



# R2 Introduction Workshop

## Advanced

26-March-2024

Lieke Hoyng , on behalf of the R2 Team

Dept. CEMM | Location AMC

Amsterdam University Medical Centers (AUMC)

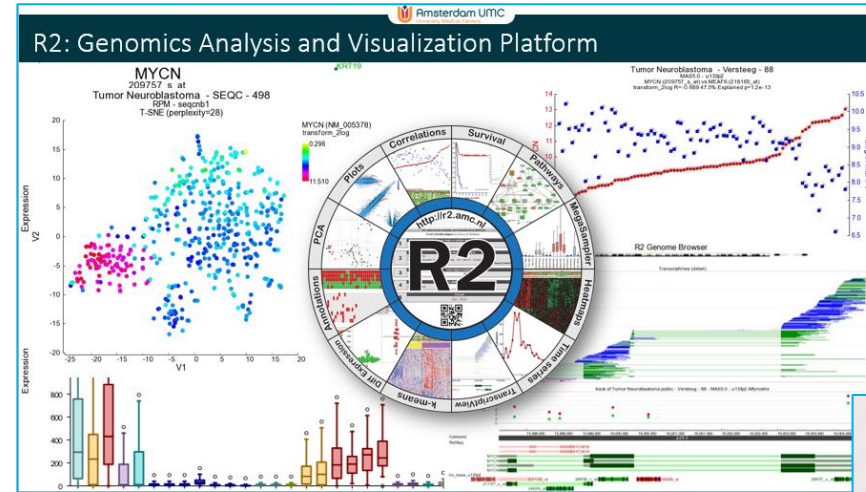
University of Amsterdam, the Netherlands

R2 Support: [r2-support@amsterdamumc.nl](mailto:r2-support@amsterdamumc.nl)

Jan Koster: [jankoster@amsterdamumc.nl](mailto:jankoster@amsterdamumc.nl)

[r2platform.com](http://r2platform.com)

# Schedule (+/-)



## This morning

9:30 - 11:00 Presentation Data / Advanced

11:00 - 11:15 Tea / coffee

11:15 - 13:00 R2 play time | Jan visit




# Genome Browser




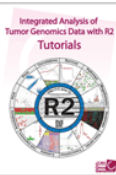
# Genome Browser

**Go to: Main**

- Main
- Time series
- AmpliconView
- Kaplan-Meier
- Sample maps
- Small Tools
- DataGrabber
- Genome Browser
- ChIP data
- TAR literature
- Change Data Scope ▶
- User Options ▶
- Help ▶
- Contact / About R2



**r2.amc.nl**

**Download the R2 Tutorials Book**

**R2: Genomics Analysis and Visualization Platform**

967627 (866122 unique) samples available

**1** Choose single or multiple dataset analysis

Single Dataset

**2** Select a dataset for analysis

Cell line Childhood Cancer (public) - Versteeg - 86 - MAS5.0 - u133p2

**3** Select type of analysis

View a Gene

**4** Proceed


**Online Tutorial**

**What is R2?**



Welcome to R2; a biologist friendly web based genomics analysis and visualization application developed by Jan Koster at the department of Oncogenomics in the Academic Medical Center (AMC) Amsterdam, the Netherlands. You can start exploring the gene expression data by following the numbered options in the center.

**For citations**, please include the following website: 'R2: Genomics Analysis and Visualization Platform (http://r2.amc.nl)'.

Financial Supporter of R2



[all news](#)

Using HGserver1(7)

Depending on how you approach the **genome browser**, different panels will become available



# Genome Browser

Go to: [Main](#) [Online Tutorial](#)

Find gene:

R2: GenomeBrowser

1 < 0.75 < 0.5 < 0.25 < << move >> 0.25 > 0.5 > 0.75 > 1 >

zoom in 20X zoom in 10X zoom in 5X zoom in 2X zoom out 2X zoom out 5X zoom out 10X zoom out 20X

hg19:chr2 R2 Genome Browser

16.077.000 16.079.000 16.081.000 16.083.000 p24.3 16.085.000 16.087.000 16.089.000

MYCNUT MYCN MYCNOS

**properties**

**TranscriptView**

Panel		Transcripts	
Intron	Exon	Intron	Exon
5'>3'	5'>3'	5'>3'	5'>3'
3'>5'	3'>5'	3'>5'	3'>5'
7?>	7?>	7?>	7?>
N.A.	N.A.	N.A.	N.A.

Draw mode:

Transcript orient.:

Type:

Min % Coverage:

Max % Coverage:

Min Alignment size:

Max Alignment size:

Min # mappings:

Max # mappings:

Best X mappings:

Min # exons:

Max # exons:

**Adjustable settings**

Genome build:

Chromosome:

Start:

End:

chr:start-end:

Vector (SVG) output:

Image Width:

Current: chr2:16075274-16092414  
Region size: 17,140 bp  
Bp per pixel: 17 bp

Cytoband click to zoom  
a10refseqDetailed, click to zoom

**Tracks**

**A: Genome Annotation**

Glensa/Cytoband

Sequence & GC windows

Conservation(PlacMammal)

CpG Islands

NAD domains Nemeth 2010

Repeats (RepeatMasker)

Sequence\_b

GC Percentage

BlackListed (Consensus)

LaminB1\_boundaries

R loop forming seq.

**B: Gene Annotation**

RefSeq(R2)

RefSeq\_features

lincRNA from Lincipedia

RefSeq(CDS)

Ensembl Gene e75

**CHIP**

ENCODE bed v1

MACS 1.4 (Public)

MACS2 (Broad) FW

MACS2 (Broad)

SuperEnhancers (ROSE)

ChIP Peaks RSEG (Public)

ENCODE bed v1 Ext

MACS2 (Narrow)

MACS2 (Broad) 2

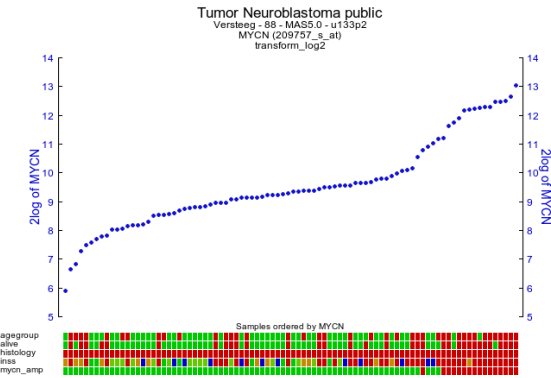
ROSE Super Enhancers (Epi Roadmap)

SuperEnhancers (ROSE AMC)

BED data

R2 has 338 tracks implemented over the different versions and access groups

# Genome Browser from basic gene expression view

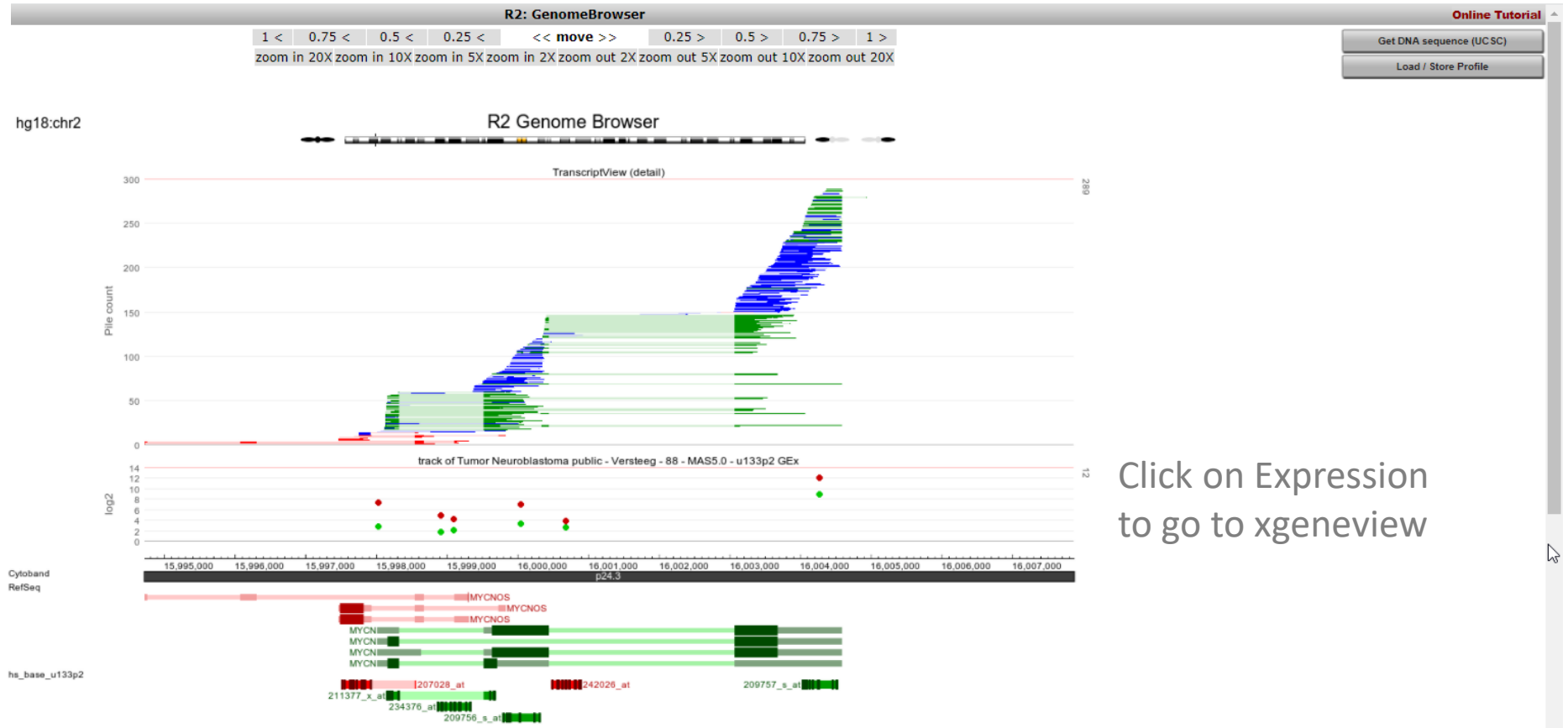


View additional details

Alternative Reporters		
Symbol Number	Reporter	
MYCN	209757_s_at (avg 1369.7 in 88 samples)	
view.all	209756_s_at (avg 235.6 in 14 samples)	
	211377_x_at (avg 158.8 in 5 samples)	
	234376_at (avg 71.7 in 3 samples)	
	242026_at (avg 22.8 in 5 samples)	

ProbesetVerification (hg18)						
symbol	probeset	rank	gene overlap	exon overlap	probes found	
MYCN	209757_s_at	GS	YES	YES	YES	
MYCN	242026_at	WS	NO	NO	NO	
MYCN	209756_s_at	3 GS	YES	YES	YES	
MYCN	211377_x_at	4 GS	YES	YES	YES	
MYCN	234376_at	5 GS	YES	YES	NO	

Probeset Genome Location (hg18):				
reporter	chrom	start	end	strand link
209757_s_at	chr2	16,004,019	16,004,525	1



Click on Expression to go to xgeneview

properties

Expression

Transformation:  ⓘ

Draw:  ⓘ

Background:  ⓘ

HugoOnce mode:  ⓘ

min # Present calls:

Slider:   ⓘ

Range:  to  ⓘ

Height:  px ⓘ

TranscriptView

Panel  Transcripts

Adjustable settings

Sample:  ⓘ

table (express): ps\_avgpres\_nbadam88\_u133p2

Select a express track:  ⓘ

Genome build:  ⓘ

Chromosome:  ⓘ

Start:  ⓘ

End:  ⓘ

chr:start-end  ⓘ

Vector (SVG) output:  ⓘ

Image Width:  ⓘ

Tracks

A: Genome Annotation

Giemsa/Cytoband  ⓘ

Sequence\_b  ⓘ

Sequence & GC windows  ⓘ

GC Percentage  ⓘ

CpG Islands  ⓘ

Fragile Sites  ⓘ

Segmental Duplications  ⓘ

Diff hg18 to hg19  ⓘ

LaminB1\_boundaries  ⓘ

NAD domains Nemeth 2010  ⓘ

Ridge boundaries  ⓘ

Repeats (RepeatMasker)  ⓘ

# Other data types than mRNA

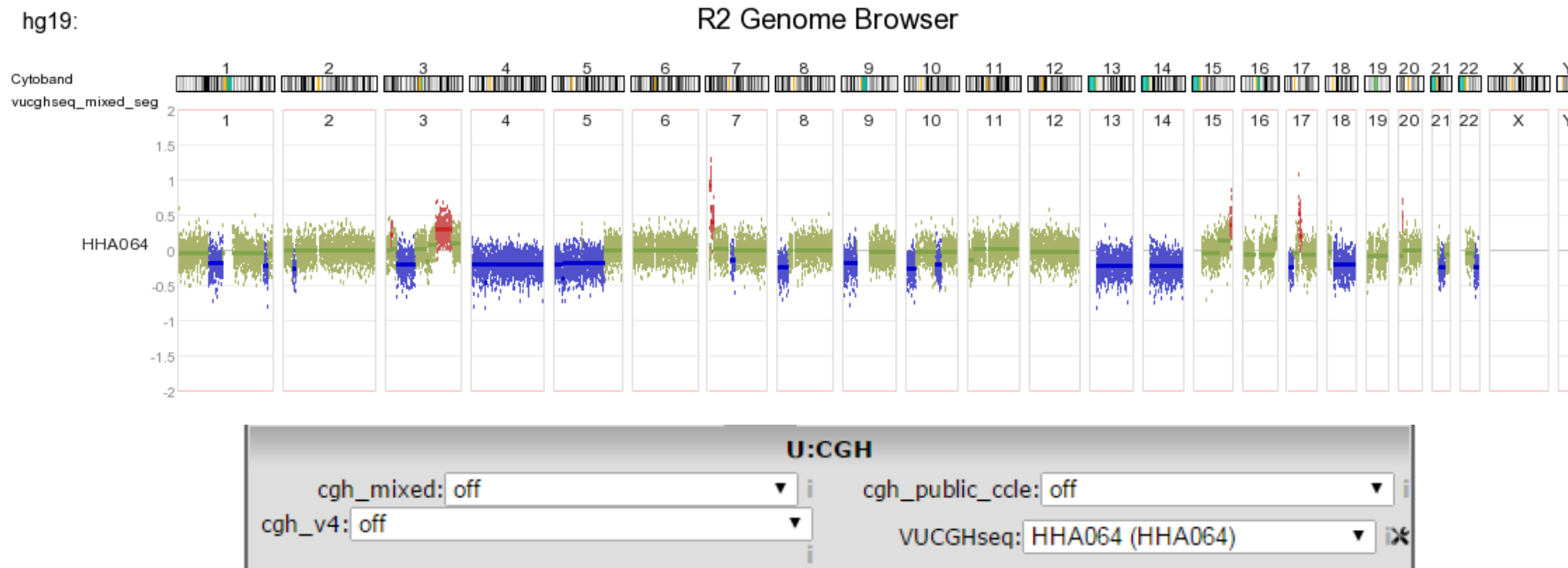




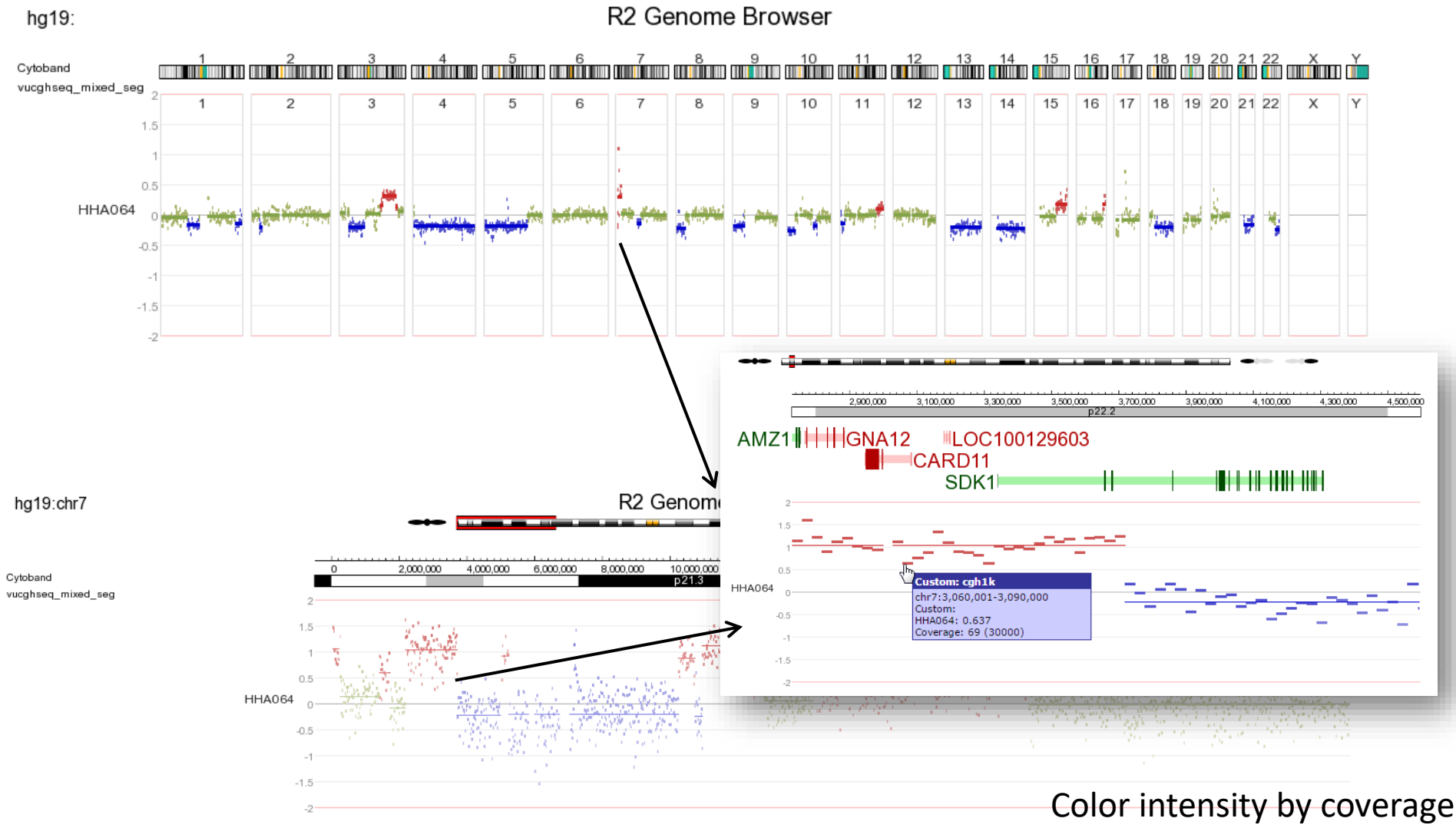
- Thus far mostly looked at mRNA
  - Similar functionalities for the other datatypes
    - aCGH / SNP / miRNA / methylation / drug response / mutation
    - Some with specific extensions/views

# aCGH / aCGH-like

- Visualization of aCGH(-like) data in the embedded genome browser of R2
  - aCGH, NGS coverage based (Whole Genome or QDNAseq)



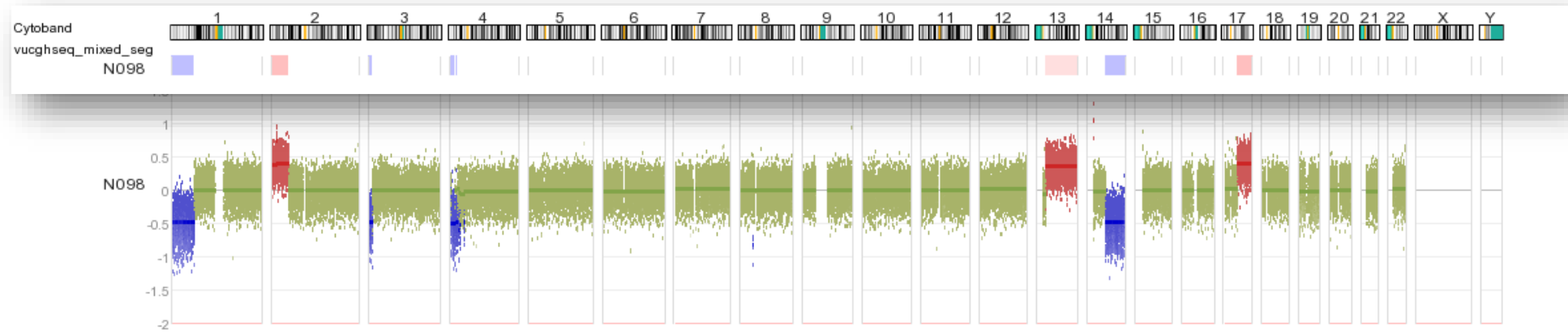
# aCGH / aCGH-like



# aCGH / aCGH-like cohort

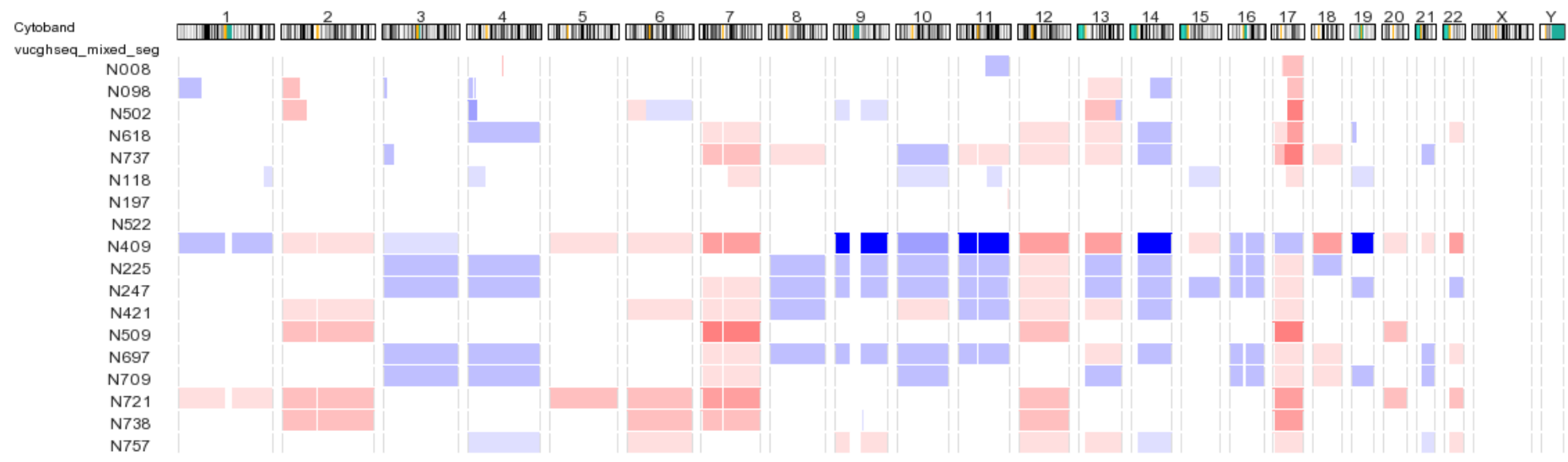
hg19:

R2 Genome Browser



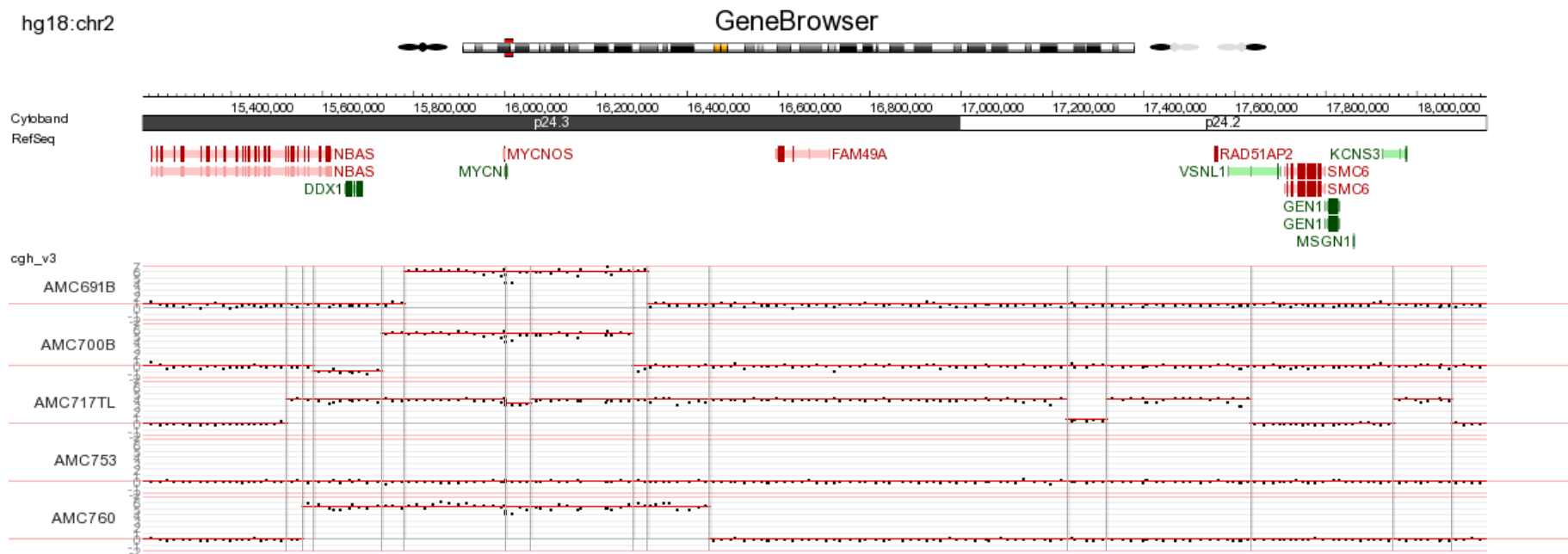
hg19:

R2 Genome Browser



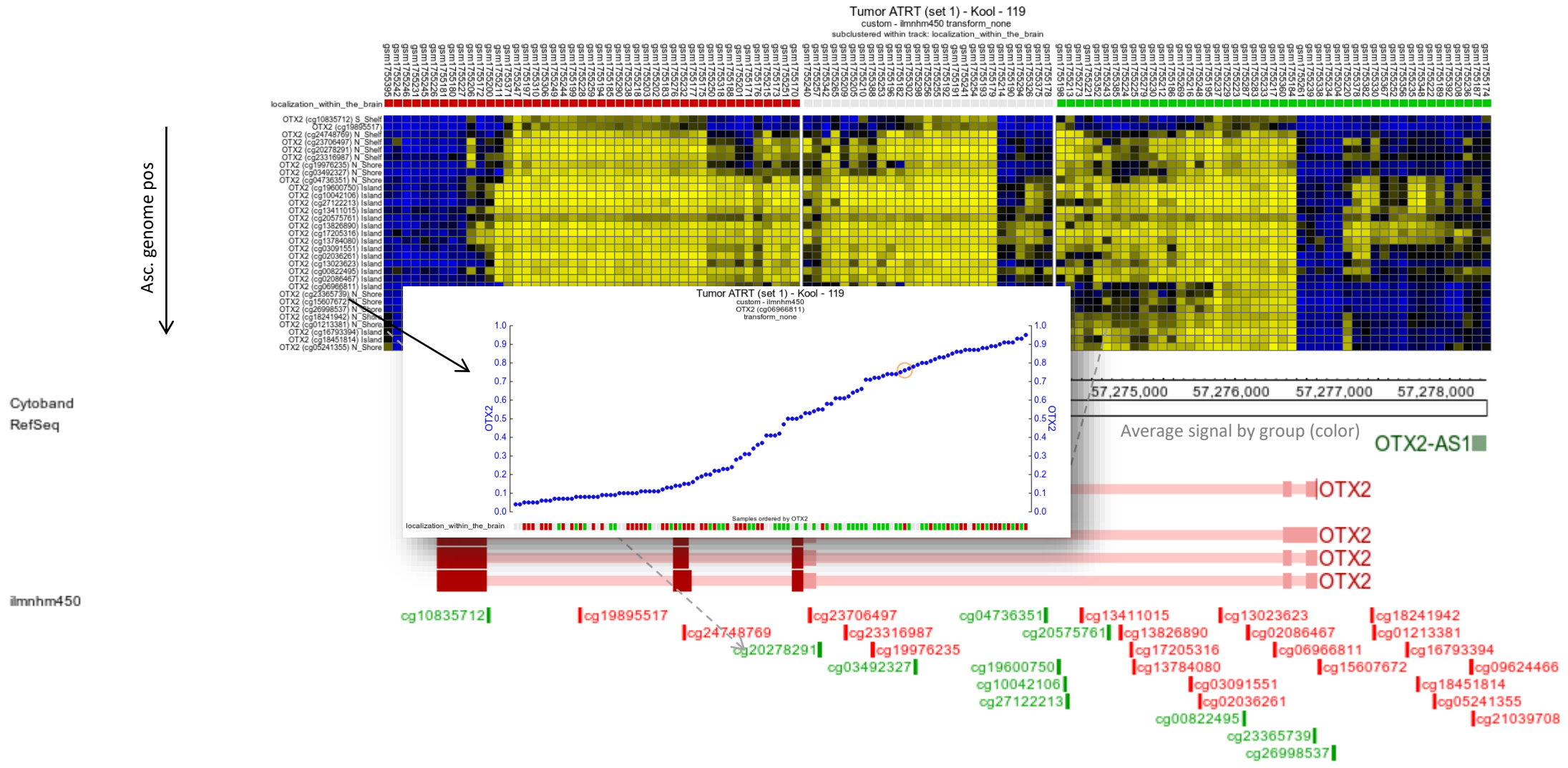
# aCGH / aCGH-like cohort dataset

- Efficient CBSbin datasets
  - Circular Binary Segmentation on raw reporter values
  - Extend segments to meet halfway
  - Superimpose (SI) all segments within a cohort
  - Annotate SI segments by overlap with gene bodies





# Methylation data



# Somatic mutation data


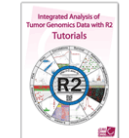
R2: Medulloblastoma 500 x R2: Variant list x +

hgserver1.amc.nl/cgi-bin/r2/main.cgi

**R2: Variant list** Online Tutorial

**Go to Main**

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- Kaplan Meier
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- Genome Browser
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**Genie:**

Genome build: hg19

Variants source: Pediatric Pan Cancer(DKFZ)

**List mode**

Detailed or summary: detailed

**Gene symbol filter**

Filter by gene name:

**Sample Filter**

Filter by sample name:

Omit samples with over: 10000 entries

**Sample Groups**

Select a track (subset): None

Selected subset: None

**Variant Filter**

Disable default filter:

Next

Default filter used: annovar\_function='exonic'.

Link	Chrom	Chromstart	Chromend	IY	RefY	AltY	Annovar_function	Gene	Exonic_classification	Dbsnp	1k_genomes	Aa_change	Add_info_r2
<a href="#">detail / view</a>	chr2	179440129	179440130	.	C	T	exonic	TTN	nonsynonymous SNV	.	.	p.D14512N,p.D146;	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr3	173997368	173997369	.	C	A	exonic	NLGN1	stopgain	.	.	p.C526X,p.C566X	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr3	38770187	38770188	.	G	A	exonic	SCN10A	nonsynonymous SNV	.	.	p.R829C	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr5	140856728	140856729	.	C	T	exonic	PCDHGC	nonsynonymous SNV	MATCH=po	.	p.A349V	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr7	25194787	25194788	.	A	G	exonic	C7orf31	nonsynonymous SNV	.	.	p.I146T	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr17	80195197	80195198	.	C	T	exonic	SLC16A3	synonymous SNV	.	.	p.G136G,p.G184G;	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr18	29432559	29432560	.	T	C	exonic	TRAPP3	nonsynonymous SNV	.	.	p.E1113G,p.E1167G;	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr19	21349176	21349177	.	C	T	exonic	ZNF431	synonymous SNV	.	.	p.L46L,p.L86L	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr1	68903929	68903930	.	A	AT	exonic	RPE65	frameshift insertion	MATCH=ine	MATCH=exact;POS=68	p.N356fs	==:FORMAT=:GT:GL:GO
<a href="#">detail / view</a>	chr1	60370620	60370621	.	T	C	exonic	CYP2J2	synonymous SNV	.	.	p.R371R	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr1	212530344	212530345	.	G	A	exonic	PPP2R5A	synonymous SNV	.	.	p.Q287Q,p.Q344Q	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr1	228645155	228645156	.	C	G	exonic	HIST3H2	synonymous SNV	MATCH=po	MATCH=position;VT=S	p.T121T	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr1	228645173	228645174	.	T	C	exonic	HIST3H2	synonymous SNV	.	.	p.V115V	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr1	228645200	228645201	.	A	G	exonic	HIST3H2	synonymous SNV	.	.	p.G106G	==:FORMAT=:GT:PL:GQ
<a href="#">detail / view</a>	chr2	9347276	9347277	.	A	T	exonic	ASAP2	nonsynonymous SNV	.	.	p.H15L	==:FORMAT=:GT:PL:GQ

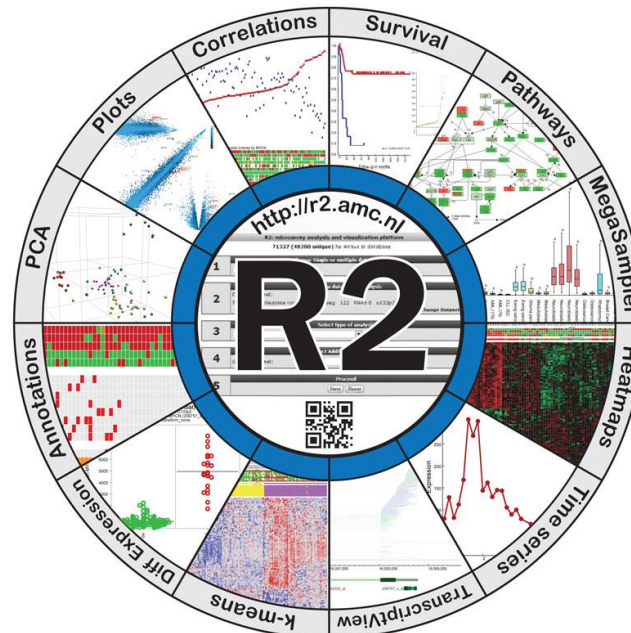
Go to page: 1 Show rows: 15 1-15 of 19188

Build a track Export to TSV

AMC: Oncogenomics Go to Main encca (2)



# Integrative Analyses



# Integrative analyses

- Multiple types of measurement on the same samples
- R2 can perform analyses on the basis of overlapping sample names in 2 different data types
  - E.g. mRNA vs miRNA
- ‘Across datasets’ option from R2 perspective
- R2 will automatically determine the intersection of overlapping samples

## Molecular data



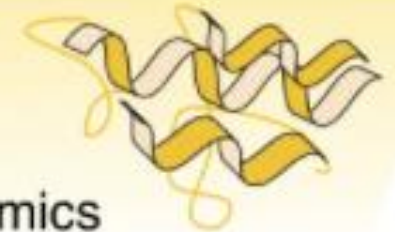
Genomics



Epigenomics



Transcriptomics



Proteomics

# Dataset Extender (2 types)

R2: Genomics Analysis and Visualization Platform

333715 (292564 unique) samples available

1 Choose single or multiple dataset analysis

2 Select an analysis

3

4 Dataset Extender (Correlate between genes)

- Dataset Extender (Correlate within genes)
- View a gene in two datatypes
- View a gene in two datasets (DataSetFuser)
- XY plot of 2 samples

**'View a gene in 2 datatypes'**  
Only look at a single combination

**'Dataset Extender'**

- Within:** only data on common genes will be analyzed.  
e.g. mRNA of MYCN vs copynumber of MYCN etc..
- Between:** all vs all  
e.g. mRNA of MYCN vs all miRNA identifiers

What is R2?  
Welcome to R2; a biologist friendly web based genomics analysis and visualization application developed by Jan Koster at the department of Oncogenomics in the Academic Medical Center (AMC) Amsterdam, the Netherlands. You can start exploring the gene expression data by following the numbered options in the center.  
For citations, please include the following website: 'R2: Genomics Analysis and Visualization Platform (http://r2.amc.nl)'.

Financial Supporter of R2

erc

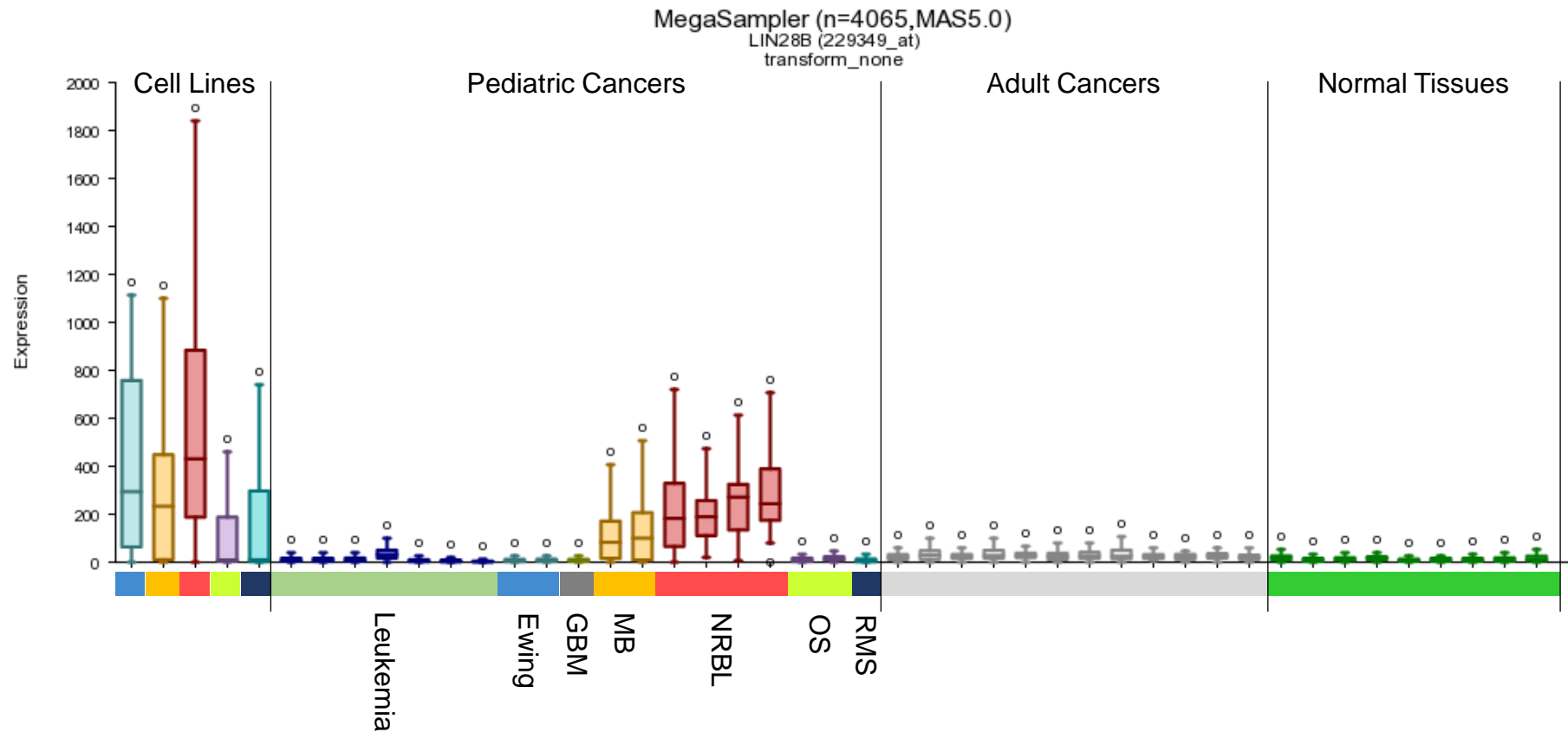
all news

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https://erc.europa.eu ics [Go to Main](#) encca (2)

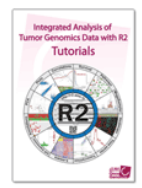
# mRNA vs miRNA

- Assess, which miRNAs are correlated with LIN28B mRNA expression



# mRNA vs miRNA

- Go to: **Main**
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  - Kaplan-Meier
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## R2: Genomics Analysis and Visualization Platform

1018043 (881501 unique) samples available

- 1** Choose single or multiple dataset analysis

Across Datasets ▼ ⓘ
- 2** Select an analysis

Select an analysis: Dataset Extender (Correlate between genes) ▼ ⓘ

  - MegaSampler (View a gene in more than 1 dataset)
  - MegaSearch (Find Differential expression between datasets)
  - 2D gene overview (View 2 genes in many datasets)
  - MegaSnitch (Associate annotations vs a gene)
  - Dataset Extender (Correlate between genes)**
  - Dataset Extender (Correlate within genes)
  - View a gene in two datatypes
  - View a gene in two datasets (DataSetFuser)
  - XY plot of 2 samples
- 3**

## Online Tutorial

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Financial Supporter of R2

all news



# mRNA vs miRNA

## R2: Dataset extender (Correlate between genes)

### Select data sets to merge

Data set collection: neuroblastoma\_amsterdam  
 Source data:  
 Target data:

- EW
- Gli
- RN
- bar
- bar
- cel
- cns
- glic
- glic
- hep
- hur
- mb
- me
- me
- me
- me
- me
- nb\_
- nb\_
- net
- net

## R2: Dataset extender (Correlate between genes)

### Select data sets to merge

Data set collection: neuroblastoma\_amsterdam  
 Source data: Expression data - Tumor Neuroblastoma (combat) - Versteeg - 122 - MAS5.0(bc) - u133p2  
 Target data: 1g

## R2: Dataset extender (Correlate between genes)

### Adjustable settings

## R2: Dataset extender (Correlate between genes)

### Adjustable settings

#### Gene Filter

Gene / Reporter: LIN28B 229349\_at advanced

#### Sample Filter

Subset track:     
 Selected sample subset: None

#### Source (ps)

Source Transform: Log2

#### Target (mir)

Target Transform: None   
 Target values: data

#### General

Corr. multiple testing: False Discovery Rate   
 p-value cutoff: 0.01

# mRNA vs miRNA

R2: hgserver2.amc.nl/cgi-bin/r2/main.cgi

Using dataset Tumor Neuroblastoma (combat) - Versteeg - 122 - MAS5.0(bc) - u133p2 vs Tumor Neuroblastic - Versteeg - 95 - custom - mir1g scanning 1 genes LIN28B (229349\_at)

22 out of 374 tests were significant at 0.01(fdr)

reporter	R	pval	reporter	R	pval
hsa-mir-18a	0.531	8.24e-06	hsa-mir-98	-0.671	5.89e-10
hsa-mir-92	0.467	2.16e-04	hsa-let-7i	-0.617	3.05e-08
hsa-mir-34c	0.476	8.80e-04	hsa-let-7d	-0.592	1.85e-07
hsa-mir-18a#	0.426	1.13e-03	hsa-let-7g	-0.580	4.08e-07
hsa-mir-15b	0.396	3.26e-03	hsa-let-7f	-0.536	6.18e-06
			hsa-mir-125b	-0.476	1.47e-04
			hsa-mir-302a	-0.458	3.24e-04
			hsa-let-7e	-0.433	8.88e-04

R2: TwoSetView

GeneID	Hugo	Description
389421	LIN28B	lin-28 homolog B (C. elegans)
406891	MIRLET71	microRNA let-7i (ALT=MIRLET7I)

Significance of correlation:  
r=-0.617 p=1.8e-09 T=-6.835 df=76

Dataset Extender Correlation  
Source: Tumor Neuroblastoma (combat) - Versteeg - 122 - MAS5.0(bc) - u133p2 (ps)  
Target: Tumor Neuroblastic - Versteeg - 95 - custom - mir1g (mir)  
LIN28B (229349\_at) vs hsa-let-7i (hsa-let-7i)

DNA amplification?

mir125B D

Let-7 transcription

LIN28B A E

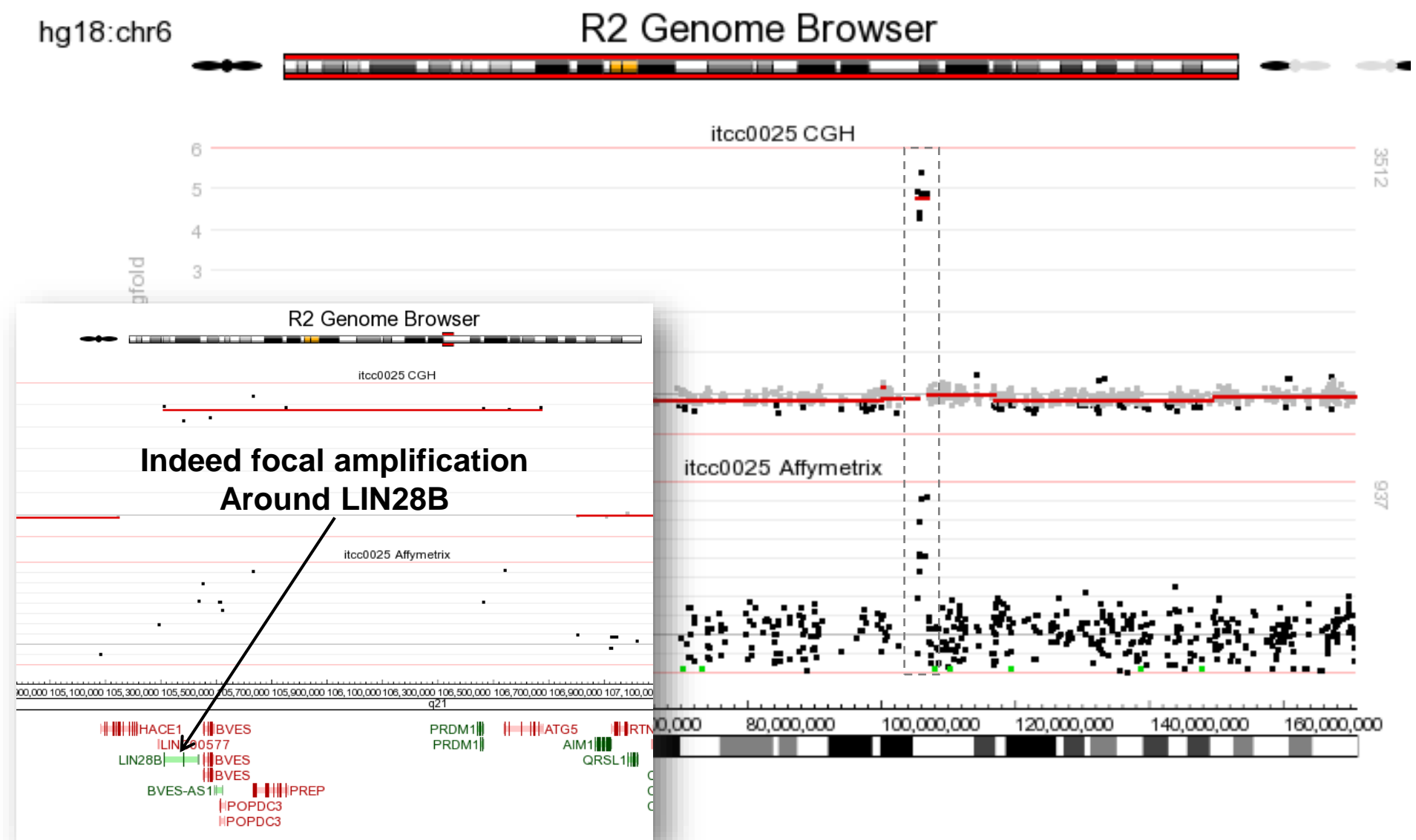
Let-7 precursors

Let-7 miRNA D

MYCN A

A Amplification  
D Deletion  
E High expression

# mRNA vs aCGH





# Best correlating pairs miRNA vs gene express

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R2: [Help\(92\)](#)
[Go to Main](#)

scanning 19843 genes  
169189 out of 8889664 tests were significant at 0.01(fdr)

reporter	R	pval	reporter	R	pval
SNORD114-3 (232355_at) vs hsa-mir-495	0.903	8.54e-29	FGF4 (206783_at) vs hsa-miR-585 (hsa-miR-585)	-0.738	9.06e-14
SNORD114-3 (232355_at) vs hsa-mir-127	0.900	3.32e-28	RAB3B (227123_at) vs hsa-mir-92 (hsa-mir-92)	-0.725	4.53e-13
SNORD114-3 (232355_at) vs hsa-mir-369-3p	0.893	3.47e-27	PHGDH (201397_at) vs hsa-mir-324-5p (hsa-mir-324-5p)	-0.721	6.56e-13
SELL (204563_at) vs hsa-mir-150	0.889	1.12e-26	MYO5A (227761_at) vs hsa-mir-92 (hsa-mir-92)	-0.719	8.61e-13
SNORD114-3 (232355_at) vs hsa-mir-655	0.889	1.27e-26	CEP152 (239413_at) vs hsa-mir-153 (hsa-mir-153)	-0.711	2.13e-12
CD3D (213539_at) vs hsa-mir-150	0.882	9.78e-26	GMPS (214431_at) vs hsa-mir-153 (hsa-mir-153)	-0.708	2.85e-12
SNORD114-3 (232355_at) vs hsa-mir-370	0.882	1.12e-25	REEP5 (208873_s_at) vs hsa-mir-92 (hsa-mir-92)	-0.707	3.41e-12
HOXC10 (218959_at) vs hsa-mir-196a	0.878	3.97e-25	ZNF496 (225335_at) vs hsa-mir-30a-5p (hsa-mir-30a-5p)	-0.706	3.54e-12
HOXC10 (218959_at) vs hsa-mir-196b	0.875	7.66e-25	RIMS1 (231986_at) vs hsa-mir-92 (hsa-mir-92)	-0.703	4.98e-12
SNORD114-3 (232355_at) vs hsa-mir-411	0.874	1.22e-24	SF3B3 (200687_s_at) vs hsa-miR-488 (hsa-miR-488)	-0.701	6.05e-12
LCK (204891_s_at) vs hsa-mir-150	0.871	2.47e-24	AHCY (200903_s_at) vs hsa-mir-328 (hsa-mir-328)	-0.699	7.51e-12
SNORD114-3 (232355_at) vs hsa-mir-379	0.868	6.02e-24	AMBP (205477_s_at) vs hsa-mir-200a# (hsa-mir-200a#)	-0.696	1.02e-11
MS4A1 (228592_at) vs hsa-mir-150	0.864	1.55e-23	BRUNOL6 (227775_at) vs hsa-mir-92 (hsa-mir-92)	-0.696	1.07e-11
SNORD114-3 (232355_at) vs hsa-mir-485-3p	0.864	1.58e-23	RNF11 (208924_at) vs hsa-mir-92 (hsa-mir-92)	-0.696	1.07e-11
CD247 (210031_at) vs hsa-mir-150	0.863	2.00e-23	TMEM97 (212282_at) vs hsa-mir-153 (hsa-mir-153)	-0.694	1.28e-11
SNORD114-3 (232355_at) vs hsa-mir-376a	0.863	2.35e-23	SNRPA (201770_at) vs hsa-miR-488 (hsa-miR-488)	-0.692	1.58e-11
SNORD114-3 (232355_at) vs hsa-mir-487b	0.862	2.79e-23	LOC728052 (1558796_a_at) vs hsa-mir-92 (hsa-mir-92)	-0.692	1.63e-11
TRBC1 (211796_s_at) vs hsa-mir-150	0.861	3.56e-23	SKP2 (203625_x_at) vs hsa-mir-153 (hsa-mir-153)	-0.691	1.73e-11
PLAC8 (219014_at) vs hsa-mir-150	0.859	5.94e-23	ZNF496 (225335_at) vs hsa-mir-148b (hsa-mir-148b)	-0.691	1.81e-11
SNORD114-3 (232355_at) vs hsa-mir-381	0.858	7.04e-23	SERPINA3 (202376_at) vs hsa-mir-200a# (hsa-mir-200a#)	-0.691	1.81e-11
SNORD114-3 (232355_at) vs hsa-mir-410	0.858	7.55e-23	RAB3B (227123_at) vs hsa-mir-19a (hsa-mir-19a)	-0.691	1.89e-11
CCR7 (206337_at) vs hsa-mir-150	0.857	1.07e-22	PPRC1 (203737_s_at) vs hsa-miR-488 (hsa-miR-488)	-0.690	1.96e-11
SNORD114-3 (232355_at) vs hsa-mir-329	0.856	1.15e-22	C16orf75 (226456_at) vs hsa-mir-204 (hsa-mir-204)	-0.688	2.54e-11
SNORD114-3 (232355_at) vs hsa-mir-369-5p	0.855	1.55e-22	NCAPG (218862_s_at) vs hsa-mir-153 (hsa-mir-153)	-0.686	3.02e-11
ICAM3 (204949_at) vs hsa-mir-142-3p	0.854	2.00e-22	SLC1A5 (208916_at) vs hsa-miR-488 (hsa-miR-488)	-0.685	3.20e-11
TRAC (209670_at) vs hsa-mir-150	0.854	2.12e-22	NOB1 (223018_at) vs hsa-miR-488 (hsa-miR-488)	-0.683	3.85e-11
RASSF5 (223322_at) vs hsa-mir-150	0.852	3.23e-22	MYBL2 (201710_at) vs hsa-mir-204 (hsa-mir-204)	-0.683	4.00e-11
IL7R (205798_at) vs hsa-mir-150	0.852	3.29e-22	E2F6 (203957_at) vs hsa-miR-585 (hsa-miR-585)	-0.682	4.52e-11
C16orf54 (1559584_a_at) vs hsa-mir-150	0.852	3.35e-22	CDC45 (224753_at) vs hsa-mir-204 (hsa-mir-204)	-0.681	4.75e-11
ICAM3 (204949_at) vs hsa-mir-150	0.851	4.28e-22	APOA2 (219466_s_at) vs hsa-mir-200a# (hsa-mir-200a#)	-0.680	5.23e-11
PLAC8 (219014_at) vs hsa-mir-142-3p	0.851	4.36e-22	MCM2 (202107_s_at) vs hsa-mir-204 (hsa-mir-204)	-0.679	5.77e-11
FAIM3 (221601_s_at) vs hsa-mir-150	0.851	4.41e-22	PPP2R2A (202313_at) vs hsa-mir-148b (hsa-mir-148b)	-0.678	6.49e-11
ICAM3 (204949_at) vs hsa-mir-142-5p	0.850	5.45e-22	LRRRC8 (228314_at) vs hsa-mir-92 (hsa-mir-92)	-0.678	6.64e-11
DENND2D (221081_s_at) vs hsa-mir-142-5p	0.849	6.39e-22	LDHA (200650_s_at) vs hsa-mir-190 (hsa-mir-190)	-0.677	6.99e-11
IKZF1 (227346_at) vs hsa-mir-150	0.849	6.58e-22	MAP1LC3A (227219_x_at) vs hsa-mir-92 (hsa-mir-92)	-0.677	7.26e-11
MEG3 (235077_at) vs hsa-mir-127	0.848	7.77e-22	TRIP13 (204033_at) vs hsa-mir-153 (hsa-mir-153)	-0.677	7.54e-11
LTB (207339_s_at) vs hsa-mir-150	0.848	7.98e-22	PSAT1 (223062_s_at) vs hsa-mir-153 (hsa-mir-153)	-0.676	7.61e-11
PTPRC (212588_at) vs hsa-mir-150	0.848	8.46e-22	TRAP1 (201391_at) vs hsa-mir-95 (hsa-mir-95)	-0.676	7.90e-11
CD2 (205831_at) vs hsa-mir-150	0.845	1.85e-21	DKK3 (214247_s_at) vs hsa-mir-20a (hsa-mir-20a)	-0.675	8.61e-11
DENND2D (221081_s_at) vs hsa-mir-150	0.844	1.96e-21	ZNF496 (225335_at) vs hsa-mir-95 (hsa-mir-95)	-0.675	9.03e-11
DENND2D (221081_s_at) vs hsa-mir-142-3p	0.844	2.20e-21	NME4 (212739_s_at) vs hsa-mir-128a (hsa-mir-128a)	-0.673	1.04e-10
SNORD114-3 (232355_at) vs hsa-mir-432	0.843	2.68e-21	DDX39 (201584_s_at) vs hsa-mir-153 (hsa-mir-153)	-0.673	1.05e-10

[Go to Main](#)
R2: TwoSetView

GeneID	Hugo	Description
767578	SNORD114-3	small nucleolar RNA, C/D box 114-3
574453	MIR495	miRNA 495 (ALT=HSA-MIR-495)

**Significance of correlation:**  
r=0.903 p=1.2e-29 T=18.350 df=76

**Dataset Extender Correlation**  
Source: Tumor Neuroblastoma - Versteeg - 88 - MASA0 - u133p2 (ps)  
Target: Tumor Neuroblastic - Versteeg - 95 - custom - mir1g (mir)  
SNORD114-3 (232355\_at) vs hsa-mir-495 (hsa-mir-495)

[Go to Main](#)
R2: TwoSetView

GeneID	Hugo	Description
5885	RAB3B	RAB3B, member RAS oncogene family

**Significance of correlation:**  
r=-0.725 p=6.4e-14 T=-9.168 df=76

**Dataset Extender Correlation**  
Source: Tumor Neuroblastoma - Versteeg - 88 - MASA0 - u133p2 (ps)  
Target: Tumor Neuroblastic - Versteeg - 95 - custom - mir1g (mir)  
RAB3B (227123\_at) vs hsa-mir-92 (hsa-mir-92)

Min source	Count
PAICS	11
ZNF496	10
SF3B3	9
NAT10	9
SLC1A5	8
BAZ1A	8
GMPS	8
ALG3	7
RAB3B	7
PHGDH	7
PPRC1	7
MARVELD1	7
KIAA0020	7
SCFD2	7
TRAP1	7
SNRPA	6
PRDX6	6
EXO1	6
DKK3	6
C19orf48	6
RUVBL1	6
POLR3D	6
SLC38A5	5
PUS7	5
WDR3	5
C4orf14	5
TMEM97	5
CCPG1	5
TAF1D	5
NOP14	4
SERINC1	4
HPDL	4
FBL	4
GABARAPL1	4
TNFRSF10B	4
MTR	4
POLD1	4
LIN28B	4
TKT	4
SLC16A1	4
ATIC	4
CEP152	4

# Relate Gene Expression to drug sensitivity

**Select data sets to merge**

Data set collection:

Source data:

Target data:

**Adjustable settings**

**Gene Filter**

Chromosome:  ⓘ

Gene ontology:

Gene set:

Manual list:  ⓘ

**Sample Filter**

Subset track:  ⚙️ ⓘ

Selected sample subset: None

**Source (ps)**

Source Transform:  ⓘ

**Target (drug)**

Target Transform:  ⓘ

Target values:

**General**

Corr. multiple testing:  ⓘ

p-value cutoff:

# Drug (IC50) data

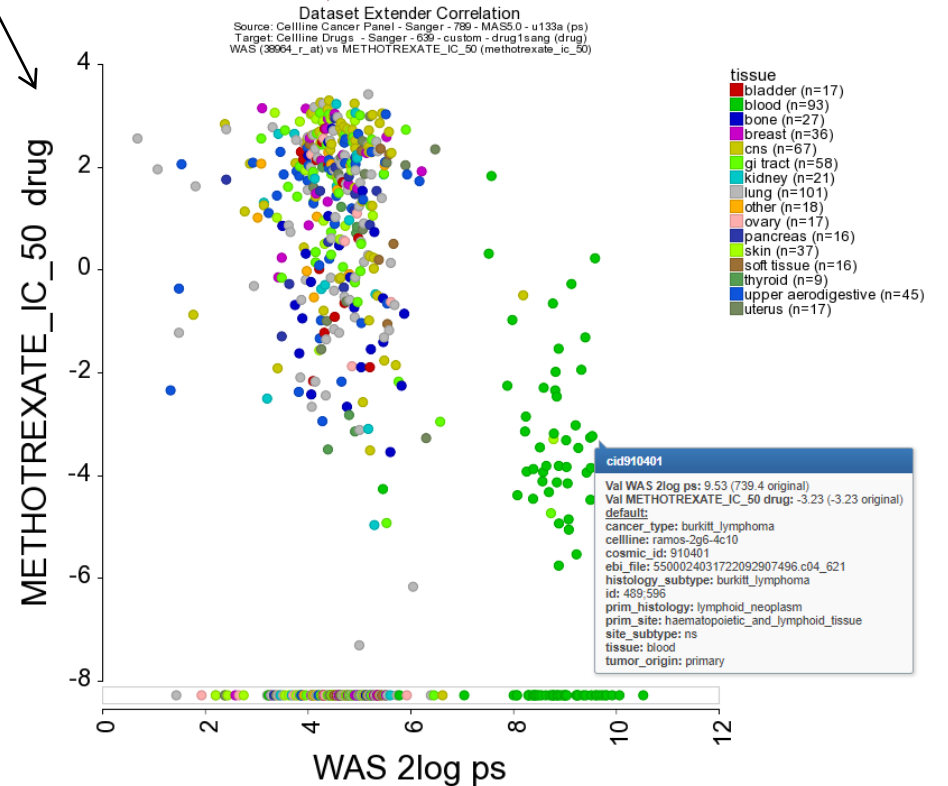
R2:

Using dataset Cellline Cancer Panel - Sanger - 789 - MASS.0 - u133a vs Cellline Drugs - Sanger - 639 - custom - drug1sang  
scanning 12503 genes

47026 out of 1637893 tests were significant at 0.01(fdr)

reporter	R	pval	reporter	R	pval
SH3BP4 (222258 s at) vs methotrexate ic 50	0.573	1.48e-30	WAS (38964 r at) vs methotrexate ic 50	-0.565	1.21e-29
GNG12 (212294 at) vs methotrexate ic 50	0.559	5.13e-29	LRMP (204674 at) vs methotrexate ic 50	-0.540	8.81e-27
TJP1 (202011 at) vs methotrexate ic 50	0.555	1.54e-28	GMFG (204220 at) vs methotrexate ic 50	-0.540	9.79e-27
FAM114A1 (213455 at) vs methotrexate ic 50	0.538	1.71e-26	NQO1 (201468 s at) vs 17-aag ic 50	-0.533	1.04e-26
AMOTL2 (203002 at) vs methotrexate ic 50	0.531	1.06e-25	NCKAP1L (209734 at) vs methotrexate ic 50	-0.533	6.97e-26
PPIC (204517 at) vs methotrexate ic 50	0.529	1.35e-25	ACAP1 (205213 at) vs methotrexate ic 50	-0.532	8.55e-26
EKBP9 (212169 at) vs methotrexate ic 50	0.530	1.38e-25	MFNG (204153 s at) vs methotrexate ic 50	-0.529	1.32e-25
ITGB5 (201125 s at) vs methotrexate ic 50	0.528	1.53e-25	DOCK2 (213160 at) vs methotrexate ic 50	-0.507	4.68e-23
NCKAP1 (207738 s at) vs methotrexate ic 50	0.515	6.70e-24	CD53 (203416 at) vs methotrexate ic 50	-0.506	5.91e-23
DAG1 (205417 s at) vs methotrexate ic 50	0.512	1.77e-23	CD48 (204118 at) vs methotrexate ic 50	-0.505	8.40e-23
CTTN (201059 at) vs methotrexate ic 50	0.508	4.03e-23	IKZF1 (205039 s at) vs methotrexate ic 50	-0.502	1.89e-22
PLS3 (201215 at) vs methotrexate ic 50	0.508	4.30e-23	MAP4K1 (206296 x at) vs methotrexate ic 50	-0.500	1.89e-22
IL13RA1 (201887 at) vs methotrexate ic 50	0.508	4.56e-23	TRAF3IP3 (213888 s at) vs methotrexate ic 50	-0.499	1.89e-22
YAP1 (213342 at) vs methotrexate ic 50	0.500	3.14e-22	NCF4 (207677 s at) vs methotrexate ic 50	-0.498	1.89e-22
PON2 (201876 at) vs methotrexate ic 50	0.497	6.99e-22	SASH3 (204923 at) vs methotrexate ic 50	-0.498	1.89e-22
TEAD1 (214600 at) vs methotrexate ic 50	0.492	1.98e-21	HCLS1 (202957 at) vs methotrexate ic 50	-0.498	1.89e-22
CAPN2 (208683 at) vs methotrexate ic 50	0.491	2.89e-21	PHLDA1 (217996 at) vs rdea119 ic 50	-0.498	1.89e-22
PARVA (217890 s at) vs methotrexate ic 50	0.490	3.24e-21	PTPN7 (204852 s at) vs methotrexate ic 50	-0.498	1.89e-22
NQO1 (201468 s at) vs methotrexate ic 50	0.486	9.33e-21	ZMAT3 (219628 at) vs nutlin-3a ic 50	-0.498	1.89e-22
SNX7 (205573 s at) vs methotrexate ic 50	0.484	1.60e-20	FLI1 (204236 at) vs methotrexate ic 50	-0.498	1.89e-22
SDC4 (202071 at) vs methotrexate ic 50	0.482	2.42e-20	CORO1A (209083 at) vs abt-263 ic 50	-0.498	1.89e-22
LAMB2 (216264 s at) vs methotrexate ic 50	0.480	3.60e-20	PTPRC (212587 s at) vs methotrexate ic 50	-0.498	1.89e-22
LAPTM4A (200673 at) vs methotrexate ic 50	0.480	3.62e-20	CORO1A (209083 at) vs methotrexate ic 50	-0.498	1.89e-22
ITGAV (202351 at) vs methotrexate ic 50	0.480	3.65e-20	DCK (203302 at) vs methotrexate ic 50	-0.498	1.89e-22
CEP170B (213242 x at) vs methotrexate ic 50	0.480	3.67e-20	ARHGAP15 (218870 at) vs methotrexate ic 50	-0.498	1.89e-22
TSPAN6 (209108 at) vs methotrexate ic 50	0.478	5.20e-20	RHOH (204951 at) vs methotrexate ic 50	-0.498	1.89e-22
TNFRSF12A (218368 s at) vs methotrexate ic 50	0.476	8.27e-20	LAPTM5 (201721 s at) vs methotrexate ic 50	-0.498	1.89e-22

Min source	Count
PTRF	7
MYOF	7
FLI1	6
ANXA1	6
GMFG	5
WWTR1	5
HNRNPD	5
NCKAP1L	5
PRKCB	5
MBP	5
ARID1A	5



# Correlate within genes (Gex vs Copynumber)

hgserver1.amc.nl/cgi-bin/r2/main.cgi

**R2: Genomics Analysis and Visualization Platform** Online Tutorial

1018043 (881501 unique) samples available

**1** Choose single or multiple dataset analysis  
Across Datasets

**2** Select an analysis  
Select an analysis: Dataset Extender (Correlate within genes)

**3** Proceed  
Next Reset

---

**R2: Dataset extender (Correlate within genes)**

Select data sets to merge

Data set collection: neuroblastoma\_amsterdam

Source data: Expression data - Tumor Neuroblastoma (combat)

Target data: aCGH data - Uncategorized - 110 - custom - gent

Select data sets

**Adjustable settings**

**Sample Filter**

Subset track:

Selected sample subset: None

**Source (ps)**

Source Transform: Log2

HugoOnce mode: yes

min # Present calls: 1

**Target (cgh)**

Target Transform: None

Target values: logfolds

**General**

Corr. multiple testing: False Discovery Rate

r\_pval\_cutoff: 0.01


Next Reset



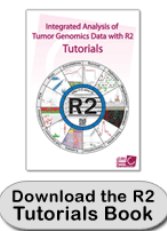
**What is R2?**

Welcome to R2; a biologist friendly web based genomics analysis and visualization application developed by Jan Koster at the department of Oncogenomics in the Academic Medical Center (AMC) Amsterdam, the Netherlands. You can start exploring the gene expression data by following the numbered options in the center.

**For citations,** please include the following website: 'R2: Genomics Analysis and Visualization Platform (http://r2.amc.nl)'.

**Financial Supporter of R2**



# Correlate within genes (Gex vs Copynumber)

Go to: [Main](#)

R2: datafusion\_gene\_search

[Online Tutorial](#)

hg18: R2 Genome Browser

hg18: R2 Genome Browser

hg18:chr2 R2 Genome Browser

hg18:chr2 R2 Genome Browser

Within Gene Cross Correlation Result

Within Gene Cross Correlation Result

Within Gene Cross Correlation Result

Within Gene Cross Correlation Result

properties

panel chrmap0

Range: a to a

Height: 150 px

Adjustable set

Genome build: HG18

Chromosome: chr2

Start: 15390187

End: 38310180

chr:start-end

Vector (SVG) output: false

ID

chr2:17,753,568-17,753,568

Reporter:

-log10 raw pvalue: 23.0754130891036

GeneSymbol: SMC6

genesymbol: SMC6

source\_id: 218781\_at

target\_id: A\_16\_P00324017

correlation: 0.835343062082847

pvalue: 8.40595209959222e-24

Dataset Extender Correlation

Source: Tumor Neuroblastoma (kombal) - Versteeg - 122 - MA25.0(b) - u133p2 (ps)

Target: Uncatalogued - 110 - custom - igent (cpg)

SMC6 (218781\_at) vs SMC6 (A\_16\_P00324017)

SMC6 cgh

SMC6 log2 ps

ATG5 chr6	0.771	9.64e-16
CAMTA1 chr1	0.756	8.59e-15
BSDC1 chr1	0.749	2.61e-14
TSFM chr12	0.748	2.73e-14
RER1 chr1	0.742	6.45e-14
FOXJ3 chr1	0.739	8.66e-14
SRRM1 chr1	0.739	8.80e-14
IGHMBP2 chr11	0.739	8.88e-14
SPCS2 chr11	0.738	0.64e-14

# View all reporters for a gene (module)

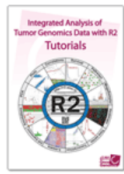
The screenshot shows the R2 Genomics Analysis and Visualization Platform interface. The browser address bar displays `https://hgserver2.amc.nl/cgi-bin/r2/main.cgi`. The main header indicates the platform name and the number of samples available: "R2: Genomics Analysis and Visualization Platform" and "348140 (198492 unique) hs samples available".

The interface is divided into several sections:

- Navigation Menu (Left):** A sidebar menu with options like Main, Time series, AmpliconView, Kaplan Meier, Tools, t-SNE Maps, Genome Browser, ChIP data, Change Data Scope, User Options, Help, and Contact / About R2.
- Workflow Steps (Center):** A series of five numbered steps:
  - 1 Choose Single or multiple dataset analysis:** A dropdown menu set to "Single Dataset".
  - 2 Select a dataset for analysis:** Shows the current dataset as "Tumor Medulloblastoma (DKFZ) - Kool - 363 - custom - ilmnhm450" with a "Change Dataset | Advanced Selection" link.
  - 3 Select type of analysis:** A dropdown menu set to "View all reporters for a gene (Heatmap)".
  - 4 Select Additional Conditions:** A text input field for "Gene:" containing "otx2".
  - 5 Proceed:** "Next" and "Reset" buttons.
- Quote (Bottom Center):** A green box containing the text: "quote The important thing is to never stop questioning. (Albert Einstein) Quote".
- Right Sidebar:**
  - What is R2?:** A text box providing a welcome message and citation information.
  - Financial Supporter of R2:** A section featuring the "erc" logo.
  - all news:** A link to news updates.
  - Social Media:** Facebook and Twitter icons.
  - Usage Statistics:** "Using: HGserver2 7,894,111 hits since 2008-04-07".
- Bottom Left:** A "Download the R2 Tutorials Book" button with a "PubMed" logo and a stack of books.

Go to: Main

- Main
- ITCC-P4 (IMIZ) ▶
- Time series
- Survival (Kaplan-Meier/Cox)
- Sample maps (UMAP/tSNE)
- Small Tools
- DataGrabber
- Genome Browser
- ChIP data
- TAR literature
- Change Data Scope ▶
- User Options ▶
- Administrator ▶
- Help ▶
- Contact / About R2



Download the R2 Tutorial Book

R2: Genomics Analysis and Visualization Platform

5,019,382 (3,775,101 unique) samples available

**1** Choose single or multiple dataset analysis

Single Dataset ▾ ⓘ

---

**2** Select a dataset for analysis

Tumor Medulloblastoma (DKFZ) - Kool - 363 - custom - ilnmhm450 ▾

---

**3** Select type of analysis

View all Met. ids for a Gene (Heatmap) ▾ ⓘ

---

**4** Proceed

Online Tutorial

What is R2?

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**For citations**, please include the following website: 'R2: Genomics Analysis and Visualization Platform (<http://r2.amc.nl>)'.

Financial Supporter of R2

Cancer Center Amsterdam



News

Try our updated vector graphs in the platform. We love to hear what you think about them.

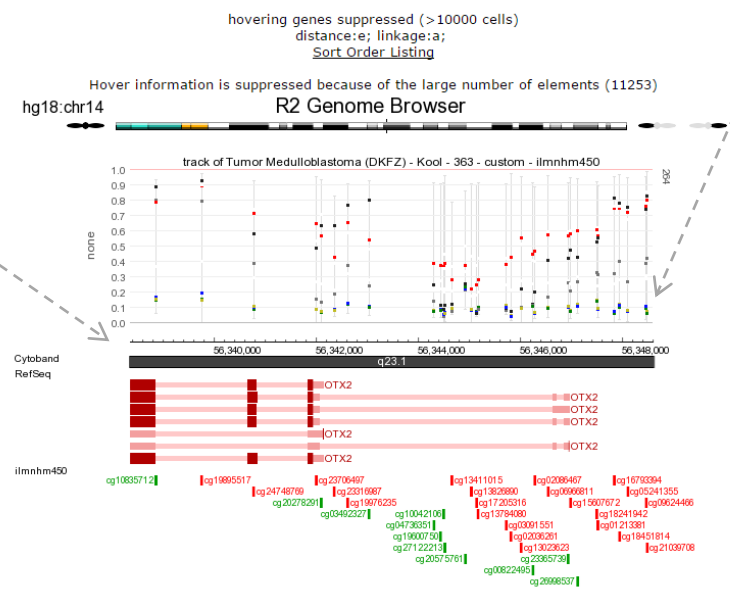
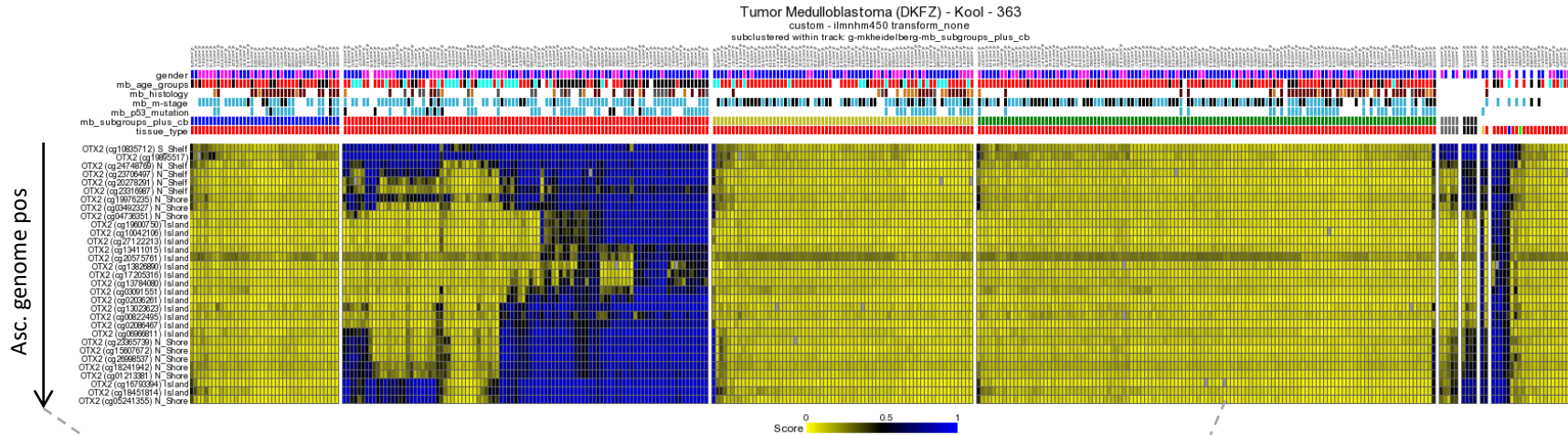
News

Check out the new ITHIR pediatric cancer precision medicine datascope. Now publicly available in R2 via the 'datascope' menu item.

[all news](#)



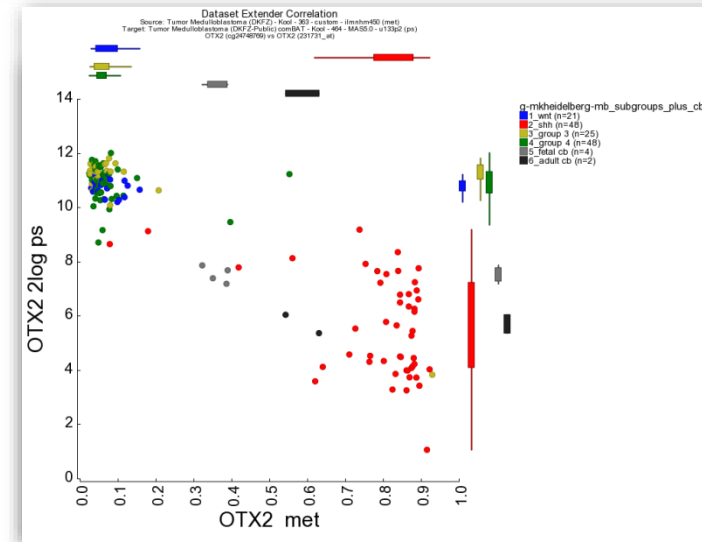
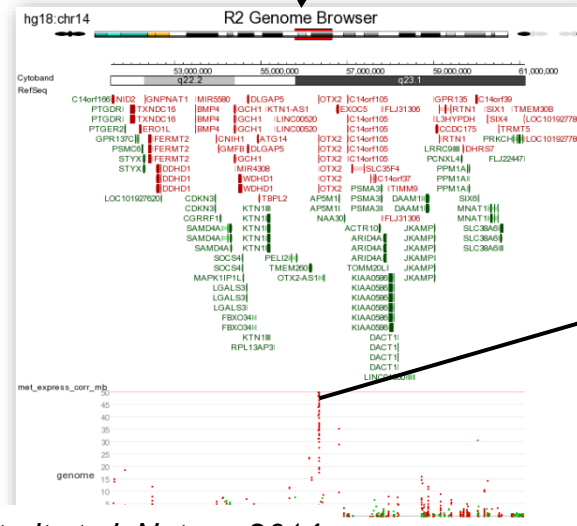
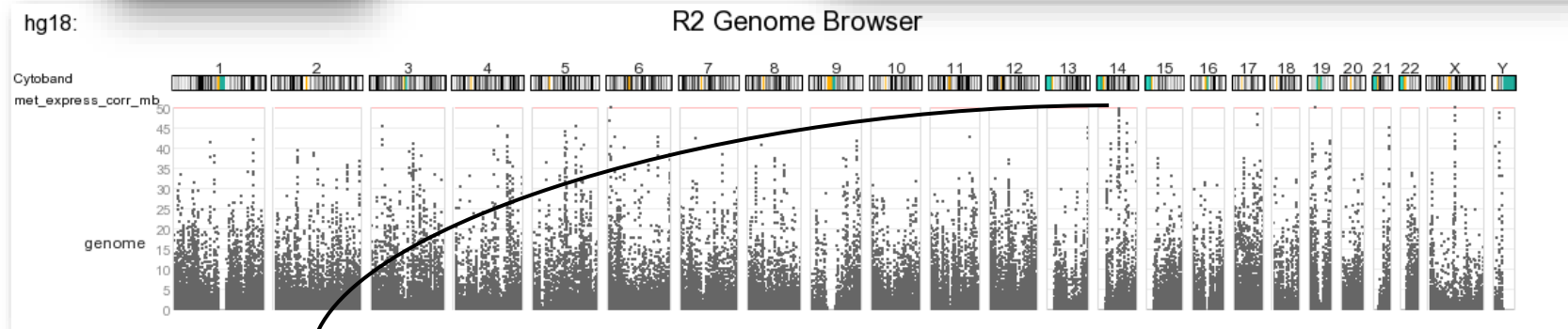
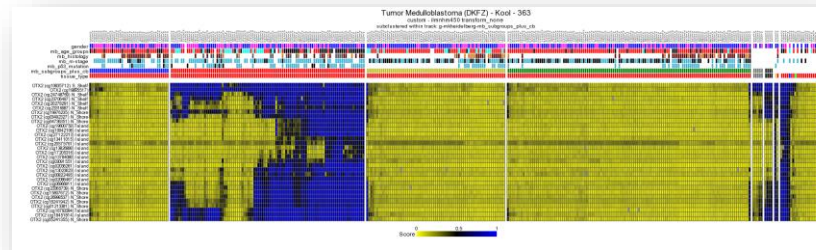
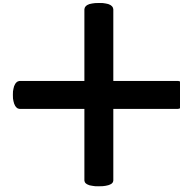
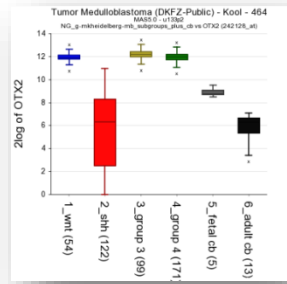
# Methylation arrays



Average signal by group (color)

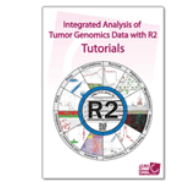


# Integration Methylation vs Gene Expression



# Whole genome sequencing data in R2

- Go to: **Main**
- Main
  - Time series
  - AmpliconView
  - Kaplan-Meier
  - Sample maps
  - Small Tools
  - DataGrabber
  - Genome Browser
  - ChIP data
  - TAR literature
  - Change Data Scope >
  - User Options >
  - Help >
  - Contact / About R2



Download the R2 Tutorials Book

**R2: Genomics Analysis and Visualization Platform**

967627 (866122 unique) samples available

- 1** Choose single or multiple dataset analysis  
Single Dataset
- 2** Select a dataset for analysis  
Tumor Neuroblastoma public - Versteeg - 88 - MAS5.0 - u133p2
- 3** Select type of analysis  
View a Gene
- 4** *(Dropdown menu open)*

- TopLister (Gene filter stdev)
- Geneset vs Genesets Correlations
- Track(#) vs Genesets Correlations
- Venn Diagram of GeneCategories
- Personalized Genomics**
- Static circos files (v3)
- Somatic Mutations (v3)
- Pers. Med. OncoPrint
- CliniSnitch (Track vs som. mutation)
- Find a Sample on omics criteria
- Plot CGH karyograms in track assisted order
- Other analyses**
- Genome Browser
- Affy-CGH-SNP (combined) Browser
- Affy Genome profiler
- ChIP Genome Browser
- ChIP data
- CliniSnitch
- CliniSnitch (by track)
- I feel lucky

**Online Tutorial**

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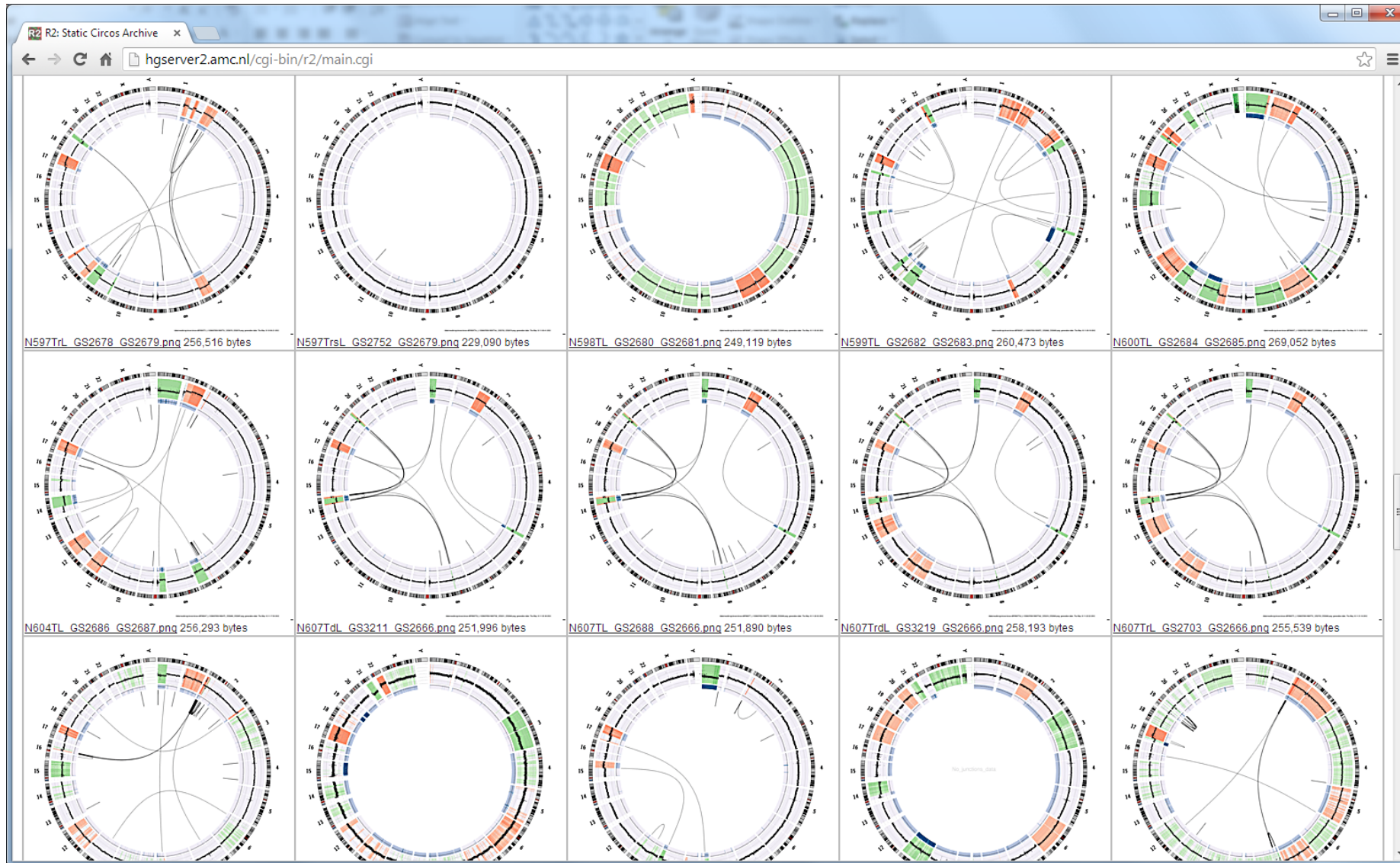
Financial Supporter of R2

Amsterdam UMC

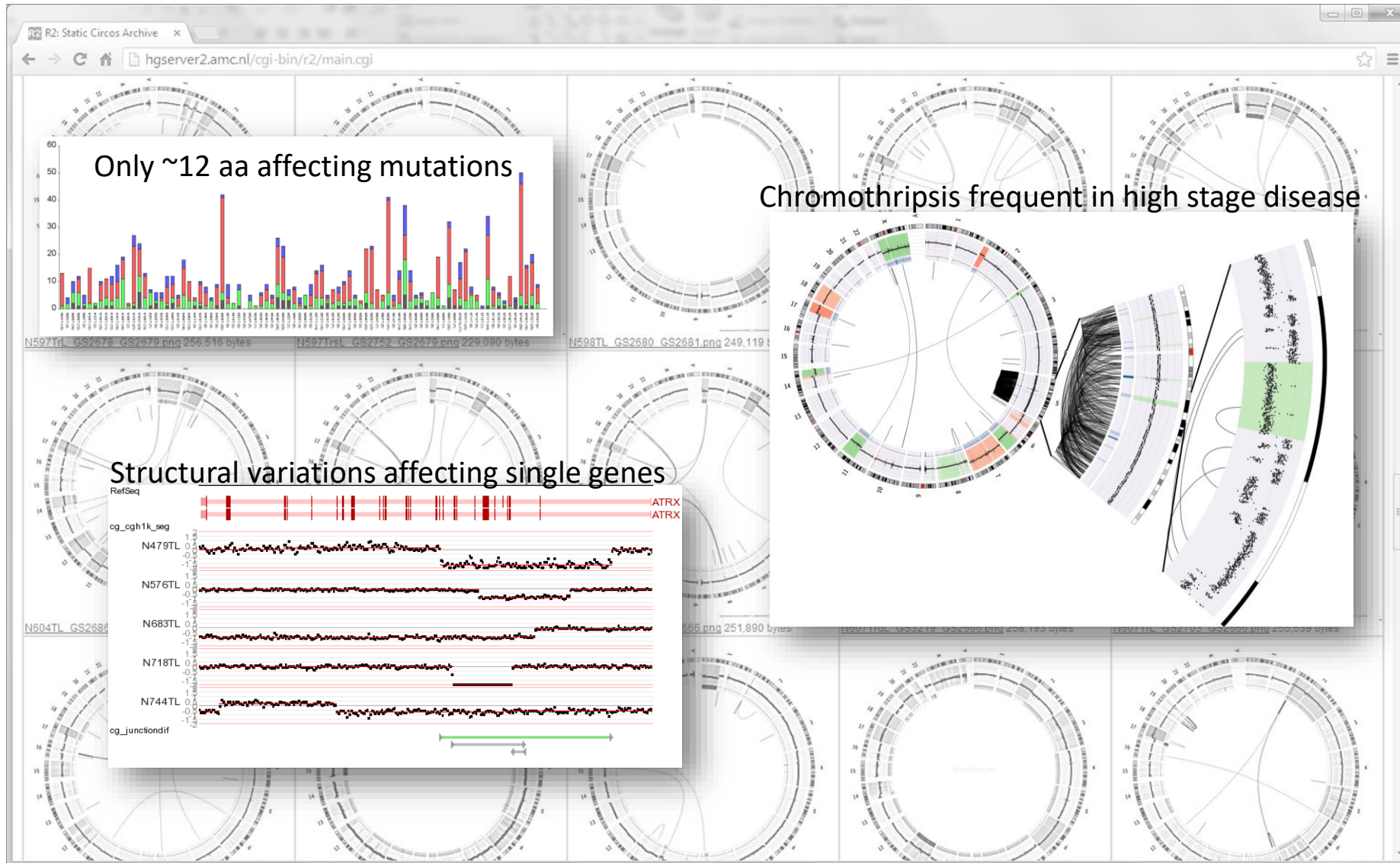
[all news](#)



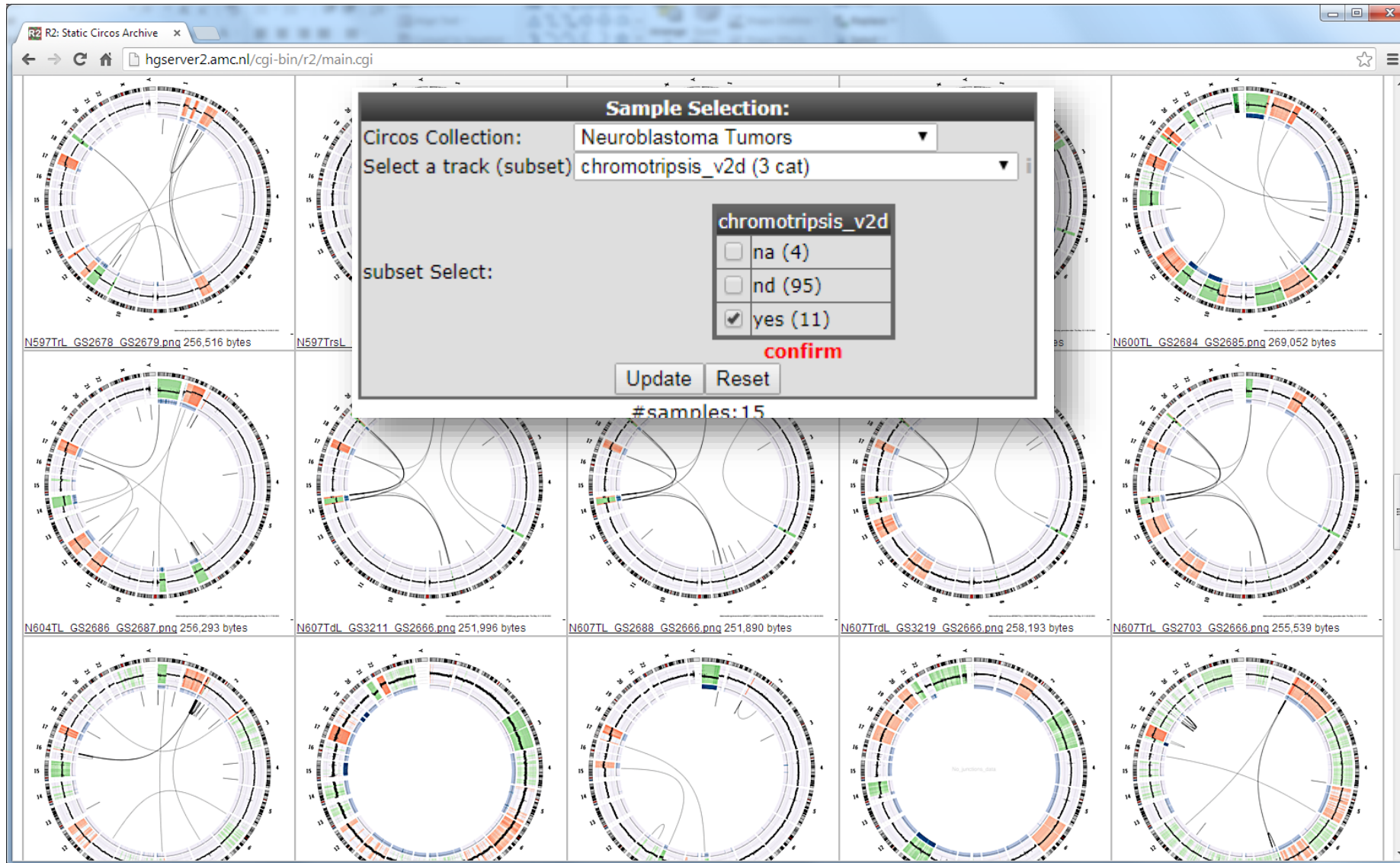
# Whole genome sequencing data in R2



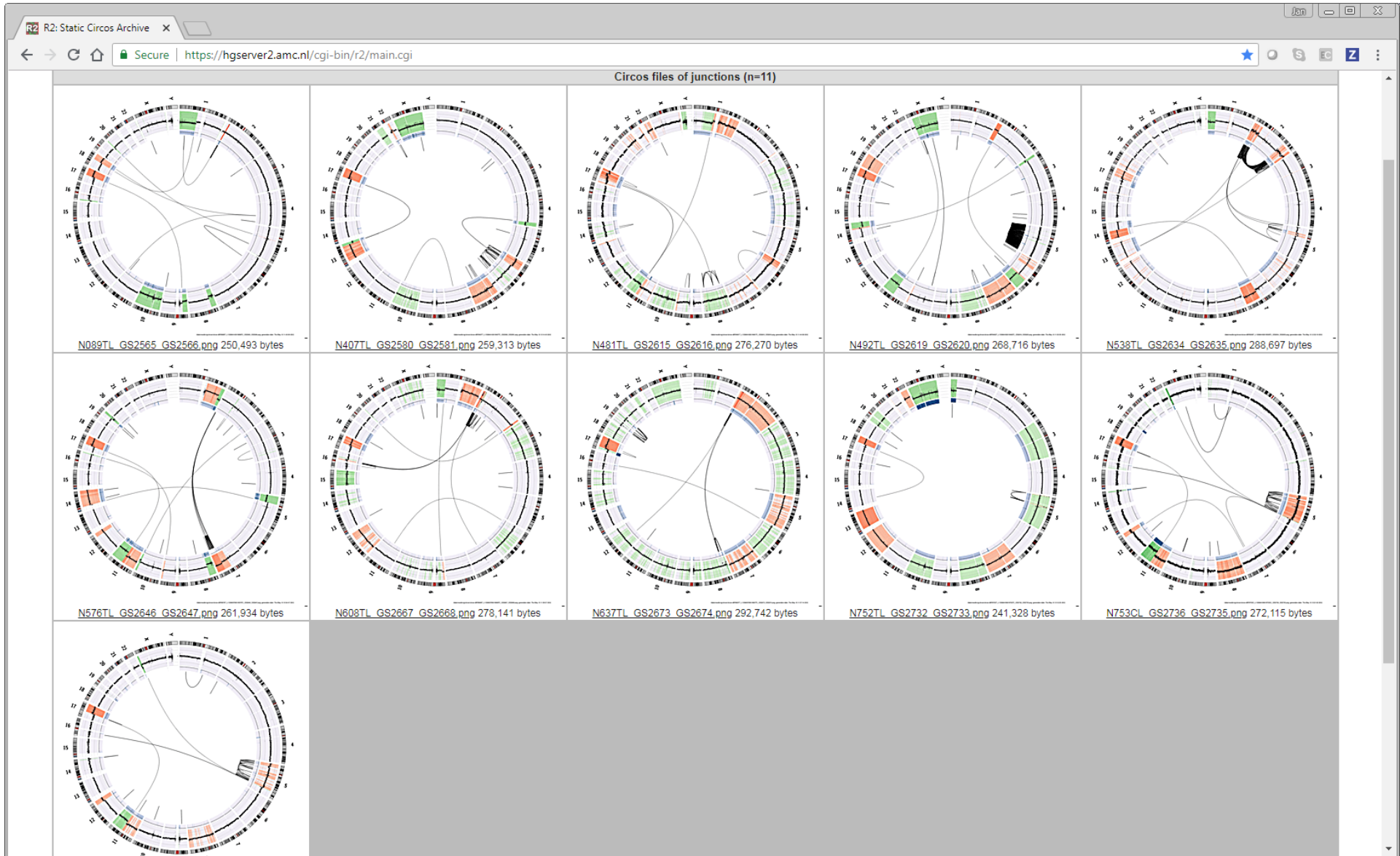
# Whole genome sequencing data in R2



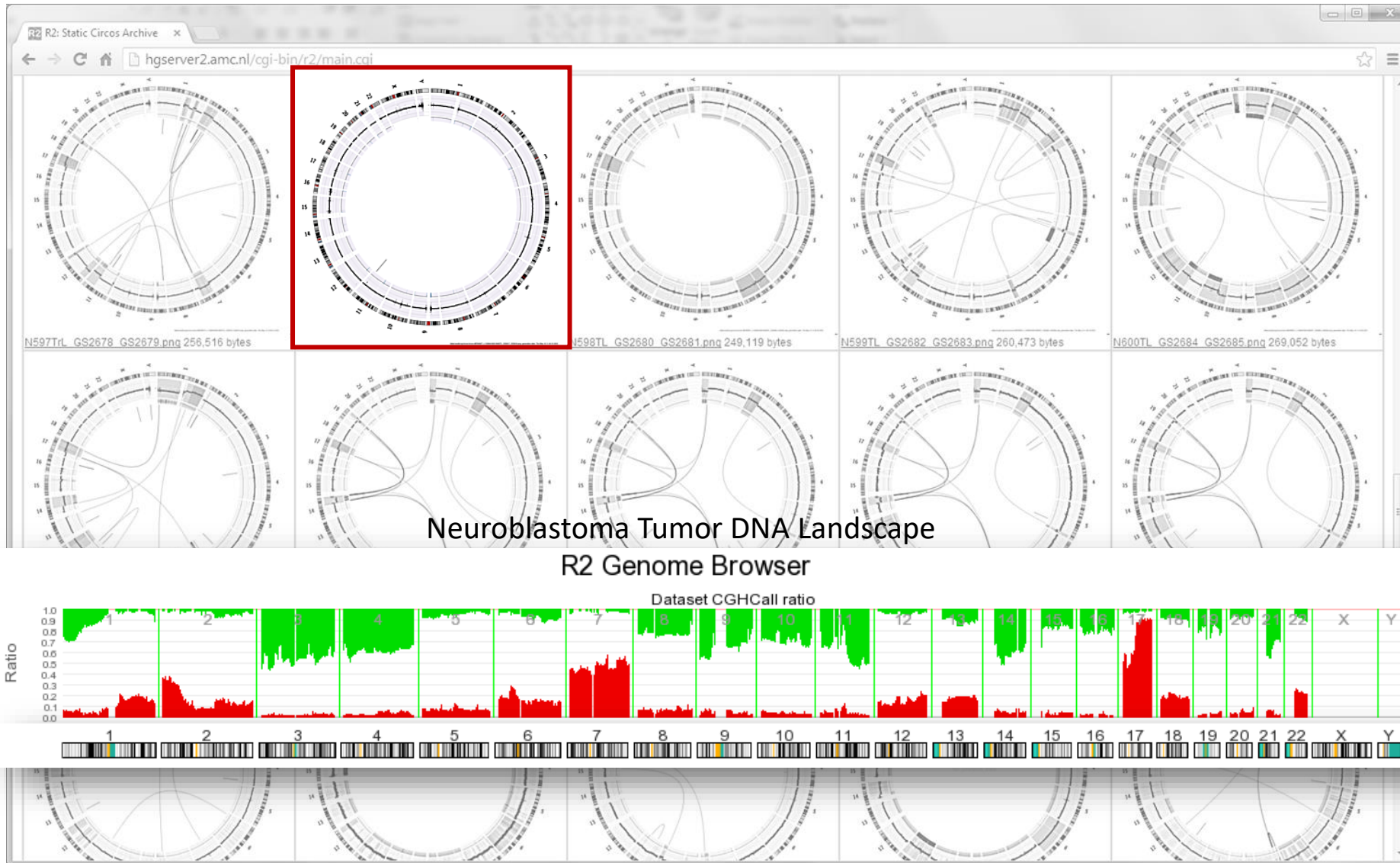
# Whole genome sequencing data in R2



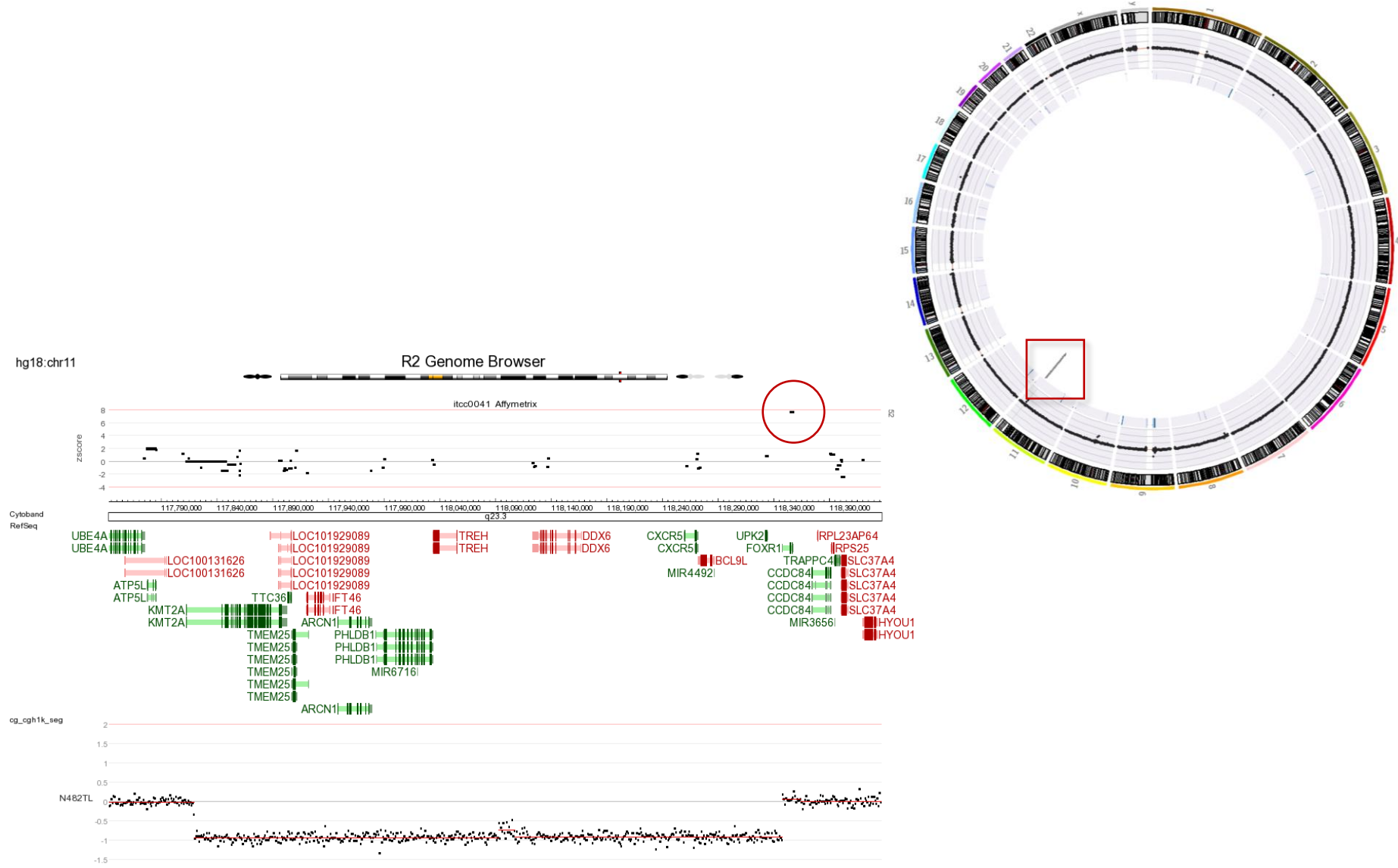
# Whole genome sequencing data in R2



# Whole genome sequencing data in R2

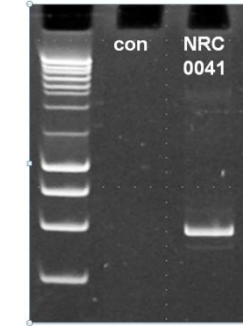
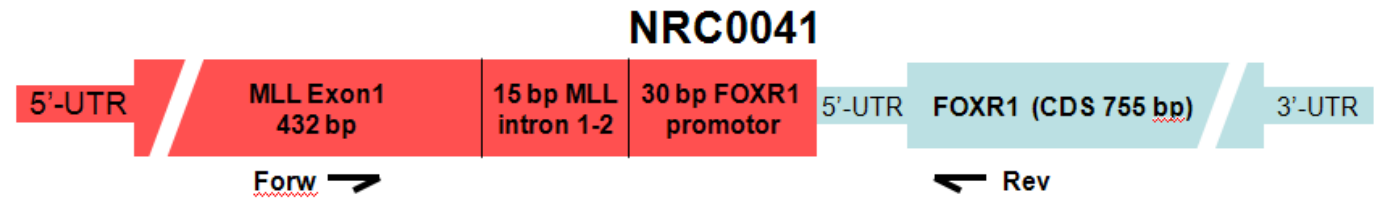
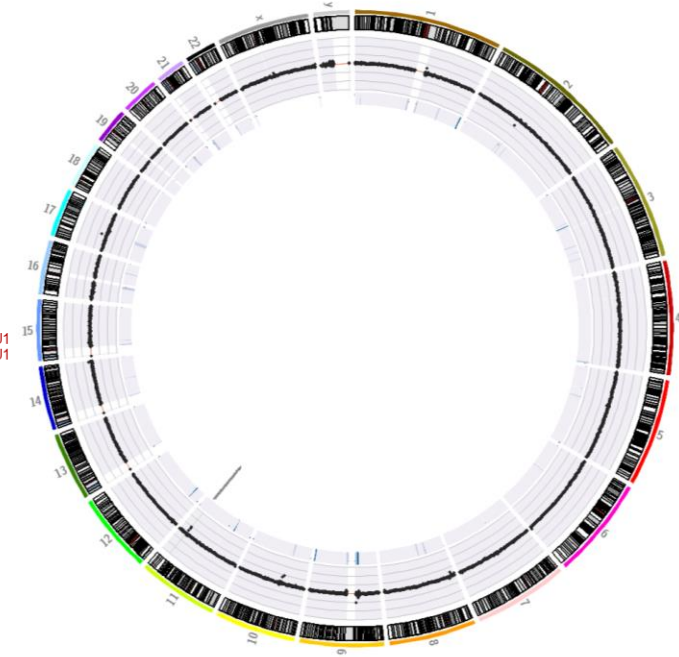
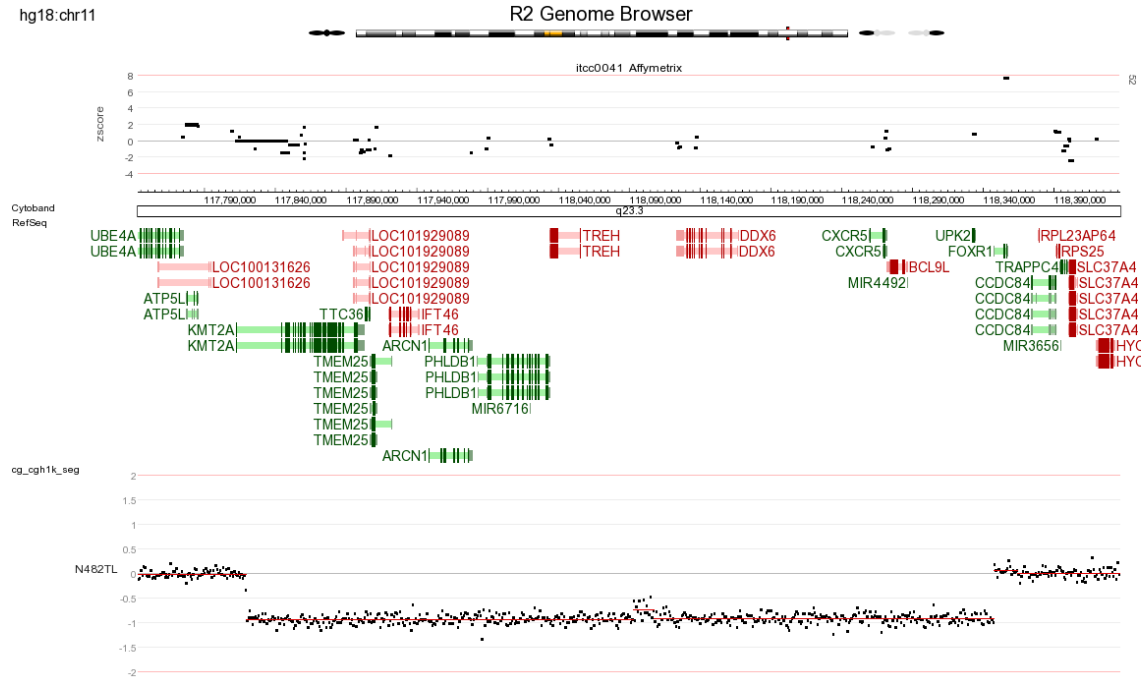


# Neuroblastoma patient with single event

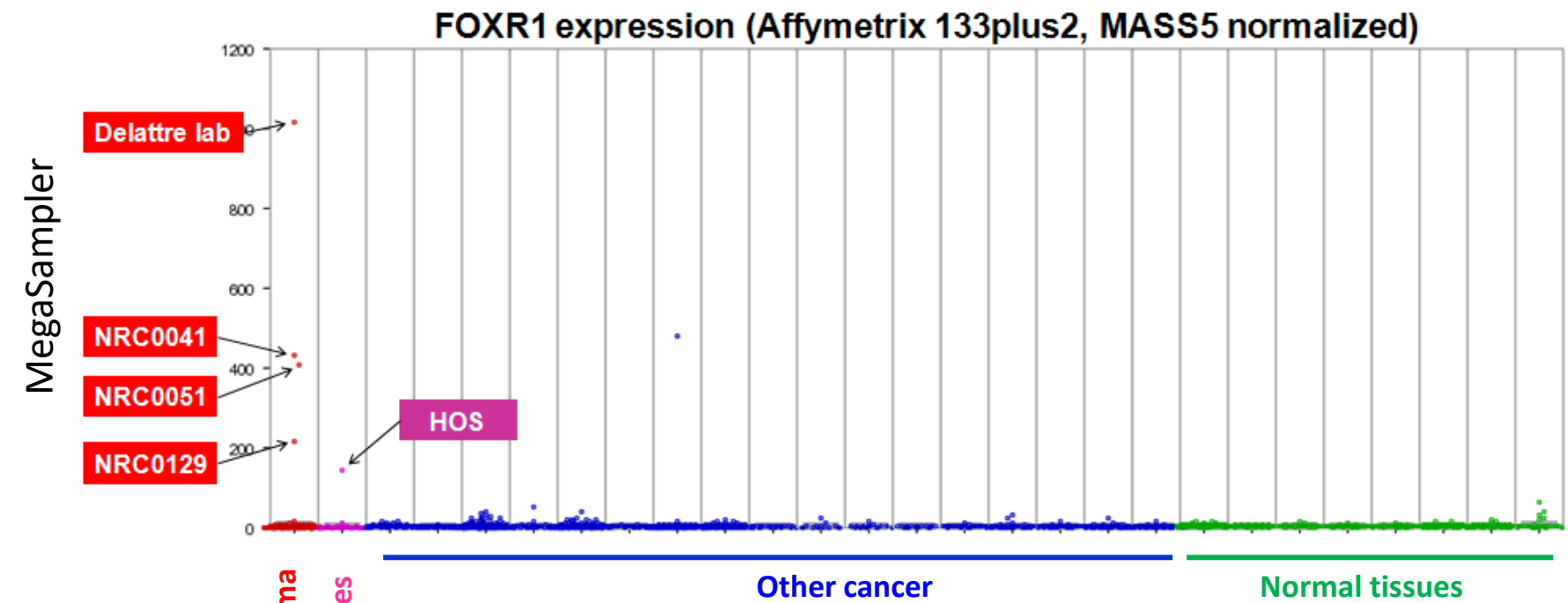




# Neuroblastoma patient with single event

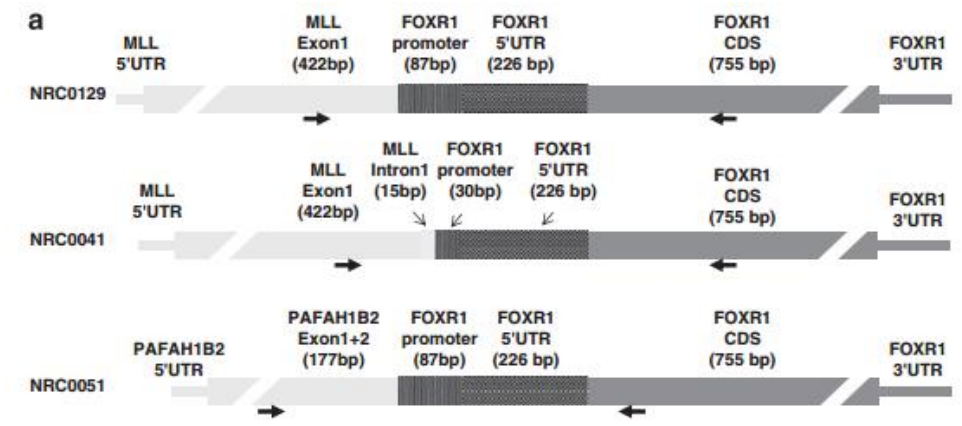


# FOXR1 only expressed in combination with Structural Variation

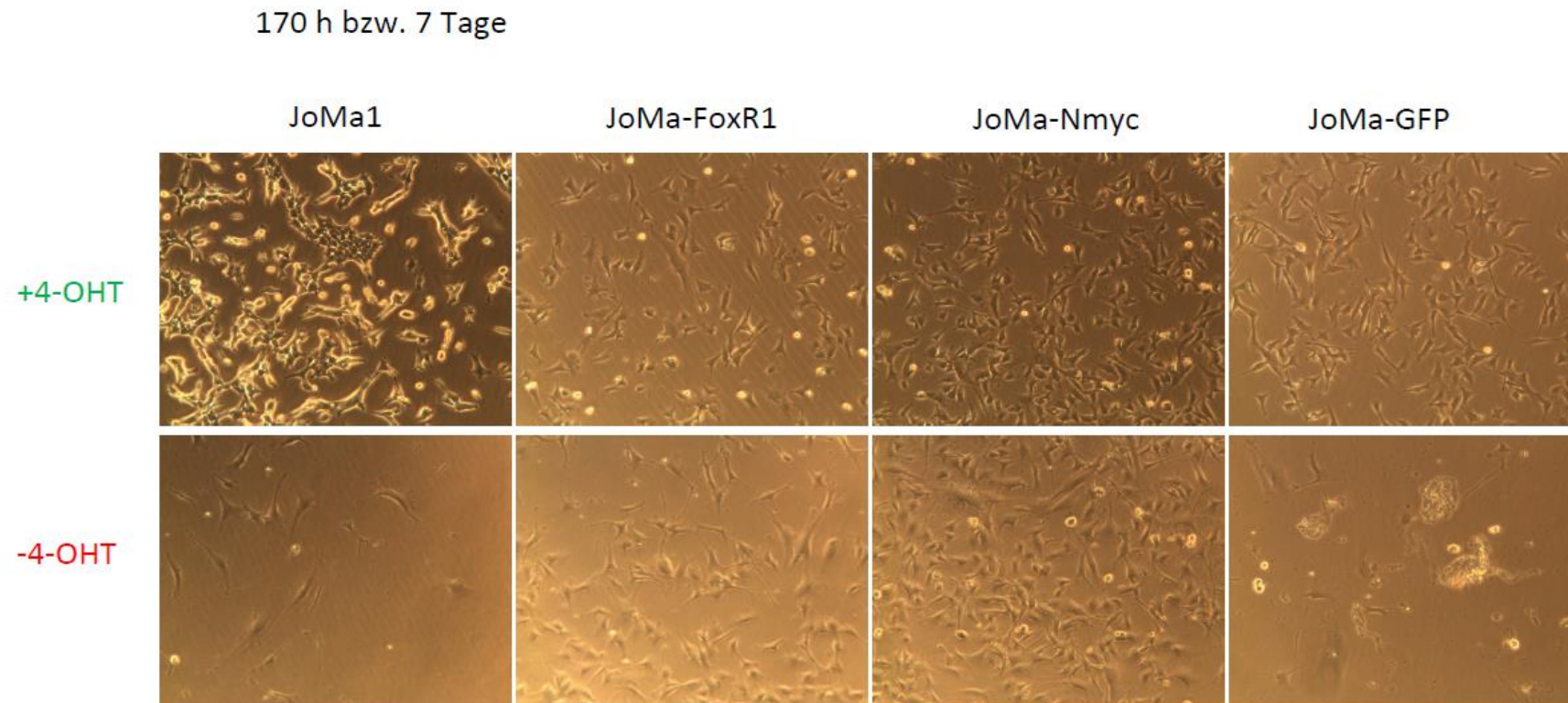


Neuroblastoma

Ped. cancer cell lines

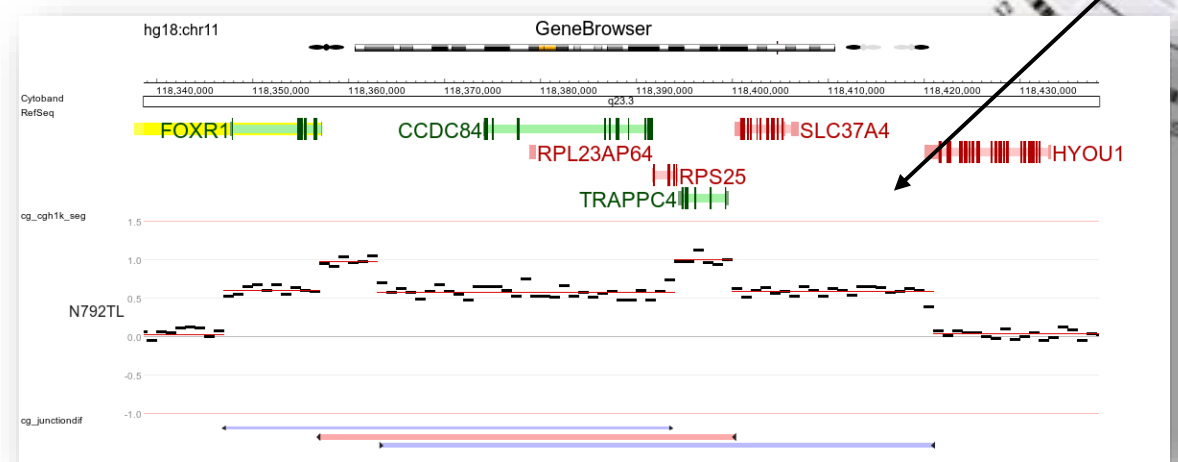
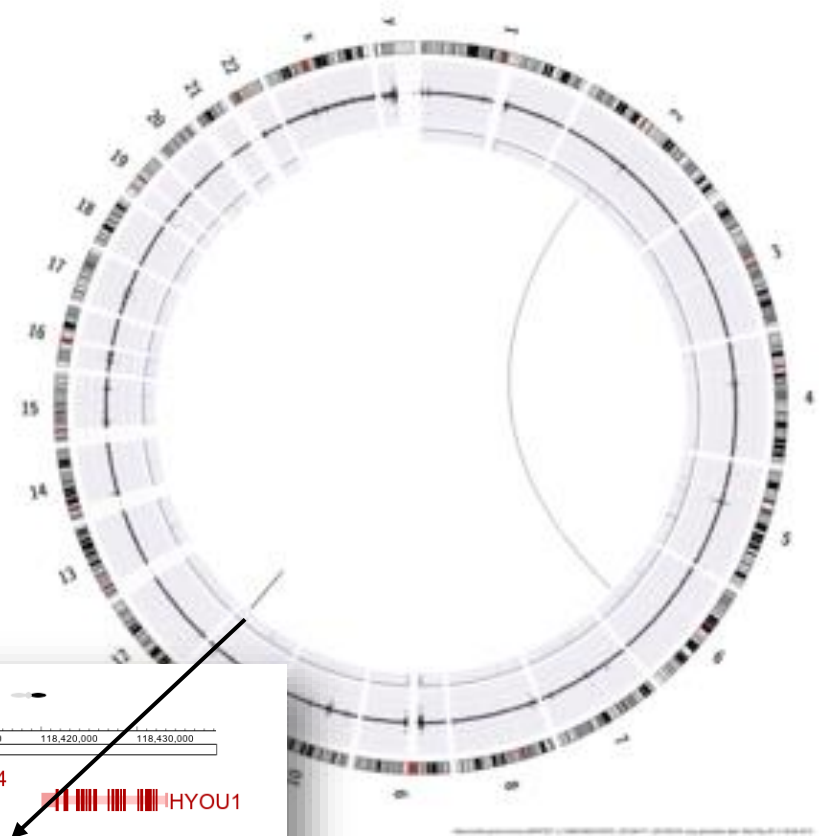


# FOXR1 causes growth maintenance in non malignant mouse neuroblasts

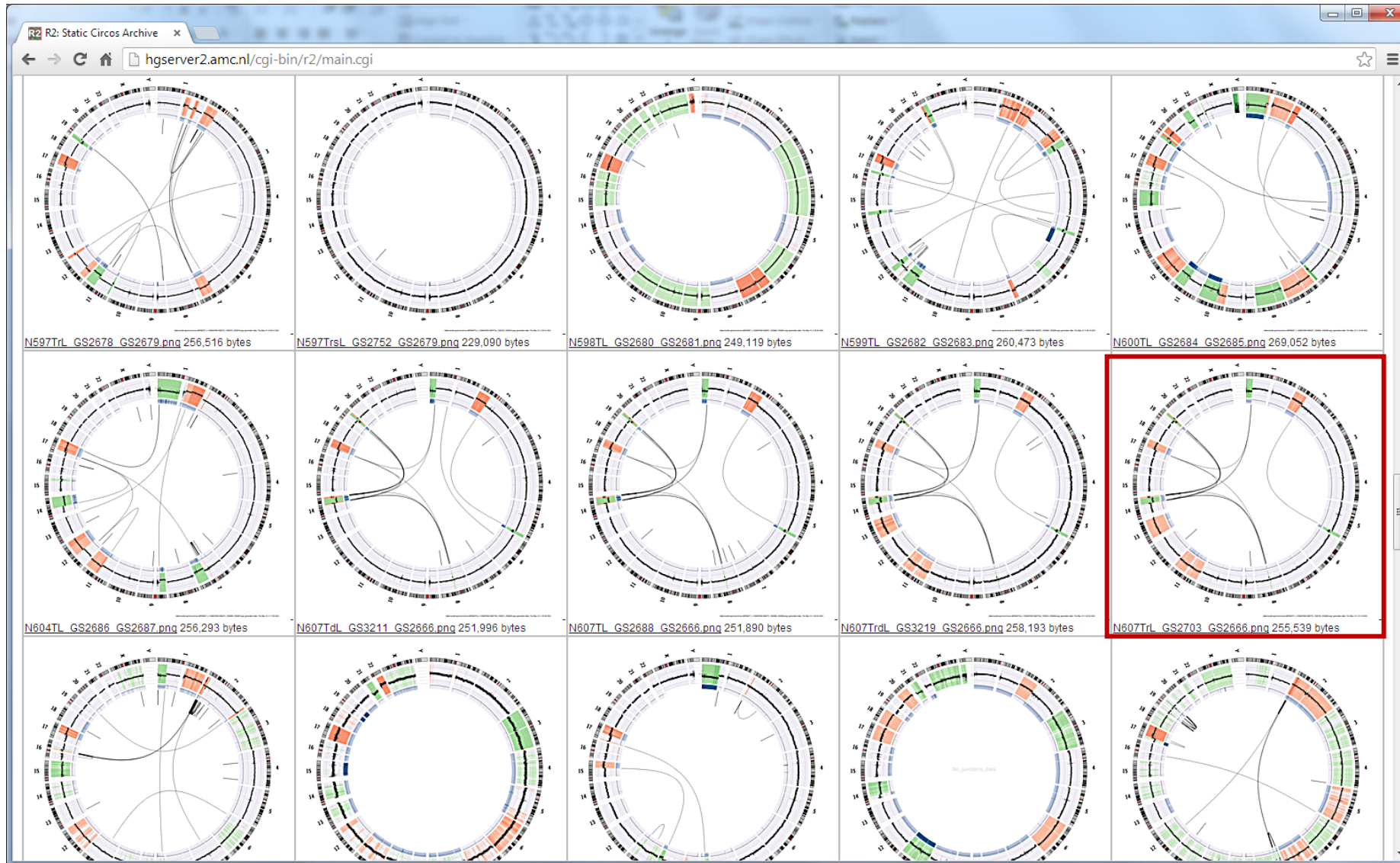


Immortalized by 4-oht regulated cMYC expression  
Removal of 4-oht > terminal differentiation

# Later on other patient with only a FOXR1 event



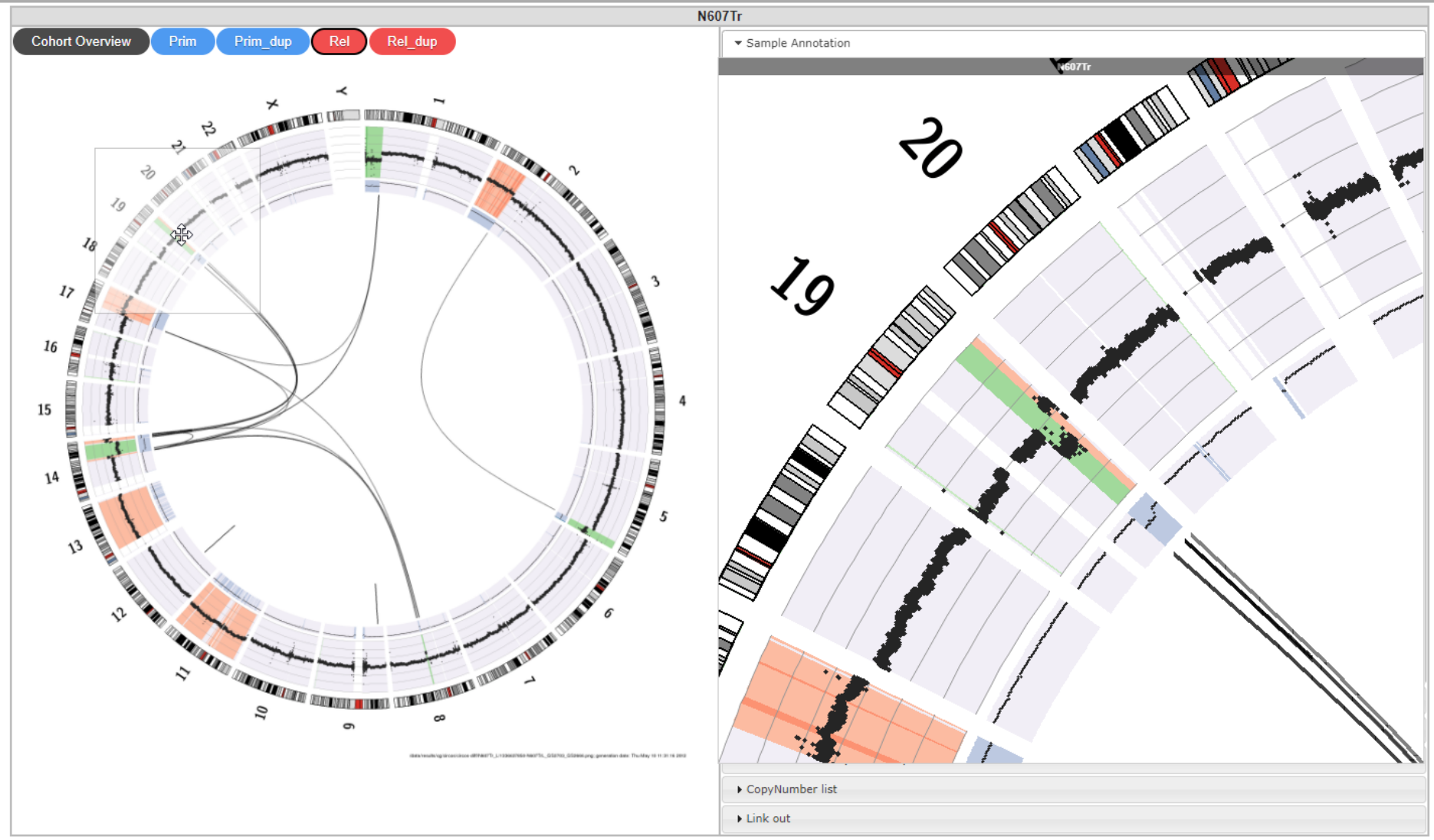
# Tumor board Patient overview



# Patient overview



# Patient overview



# Patient overview

Cohort Overview
Prim
Prim\_dup
Rel
Rel\_dup

N607Tr

Sample Annotation

▼ Somatic Structural Variants (23 entries)

Link	leftchr	leftpos	rightchr	rightpos	discordantmates	junctionseqres	event_type
<a href="#">view</a>	chr1	33844571	chr14	65953136	22	Y	complex
<a href="#">view</a>	chr1	33848331	chr14	65945324	22	Y	interchromosomal
<a href="#">view</a>	chr1	33852272	chr17	36301741	89	Y	complex
<a href="#">view</a>	chr12	48468933	chr12	48469387	12	Y	probable-inversion
<a href="#">view</a>	chr14	65947501	chr14	65955379	51	Y	tandem-duplication
<a href="#">view</a>	chr14	65953106	chr17	36301672	15	Y	complex
<a href="#">view</a>	chr14	65971282	chr14	104504570	61	Y	complex
<a href="#">view</a>	chr14	71013817	chr19	58837486	23	Y	interchromosomal
<a href="#">view</a>	chr14	98012077	chr19	63677849	64	Y	interchromosomal
<a href="#">view</a>	chr14	99329065	chr14	101061445	16	N	complex
<a href="#">view</a>	chr14	101061746	chr19	49302899	25	Y	complex
<a href="#">view</a>	chr14	101061766	chr19	49302899	20	Y	complex
<a href="#">view</a>	chr14	104550136	chr14	105674704	19	Y	complex
<a href="#">view</a>	chr14	104550325	chr19	58838952	29	Y	complex
<a href="#">view</a>	chr19	58838790	chr19	58839003	70	Y	complex
<a href="#">view</a>						Y	interchromosomal
<a href="#">view</a>						Y	interchromosomal
<a href="#">view</a>						N	probable-inversion
<a href="#">view</a>						Y	interchromosomal
<a href="#">view</a>						Y	interchromosomal
<a href="#">view</a>						Y	complex
<a href="#">view</a>						Y	interchromosomal
<a href="#">view</a>						Y	deletion

[Go to Main](#)

R2: GeneBrowser

hg18:chr1

View chr1:33802272-33902272 in GenomeBrowser

[Help\(93\)](#)

[Go to Main](#)

R2: GeneBrowser

hg18:chr17

View chr17:36251741-36351741 in GenomeBrowser

R2-support@amsterdamumc.nl

R2-Platform



# Patient overview



# Patient overview

The image displays two overlapping browser windows from the R2 platform. The top window shows a Circos plot of the genome with a table of somatic variants for sample N607TrL. The bottom window shows a detailed view of a variant on chromosome 18 (hg18:chr21).

**Table 1: Somatic Variants (Callidif Somatic Output (so >= .10))**

link	chrom	chromstart	var	r2_refflat	aa_change	category	Somatic	validation_result	reference	alleleseq
view	chr1	149546735	het_del PI4KB	F385L	FRAMESHIFT	0.113	na		A	
view	chr11	48323628	het_ins OR4C45	>10char	frameshift	0.167	na			AG
view	chr12	11066031	het_snp TAS2R19	P135S	MISSENSE	0.342	na		G	A
view	chr12	11066052	het_snp TAS2R19	V128L	MISSENSE	0.159	na		C	G
view	chr12	11075102	het_sub TAS2R31	I32T	MISSENSE	0.316	na		AA	GG
view	chr14	91123334	het_ins CATSPERB	GV1032GS	FRAMESHIFT	0.106	:			C
view	chr16	1817793	het_ins FAHD1	I187S	FRAMESHIFT	0.101	:			G
view	chr16	24829250	het_del SLC5A11	S591	DELETE	0.103	na		AGC	
view	chr19	4463888	het_sub PLIN4	T346I	MISSENSE	0.339	na		AG	GA
view	chr19	4463888	het_snp ALK	F1173L	MISSENSE	0.052	yes;as_expected		G	T
view	chr19	4463888	het_ins ZNF335			0.196	:			TTC
view	chr19	4463888	het_snp TIAM1	W1284*	NONSENSE	0.359	yes;as_expected		C	T
view	chr19	4463888	het_snp PLATN	Q148*	NONSENSE	0.567	na		C	T
view	chr19	4463888	het_ins ODZ3	GR588GS	FRAMESHIFT	0.695	na			G
view	chr19	4463888	het_del ZBTB24	>10char	FRAMESHIFT	0.76	na			>8bases

**Table 2: Variant Details (hg18:chr21)**

Variant	Effect	Impact	Cell lines	Tumor	TICs	AF	AF (95%)	AF (99%)	AF (99.9%)	AF (99.99%)	AF (99.999%)
chr21:31,430,151-31,430,151	het C>T (G=4)	High	0	4	0	0.359	0.359	0.359	0.359	0.359	0.359


# CiViC / Clinvar

R2: Static Circos Archive | R2: Single Sample Circos | R2: Two Gene View for N

Secure | <https://hgserver2.amc.nl/cgi-bin/r2/main.cgi>

Main | R2: Single Sample Circos Overview | Help (RTD)

I007\_013\_2M1



I007\_013\_2M1L

Sample Annotation

Somatic Variants

ClinVar

CiViC

gene	variant	disease	drugs	type	direction	level	significance	statement	pubmed_id	rating	chr	start	stop	varsum	origin
PDCD4	EXPRESSION	Lung Cancer	Paclitaxel	Predictive	Supports	B	Sensitivity	Info	<a href="#">25928036</a>	4	10	112631596	112659763	Info	Somatic Mutation
PTCH1	LOH	Brain Medulloblastoma	Vismodegib	Predictive	Supports	B	Sensitivity	Info	<a href="#">26169613</a>	2	9	98205262	98270943		Somatic Mutation
PTCH1	MUTATION	Brain Medulloblastoma	Vismodegib	Predictive	Supports	B	Sensitivity	Info	<a href="#">24651015</a>	4	9	98205262	98270943		Somatic Mutation
REL	AMPLIFICATION	Diffuse Large B-cell Lymphoma		Diagnostic	Supports	B	Positive	Info	<a href="#">12075054</a>	4	2	61108709	61149800		Somatic Mutation
TSC1	FRAMESHIFT TRUNCATION	Invasive Bladder Transitional Cell Carcinoma	Everolimus	Predictive	Supports	B	Sensitivity	Info	<a href="#">22923433</a>	3	9	135766735	135820008	Info	Somatic Mutation
TSC1	FRAMESHIFT TRUNCATION	Non-small Cell Lung Carcinoma	Rapamycin (Sirolimus)	Predictive	Supports	D	Sensitivity	Info	<a href="#">19966866</a>	3	9	135766735	135820008	Info	Somatic Mutation
TSC1	LOSS-OF-FUNCTION	Bladder Carcinoma	Everolimus	Predictive	Supports	B	Sensitivity	Info	<a href="#">22923433</a>	3	9	135766735	135820008	Info	Somatic Mutation

Gene Expression list

11:58  
4/11/2017

Other patient

# Gene Expression

Go to: [Main](#) | [Graduate Training Course portal](#)

[Online Tutorial](#)

R2: Single Sample Circos Overview

**st4 | non-amp (N170T)**

Cohort Overview

**Tumor Neuroblastoma (combat)**  
Versteeg - 122 - MAS5.0(bc) - u133p2  
Marked: **itcc0061**

**Expression**

**Tumor Neuroblastoma (combat)**  
Versteeg - 122 - MAS5.0(bc) - u133p2  
NG\_stage(inss) vs ATF1 (222103\_at)  
p=0.099 (anova)

Tumor Neuroblastoma (combat) - Versteeg - 122 - MAS5.0(bc) - u133p2 (sample\_id=42)  
Using Genelist: geneannot::base::cancer\_gene\_census::cancer\_gene\_census

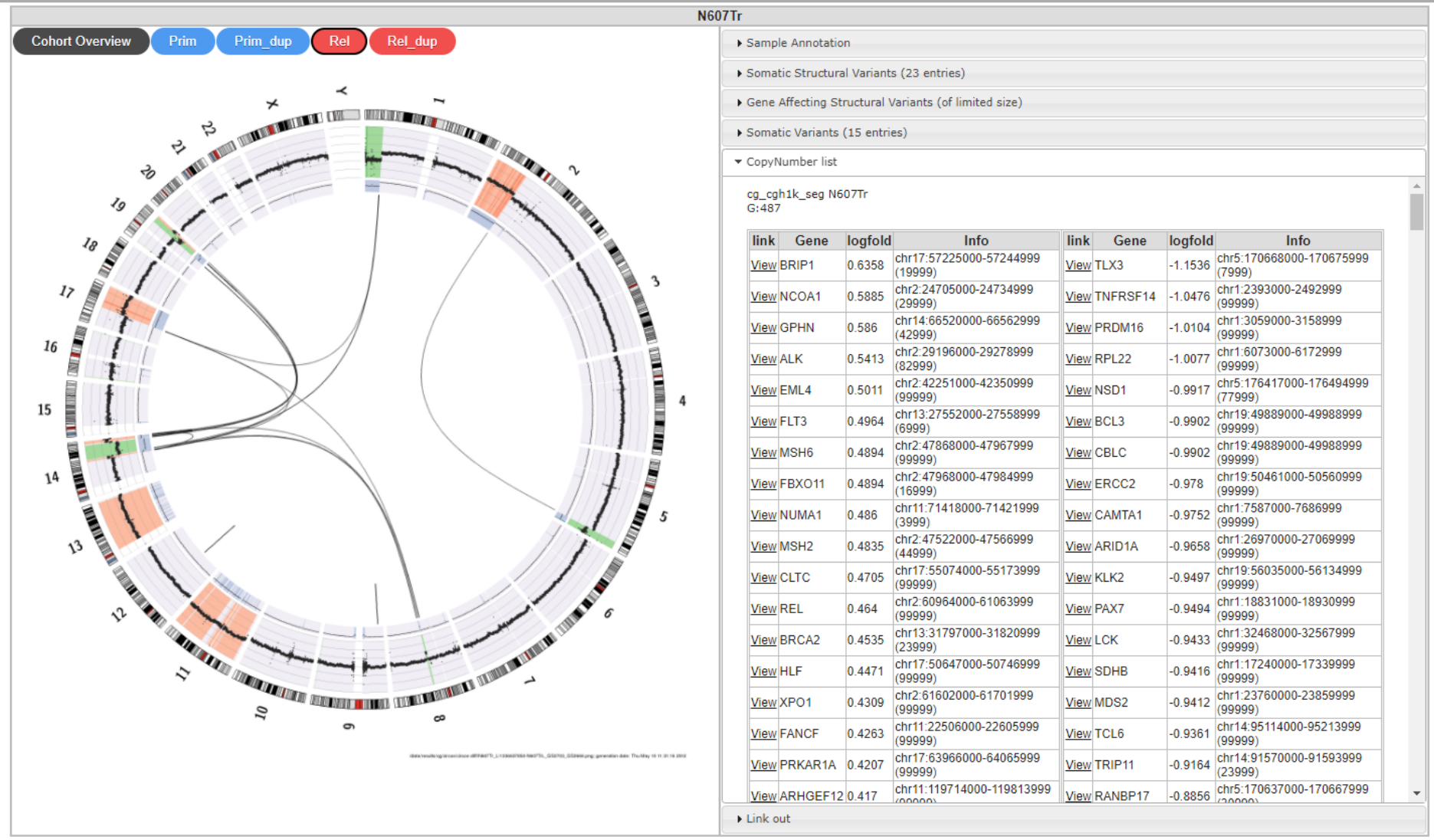
Plot Top10 Genes  
View Top25 Overview

GeneView	Gene	zscore	Detailed
222103_at	ATF1	2.501	<a href="#">Detailed</a>
235215_at	ERCC4	2.430	<a href="#">Detailed</a>
203409_at	DDB2	2.330	<a href="#">Detailed</a>
210793_s_at	NUP98	1.931	<a href="#">Detailed</a>
227891_s_at	TAF15	1.781	<a href="#">Detailed</a>
208212_s_at	ALK	1.655	<a href="#">Detailed</a>
223068_at	EML4	1.610	<a href="#">Detailed</a>
212287_at	SUZ12	1.593	<a href="#">Detailed</a>
221007_s_at	FIP1L1	1.479	<a href="#">Detailed</a>
201244_s_at	RAF1	1.438	<a href="#">Detailed</a>
204379_s_at	FGFR3	1.383	<a href="#">Detailed</a>
204159_at	CDKN2C	1.352	<a href="#">Detailed</a>
215088_s_at	SDHC	1.349	<a href="#">Detailed</a>
219492_at	CHIC2	1.313	<a href="#">Detailed</a>

patient

Below, button to scan all genes

# Copy number events by genes



The cn level of gene-span of a selection of genes is shown

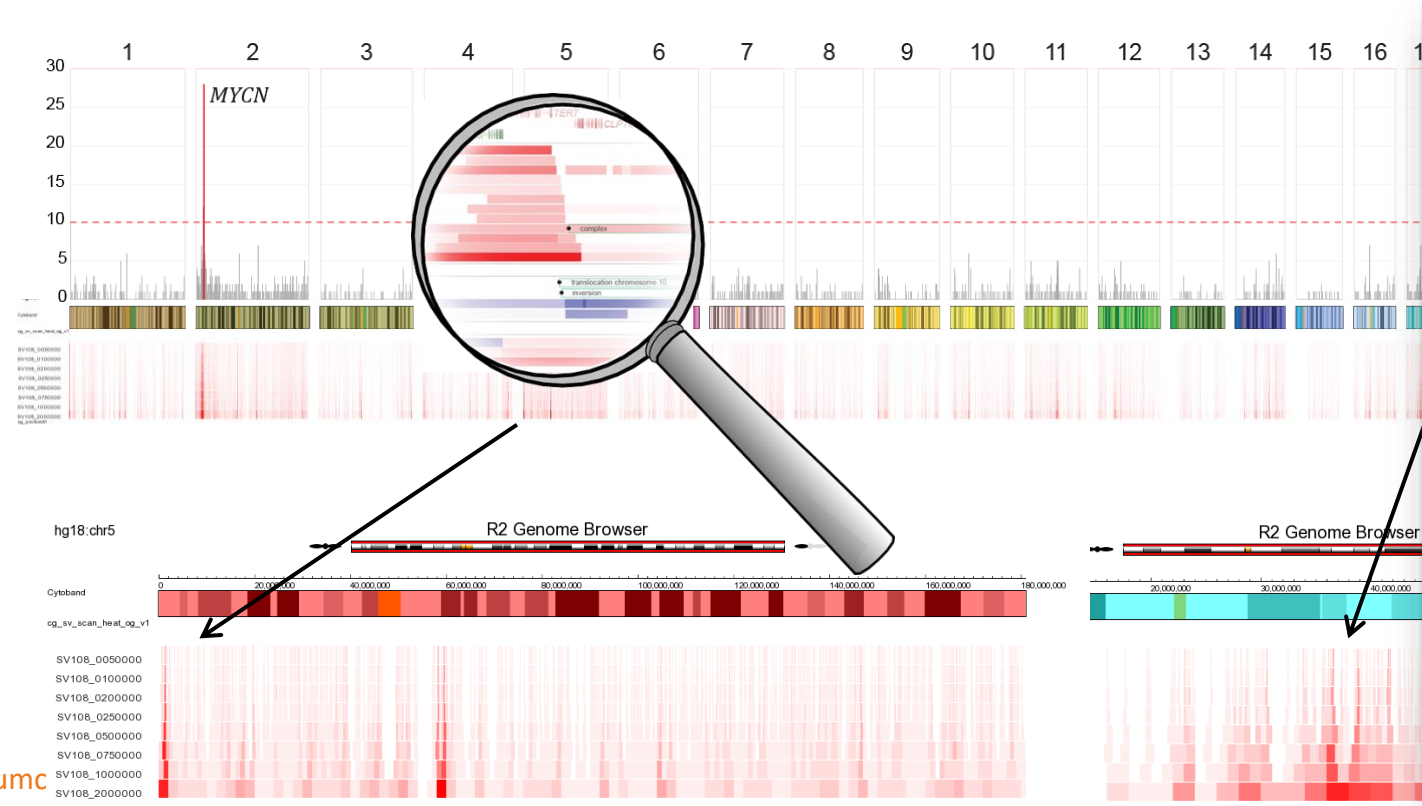
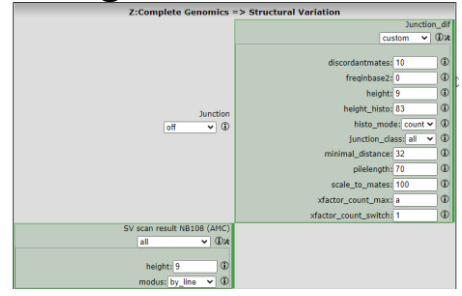
# Structural Variations



# Scan for Enrichment in SV

- Are there hotspots of SV in the neuroblastoma genome?

R2 genome browser



Gene	#_samples	Where
MYCN	28	SURR
TERT	13	US
ATRX	8	INTRA
TENM3	6	INTRA
PDE4D	6	INTRA
PTPRD	6	INTRA
GATA3	5	DS
CCND1	5	US
SHANK2	5	INTRA
PSEN1	5	US
ASCC2	5	INTRA
EIF4E	5	DS
MAML3	4	INTRA
CSMD2	3	INTRA
ADAM12	3	DS
UNC79	3	INTRA
ZFH3	3	INTRA
ASIC2	3	INTRA
STAG2	3	INTRA

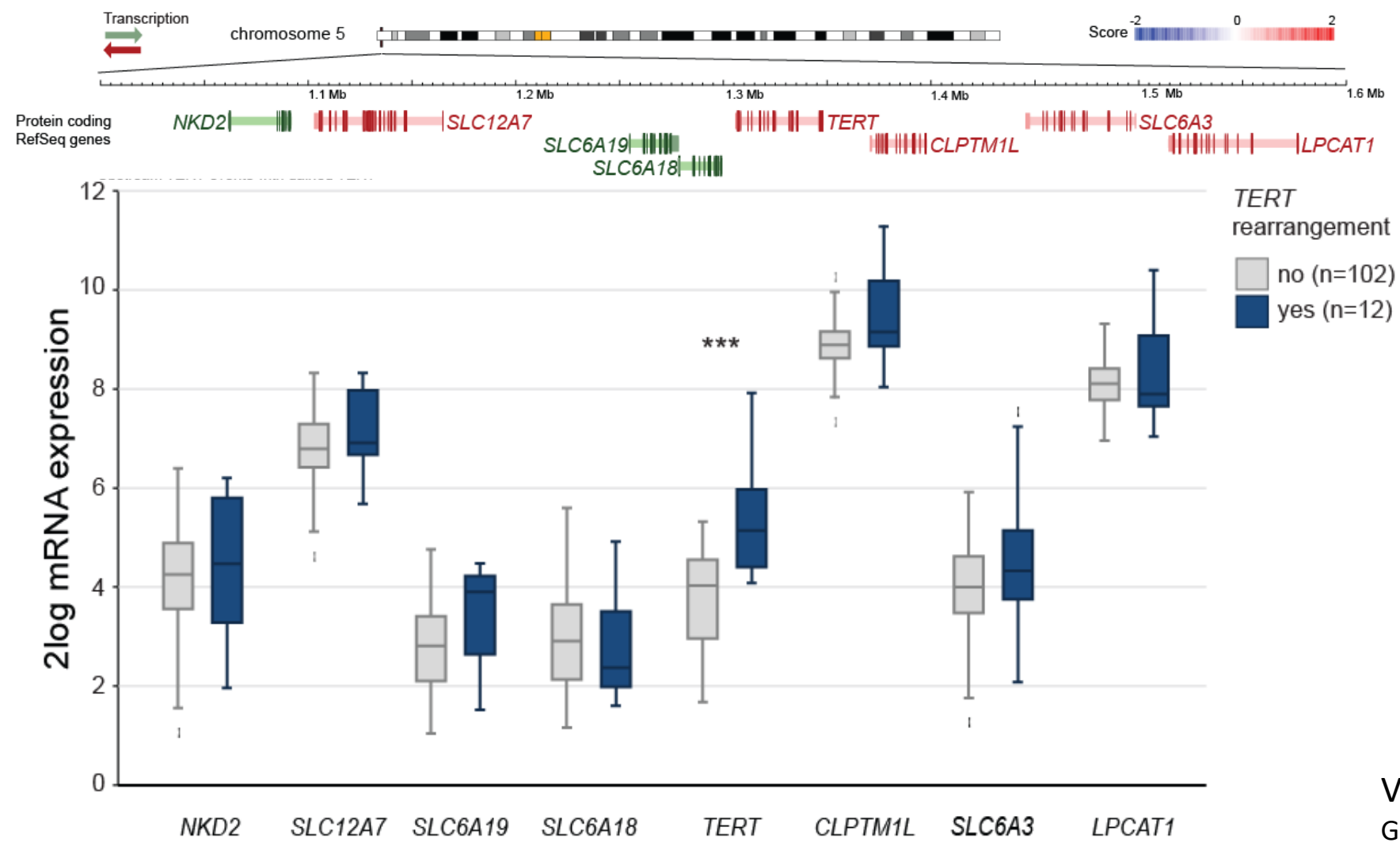
108 selected cases

# TERT region has gains, losses and inversions in 23% of high stage NB



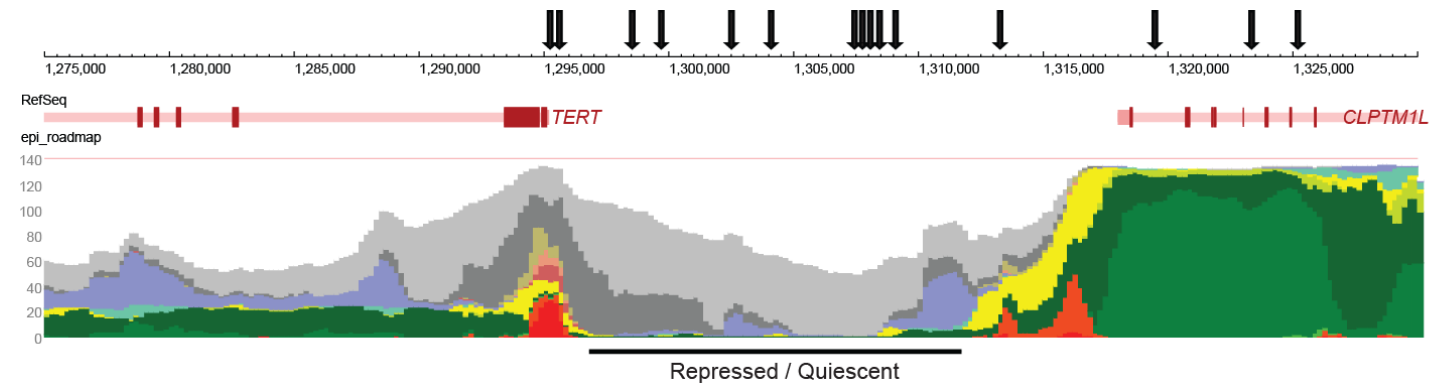


# TERT expression affected



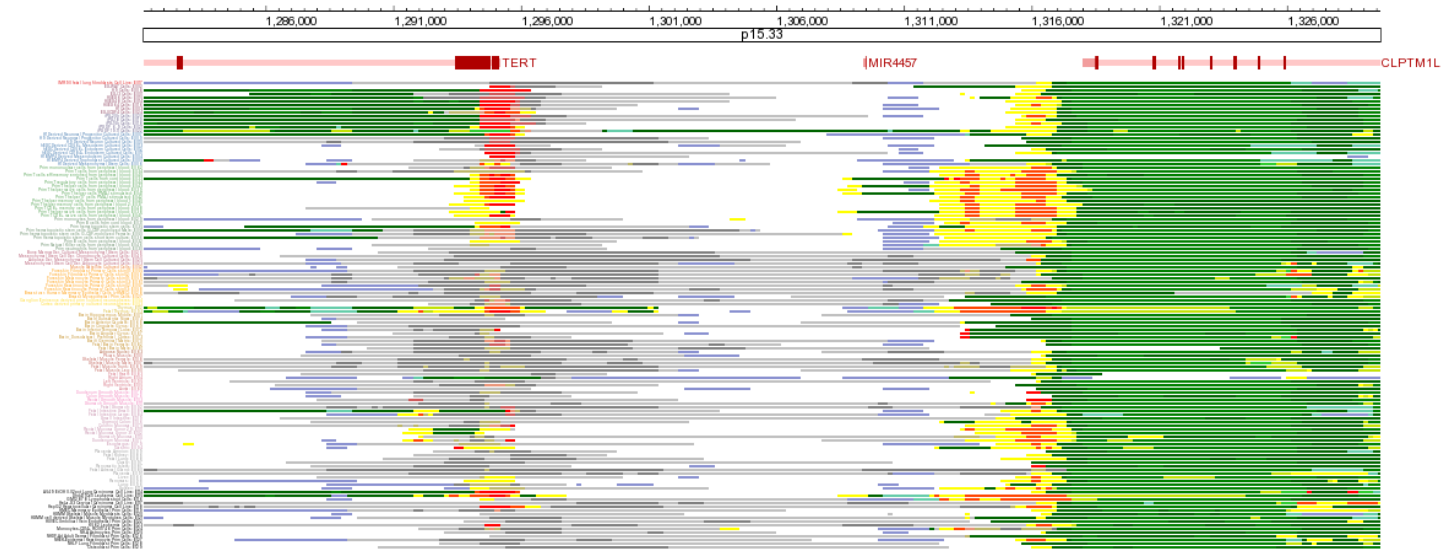
View Multiple Genes  
Genes in order of genome

# TERT upstream region mostly quiescent / repressed



NIH epi roadmap HMM15

- Active TSS
- Flanking Active TSS
- Transcr. at gene 5 and 3
- Strong transcription
- Weak transcription
- Genic enhancers
- Enhancers
- ZNF genes & repeats
- Heterochromatin
- Bivalent/Poised TSS
- Flanking Bivalent TSS/Enh
- Bivalent Enhancer
- Repressed PolyComb
- Weak Repressed PolyComb
- Quiescent/Low



NIH Epigenome Roadmap

all

height: 10

modus: overview

NIH Epigenome Roadmap

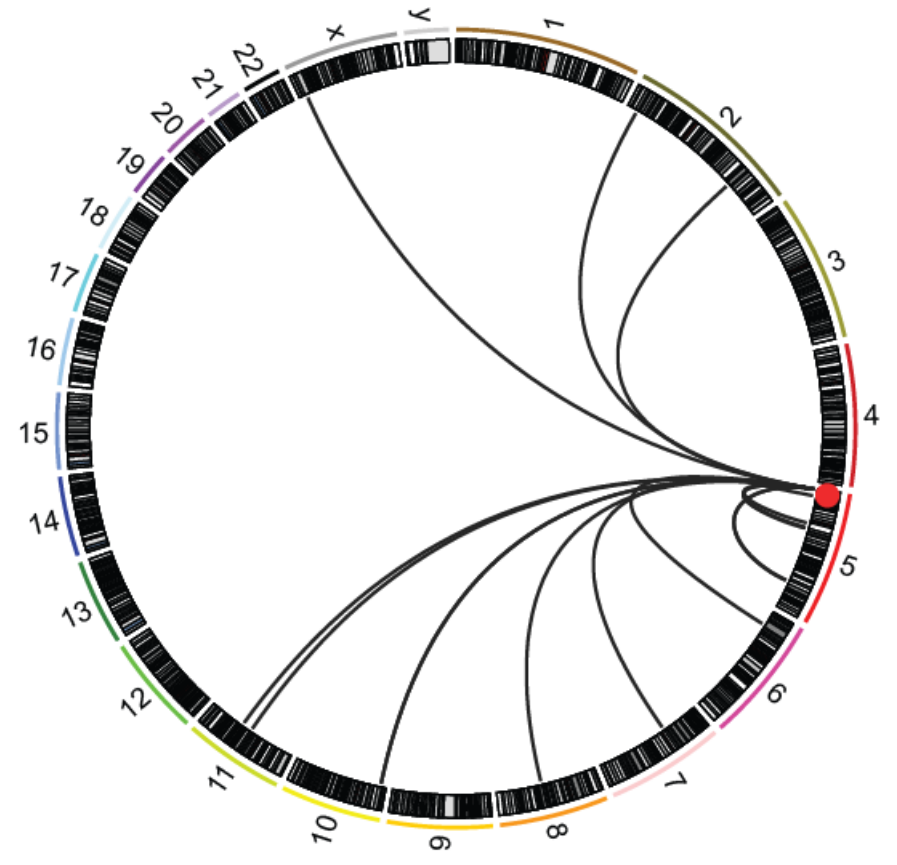
all

height: 10

modus: plugin

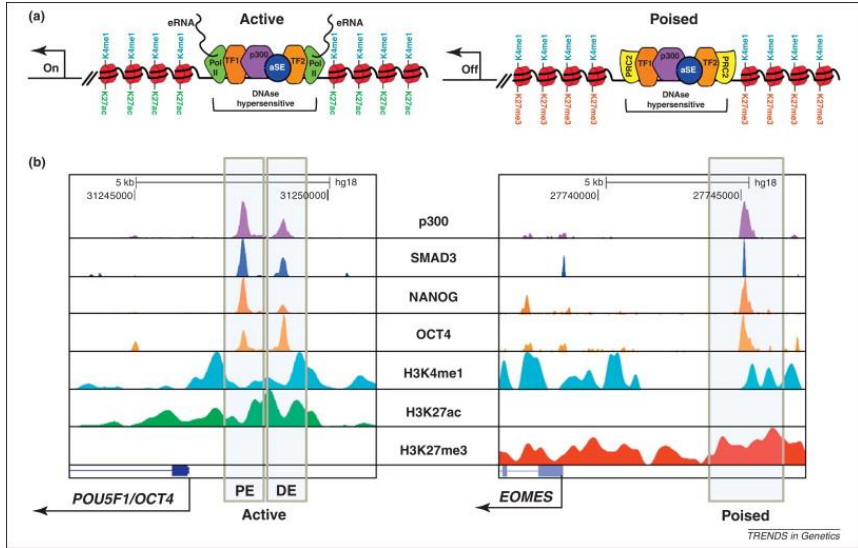
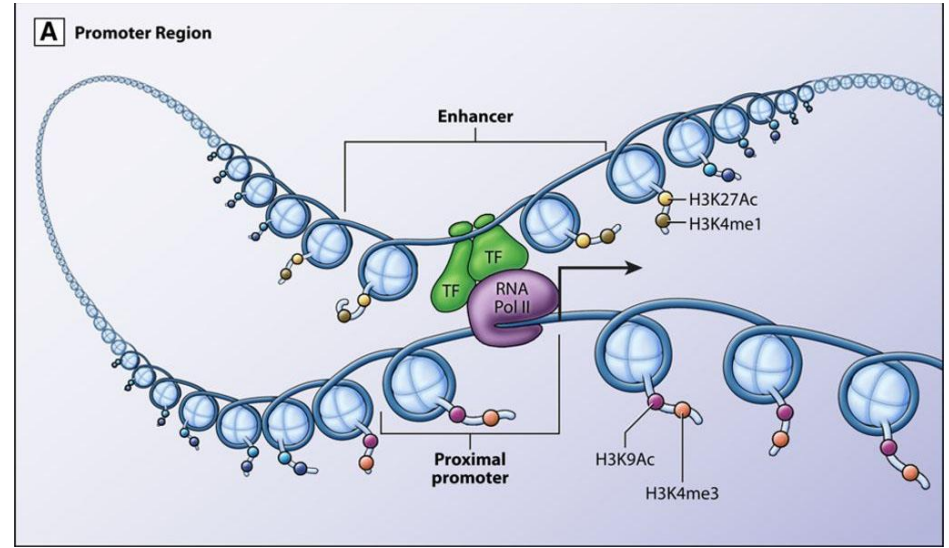
Valentijn & Kosters et al. 2012

# TERT rearrangements have many different translocation partners

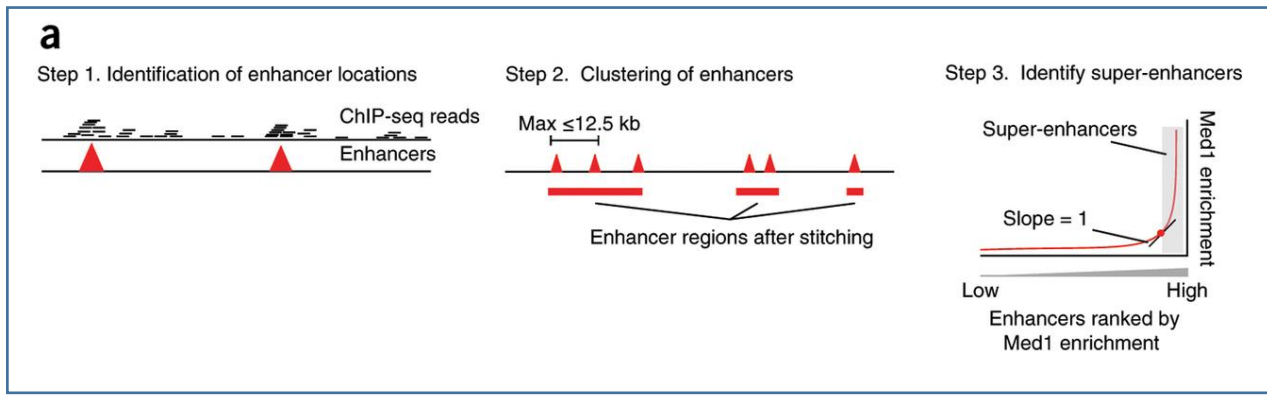


Valentijn & Koster et al 2015 Nat. Genet

# Enhancers

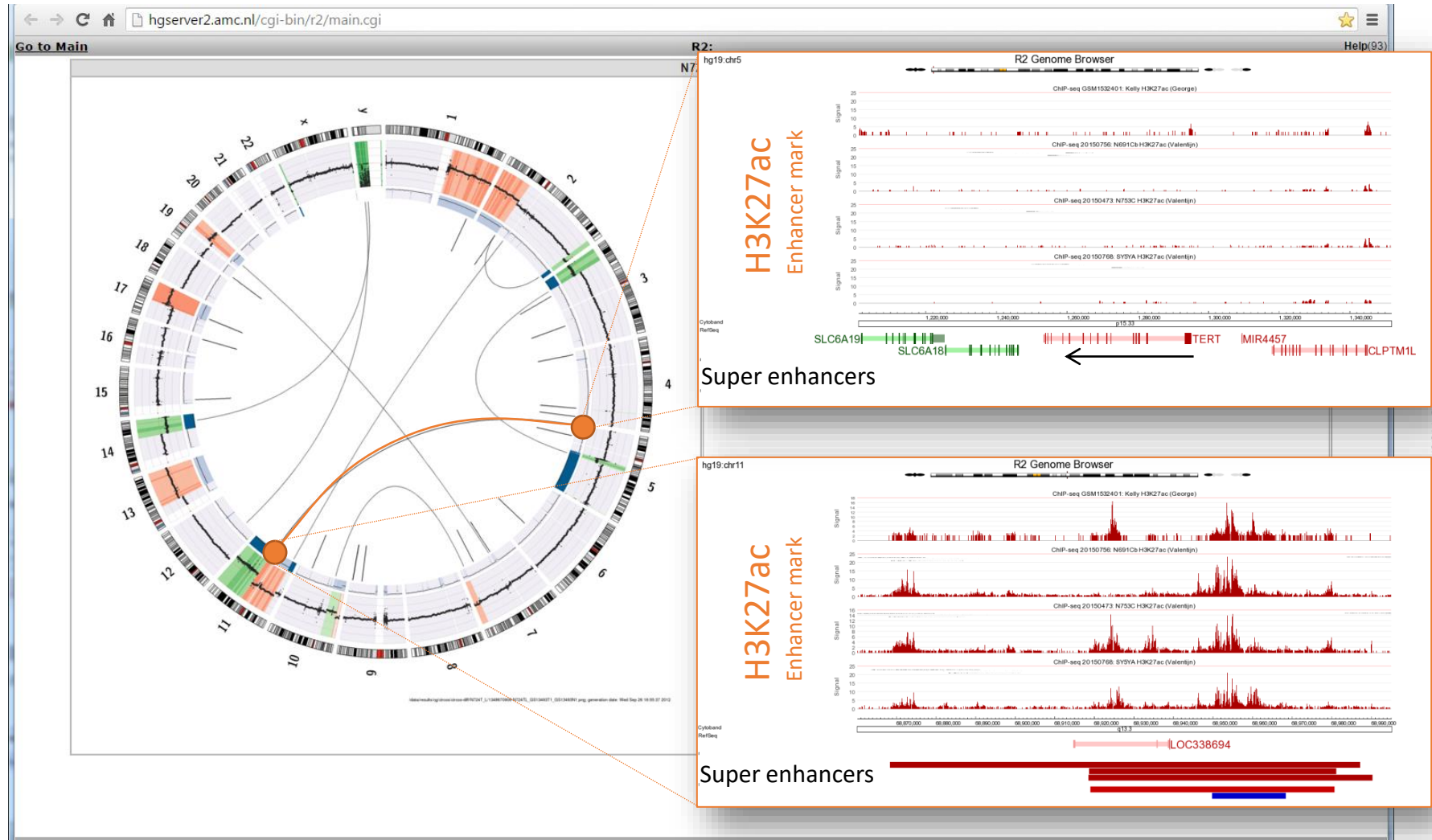


Active enhancers are marked by H3K27Ac



Super Enhancers are the most 'active / enriched' enhancers

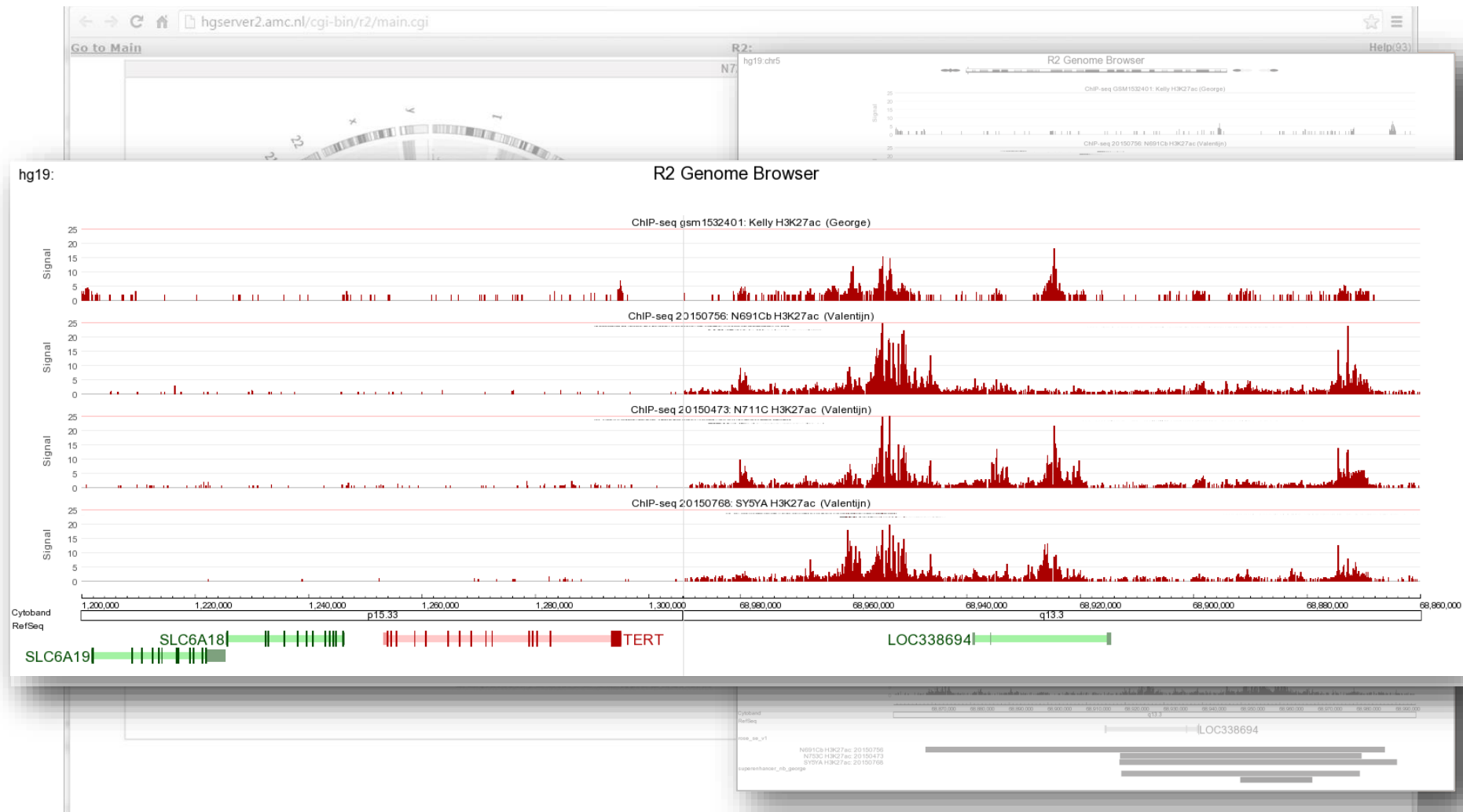
# TERT translocates to super-enhancers



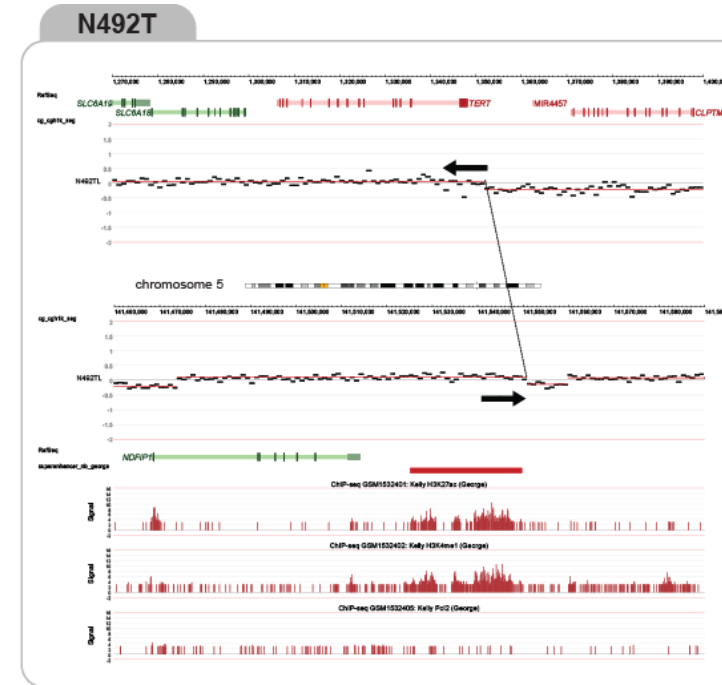
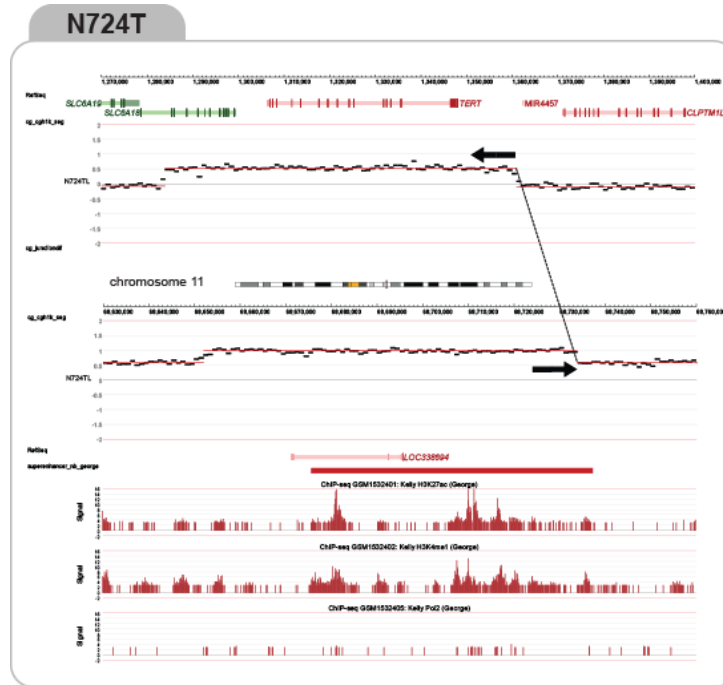
Valentijn & Kosters et al 2015 Nat. Genet.

We will cover ChIP profiles later

# TERT translocates to super-enhancers



# TERT translocates to super-enhancers

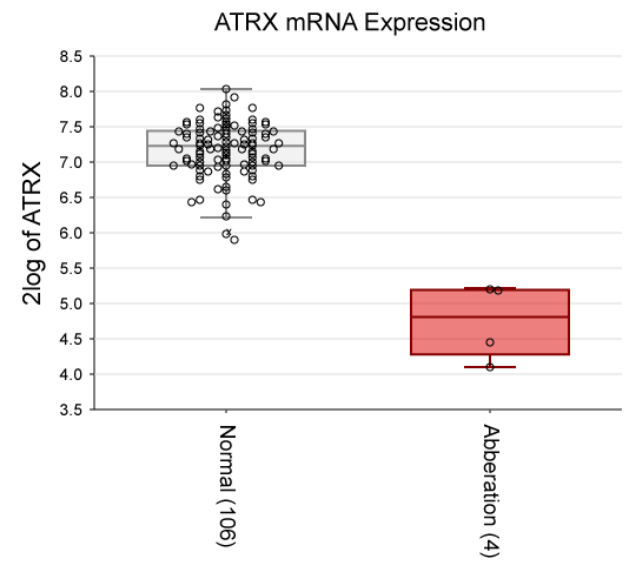
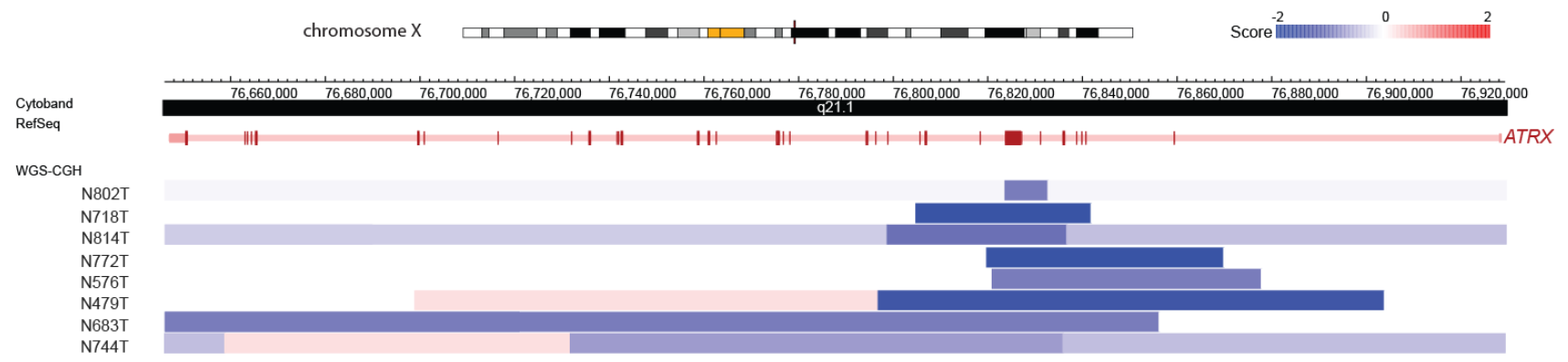


Valentijn & Koster et al 2015 Nat. Genet

	r100k	r250k	r500k	r750k	r1000k
p count>=5	0.00034				
p count>=6		0.0011			
p count>=7			0.00306		
p count>=9				0.00045	0.00234

100,000 iterations of random breakpoints

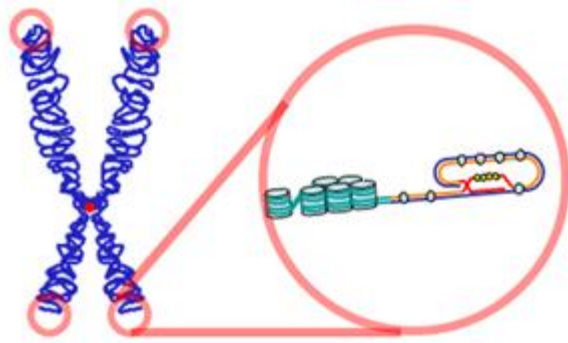
# ATRX impairment => ALT



Valentijn & Koster et al 2015 Nat. Genet



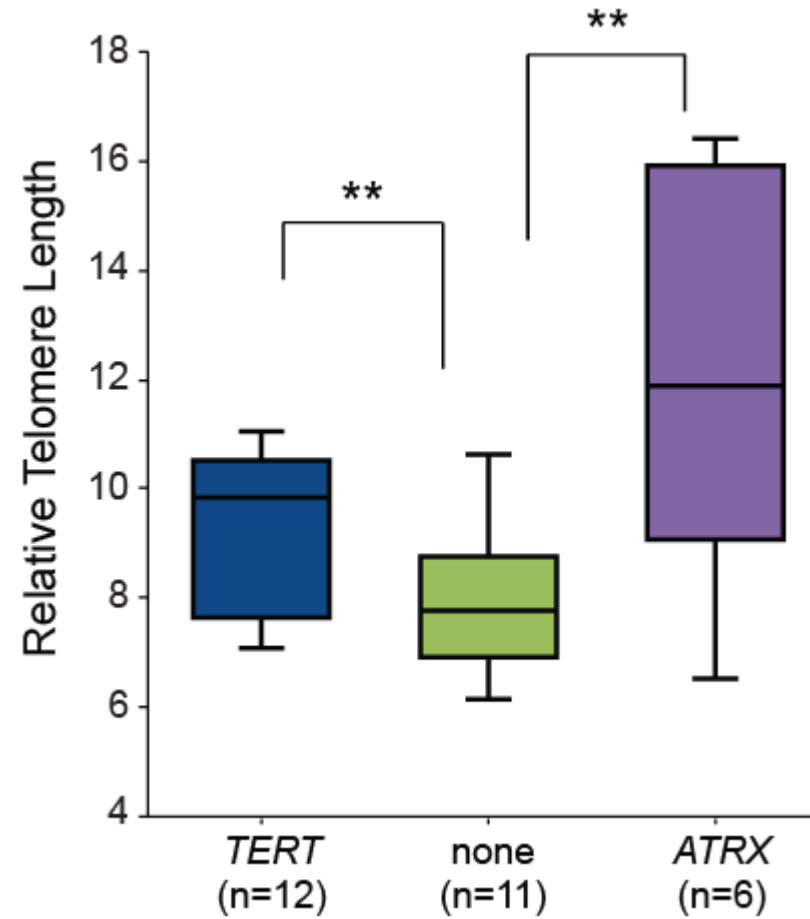
# Telomeres length NGS



Telomeres are  $(TTAGGG)_n$   
Search for reads containing multiple instances ( $\geq 4$ ) of this sequence and relate to total reads

## Small Tools

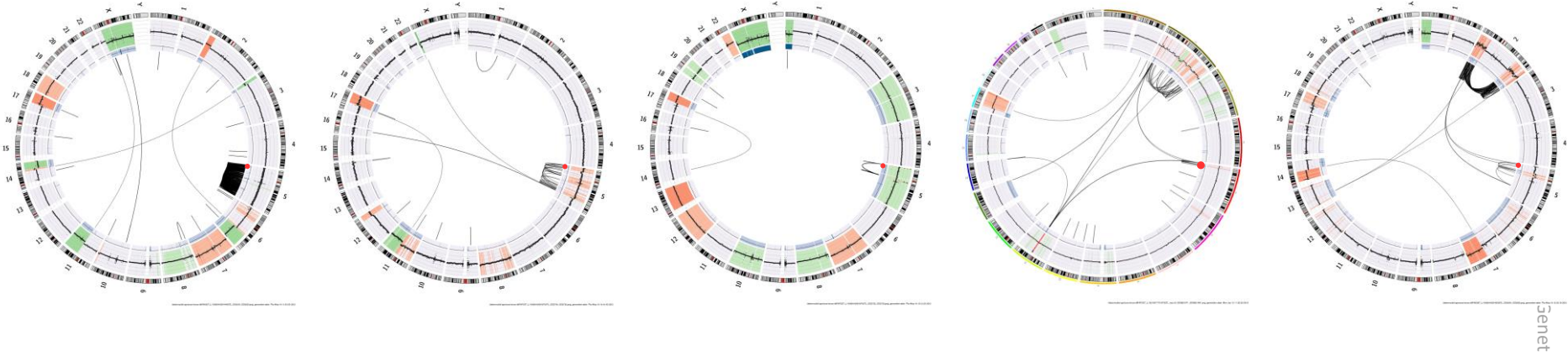
- [Gene Ontology on R2 texture](#)
- [Draw a Venn diagram](#)
- [Create Boxplot with user data](#)
- [Sample Ranked GeneSet Scores \(Alpha\)](#)
- [Strip header from R2 result file](#)



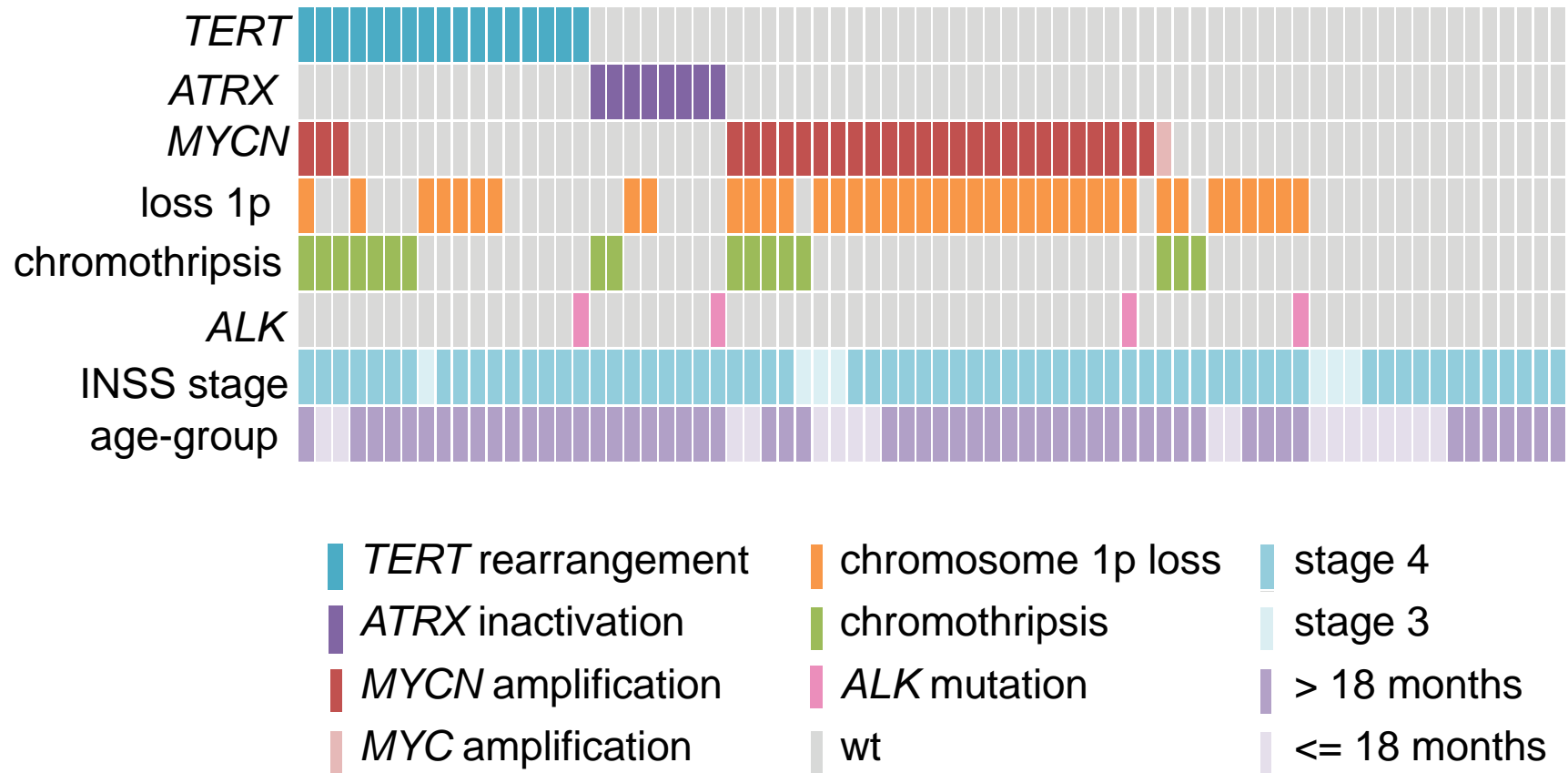
Valentijn & Koster et al 2015 Nat. Genet

# Chromothripsis

- 5 chromothripsis cases directly affect TERT

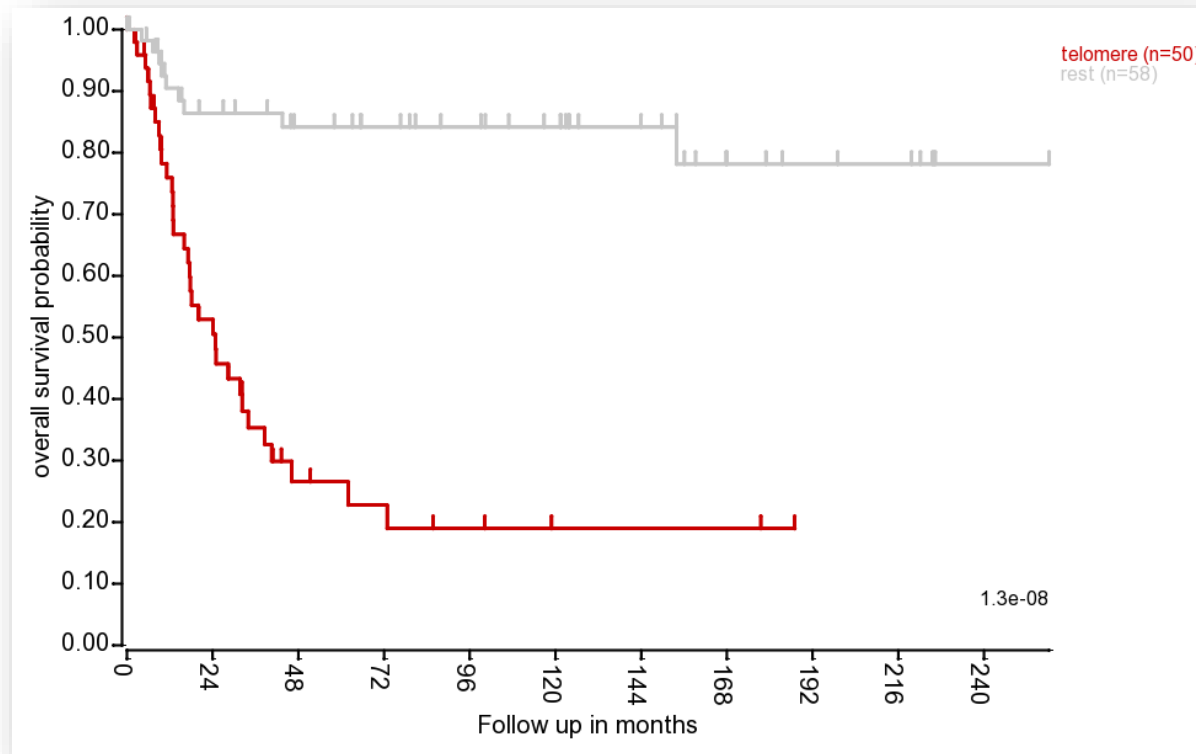


# Clinical / Molecular association



Valentijn & Koster et al 2015 Nat. Genet

# Clinical / Molecular association



Valentijn & Koster et al 2015 Nat. Genet

Kaplan by groups

# ChIPseq analysis

The screenshot displays the R2: ChIP web application interface. The browser window shows the URL `hgserver1.amc.nl/cgi-bin/r2/main.cgi`. The page layout includes a navigation menu on the left, a main content area with various tool links, and a footer with contact information.

**Navigation Menu (Left):**

- Main
- Time series
- AmpliconView
- Kaplan Meier
- Tools
- t-SNE Maps
- Genome Browser
- ChIP data** (selected)
- Change Data Scope
- User Options
- Help
- Contact / About R2

**Main Content Area (Right):**

**R2: ChIP**

- [ChIP \(Transcriptional Start Site\)](#)
- [TSS graphs within experiment](#)
- [TSS graphs](#)
- [TSS genesets scan](#)
- [TSS XY plot 2 experiments](#)
- ChIPseq**
- [ROSE Super Enhancer Plot](#)
- [MACS Plot](#)
- [RSEG Plot](#)
- [ChIPseq TSS Peak/Coverage Plotter](#)
- ChIP Browser**
- [ChIP-chip Genome Browser](#)
- [ChIP-chip Genome Browser \(+GEx\)](#)

**Additional Elements:**

- PubMed** logo with a 3D figure holding a green checkmark and a stack of books.
- r2.amc.nl** logo.
- Integrated Analysis of Tumor Genomics Data with R2 Tutorials (with R2 logo).
- Download the R2 Tutorials Book** button.

**Footer:**

- AMC: Oncogenomics
- [Go to Main](#)
- encca (2)

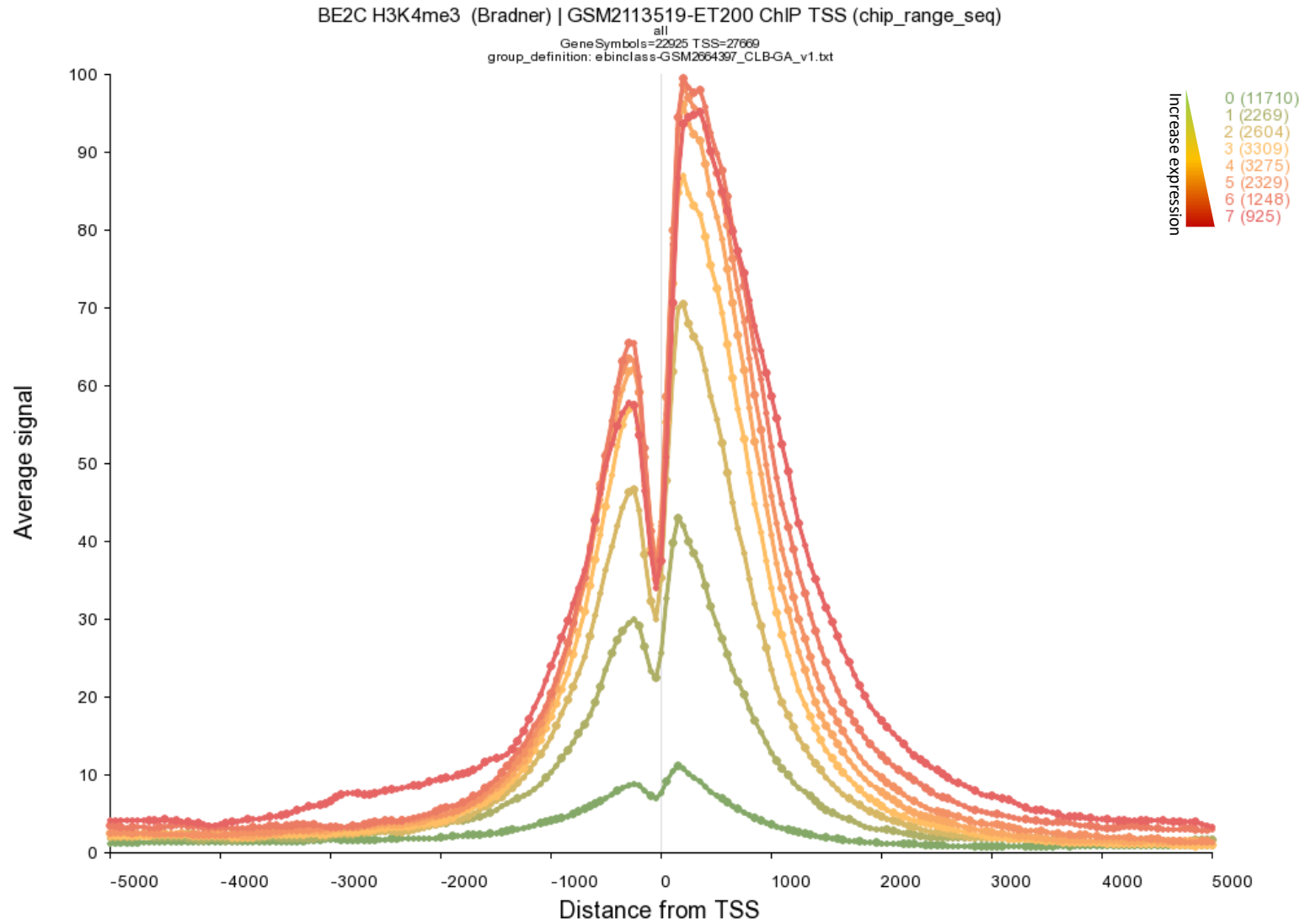
# ChIPseq analysis

The screenshot displays the R2 ChIPseq analysis interface. On the left, a plot titled 'ROSE' shows 'Signal SHEP2 H3K27ac (Valentijn) - Input' on the y-axis (0 to 180,000) and 'Rank' on the x-axis (-10,000 to -70,000). A red line shows a sharp peak at rank -70,000. An arrow points from this peak to the 'R2 Genome Browser' view below. The browser shows 'hg19:chr5' with a signal plot for 'ChIP-seq 20150766: SHEP2 H3K27ac (Valentijn)'. The x-axis is 'q11.2' with coordinates from 56,000,000 to 58,400,000. Below the plot, gene models for C5orf67, MAP3K1, SETD9, MIER3, GPBP1, LINC-R-0003, LOC101928505, ACTBL2, LOC101928539, LOC101928569, PLK2, GAPTI, RAB3C, and PDE4D are shown. A 'ROSE Super Enhancer Plot' is highlighted in the navigation menu on the right.

# ChIPseq analysis

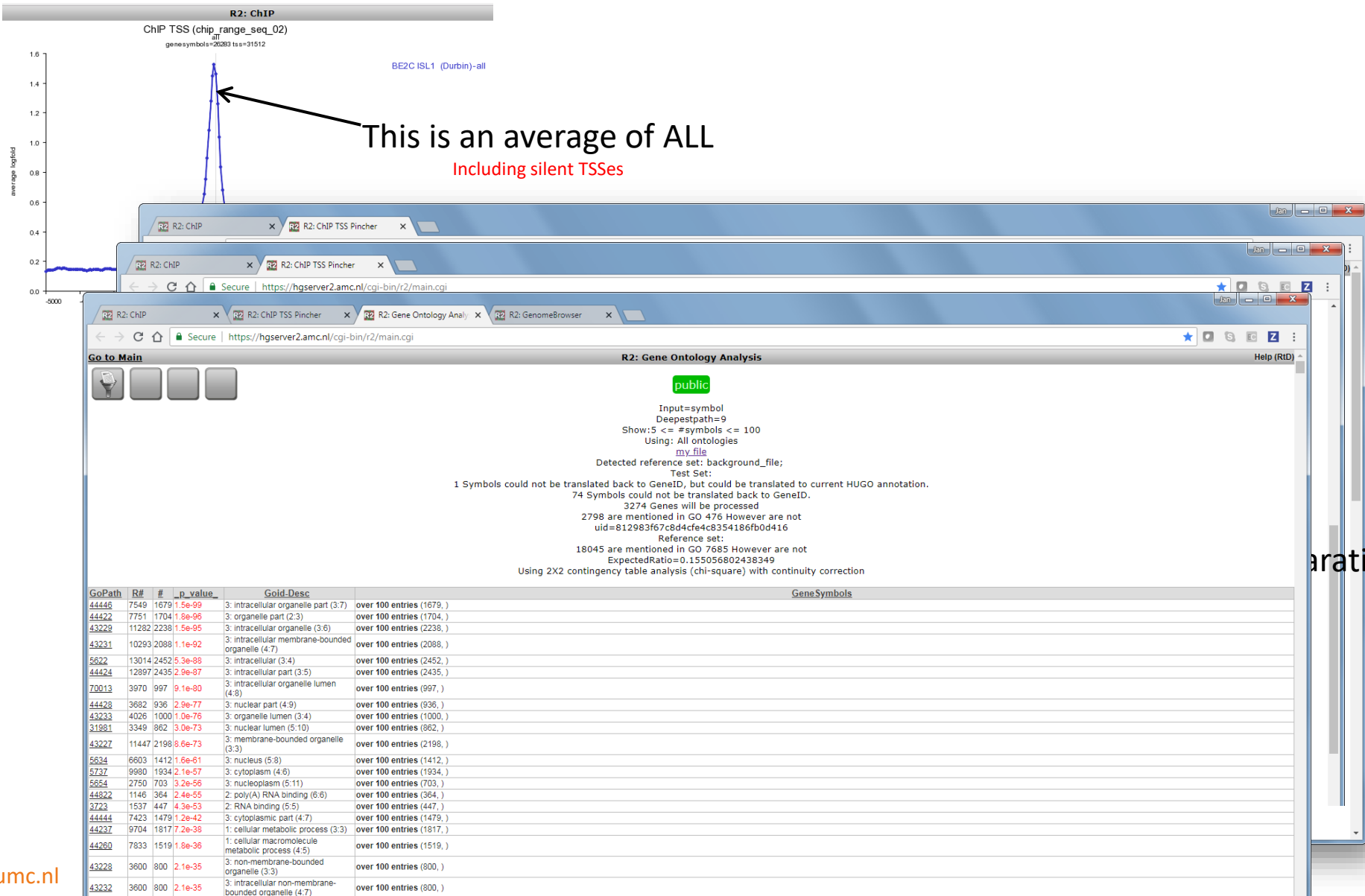
The screenshot displays the R2 ChIPseq web interface. On the left is a navigation menu with options like 'Main', 'Time series', 'AmpliconView', 'Kaplan Meier', 'Tools', 't-SNE Maps', 'Genome Browser', 'ChIP data', 'Change Data Scope', 'User Options', 'Help', and 'Contact / About R2'. Below the menu is a 'PubMed' logo and a 'Download the R2 Tutorials Book' button. The main content area is titled 'R2: ChIP' and contains a list of analysis tools: 'ChIP (Transcriptional Start Site)', 'TSS graphs within experiment', 'TSS graphs', 'TSS genesets scan', 'TSS XY plot 2 experiments', 'ChIPseq', 'ROSE Super Enhancer Plot', 'MACS Plot', 'RSEG Plot', 'ChIPseq TSS Peak/Coverage Plotter', 'ChIP Browser', 'ChIP-chip Genome Browser', and 'ChIP-chip Genome Browser (+GEx)'. Two arrows point from the 'ChIPseq TSS Peak/Coverage Plotter' and 'ChIP Browser' links to two signal plots. The first plot is for 'SY5YA H3K27ac (Valentijn)' and the second is for 'N691Cb H3K27ac (Valentijn)'. Both plots show signal intensity across a genomic region with a color scale from -2000 to 2000. The bottom of the interface includes 'AMC: Oncogenomics' and a 'Go to Main' button.

# TSS graphs





# ChIP Pincher



aration

# TSS geneset scans

R2: ChIP

hgserver1.amc.nl/cgi-bin/r2/main.cgi

Go to Main

- Main
- Time series
- AmpliconView

R2: ChIP

ChIP (Transcriptional Start Site)

- TSS graphs within experiment
- TSS graphs
- TSS genesets scan
- TSS XY plot 2 experiments

Online Tutorial

R2: ChIP\_chip

setname	stats	Kelly_ISL1 (Durbin)   GSM2486162-ET200   GSM2486166-ET200	Kelly_HAND2 (Durbin)   GSM2486166-ET200   GSM2486166-ET200
One_carbon_pool_by_folate	g:20 t:23	3.43 Footprint	5.12 Footprint
Mismatch_repair	g:23 t:26	3.15 Footprint	2.99 Footprint
Glycosaminoglycan_biosynthesis_heparan_sulfate_heparan	g:24 t:26	2.12 Footprint	3.08 Footprint
Tyrosine_metabolism	g:35 t:48	1.68 Footprint	4.76 Footprint
Phenylalanine_metabolism	g:17 t:24	1.61 Footprint	3.14 Footprint
Transcriptional_misregulation_in_cancer	g:179 t:248	1.37 Footprint	2.47 Footprint
Pantothenate_and_CoA_biosynthesis	g:18 t:24	1.33 Footprint	3.43 Footprint
Biosynthesis_of_unsaturated_fatty_acids	g:23 t:29	1.20 Footprint	2.94 Footprint
Histidine_metabolism	g:23 t:29	1.16 Footprint	3.17 Footprint
Glycosphingolipid_biosynthesis_lacto_and_neolacto_series	g:26 t:34	1.14 Footprint	3.44 Footprint
ABC_transporters	g:44 t:56	1.10 Footprint	2.19 Footprint
Colorectal_cancer	g:62 t:84	1.07 Footprint	2.69 Footprint
Protein_export	g:23 t:24	1.04 Footprint	2.35 Footprint
Fatty_acid_elongation	g:25 t:30	1.02 Footprint	2.81 Footprint
Citrate_cycle_TCA_cycle	g:30 t:34	0.94 Footprint	3.92 Footprint
Other_glycan_degradation	g:18 t:21	0.94 Footprint	2.87 Footprint
Steroid_biosynthesis	g:20 t:25	0.89 Footprint	3.39 Footprint
Terpenoid_backbone_biosynthesis	g:22 t:23	0.87 Footprint	3.95 Footprint
SNARE_interactions_in_vesicular_transport	g:34 t:35	0.85 Footprint	1.73 Footprint
Glycosylphosphatidylinositol_GPI_anchor_biosynthesis	g:25 t:25	0.85 Footprint	2.81 Footprint
Glycosaminoglycan_biosynthesis_keratan_sulfate	g:15 t:18	0.84 Footprint	2.48 Footprint
Other_types_of_O_glycan_biosynthesis	g:31 t:40	0.84 Footprint	4.45 Footprint
Fanconi_anemia_pathway	g:51 t:56	0.84 Footprint	2.89 Footprint
Dorso_ventral_axis_formation	g:27 t:32	0.82 Footprint	1.77 Footprint
Ribosome	g:133 t:138	0.82 Footprint	2.15 Footprint
VEGF_signaling_pathway	g:61 t:77	0.81 Footprint	2.68 Footprint
Cocaine_addiction	g:49 t:82	0.80 Footprint	3.10 Footprint
Nicotinate_and_nicotinamide_metabolism	g:29 t:35	0.79 Footprint	2.78 Footprint
Arginine_biosynthesis	g:20 t:27	0.78 Footprint	2.02 Footprint
Homologous_recombination	g:29 t:32	0.78 Footprint	2.81 Footprint
Circadian_rhythm	g:31 t:39	0.78 Footprint	3.32 Footprint
Tryptophan_metabolism	g:40 t:47	0.77 Footprint	3.37 Footprint
Type_1_diabetes_mellitus	g:41 t:45	0.77 Footprint	2.40 Footprint
Glycosaminoglycan_biosynthesis_chondroitin_sulfate_dermatan_sulfate	g:20 t:24	0.76 Footprint	2.72 Footprint
RNA_polymerase	g:31 t:33	0.75 Footprint	2.42 Footprint
Amphetamine_addiction	g:67 t:91	0.75 Footprint	2.86 Footprint
N_Glycan_biosynthesis	g:49 t:62	0.75 Footprint	1.83 Footprint
DNA_replication	g:36 t:39	0.75 Footprint	2.34 Footprint
Pyrimidine_metabolism	g:104 t:121	0.73 Footprint	2.33 Footprint
Spliceosome	g:132 t:142	0.73 Footprint	2.45 Footprint
Gastric_acid_secretion	g:74 t:112	0.73 Footprint	1.78 Footprint
Amyotrophic_lateral_sclerosis_ALS	g:51 t:62	0.72 Footprint	1.87 Footprint
Renal_cell_carcinoma	g:66 t:85	0.71 Footprint	2.11 Footprint
Propanoate_metabolism	g:32 t:41	0.71 Footprint	2.44 Footprint
Endocrine_and_other_factor_regulated_calcium_reabsorption	g:47 t:70	0.71 Footprint	2.26 Footprint
Glyoxylate_and_dicarboxylate_metabolism	g:28 t:32	0.70 Footprint	2.84 Footprint
Sphingolipid_metabolism	g:47 t:56	0.70 Footprint	2.10 Footprint
Aminocycl_RNA_biosynthesis	g:44 t:45	0.69 Footprint	2.55 Footprint
Vibrio_cholerae_infection	g:51 t:60	0.69 Footprint	2.08 Footprint
Acute_myeloid_leukemia	g:57 t:77	0.69 Footprint	2.68 Footprint
Selenocompound_metabolism	g:17 t:18	0.69 Footprint	4.78 Footprint
Nucleotide_excision_repair	g:47 t:53	0.69 Footprint	1.92 Footprint
Glycosaminoglycan_degradation	g:19 t:24	0.69 Footprint	2.32 Footprint

ChIP-chip tss  
kelly\_Transcriptional\_misregulation\_in\_cancer  
g:180 t:248

ChIP-chip tss  
kelly\_Transcriptional\_misregulation\_in\_cancer  
g:180 t:248

ChIP-chip tss  
kelly\_Transcriptional\_misregulation\_in\_cancer  
g:180 t:248

TSS\_detail Plot  
kelly\_Transcriptional\_misregulation\_in\_cancer  
Kelly\_HAND2 (Durbin) | GSM2486166-ET200  
Kelly\_ISL1 (Durbin) | GSM2486162-ET200

Score

# ChIPseq genome browser

The screenshot displays the R2: ChIP genome browser interface. The main window shows a genomic track for hg19:chr2 with a zoom level of 20X. A table titled "Select a ChIP-seq analysis result:" is overlaid on the track, listing various ChIP-seq experiments. The table includes columns for selection, sample ID, group, ID, cell line, target, author, color, and update status. The table contains 20 rows of data.

<input type="checkbox"/>	sol	chip_group	id	chip_cellline	chip_target	chip_author	colors	upd
<input type="checkbox"/>								
<input type="checkbox"/>		GSE94822	gsm3128276-et200	BE2C	input	Durbin	#C1C1C1	c.c.
<input type="checkbox"/>		GSE94822	gsm3128275-et200	BE2C	H3K27ac	Durbin	#AA0000	c.c.
<input type="checkbox"/>		GSE94822	gsm2915911-et200	Kelly	TBX2	Durbin	#0000AA	c.c.
<input type="checkbox"/>		GSE94822	gsm2915910-et200	Kelly	PHOX2B	Durbin	#0000AA	c.c.
<input type="checkbox"/>		GSE94822	gsm2915909-et200	Kelly	MYCN	Durbin	#0000AA	c.c.
<input type="checkbox"/>		GSE101295	gsm2871003-et200	SJNB3	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2871002-et200	SJNB3	H3K27ac	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE101295	gsm2871001-et200	NB69-MWZ4975	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2871000-et200	NB69-MWZ4975	CTCF	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE101295	gsm2870999-et200	SKNAS-MWZ4975	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2870998-et200	SKNAS-MWZ4975	CTCF	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE101295	gsm2870997-et200	SYSYS-MWZ4975	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2870996-et200	SYSYS-MWZ4975	CTCF	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE101295	gsm2700779-et200	NB69	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2700778-et200	NB69	H3K27ac	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE101295	gsm2700777-et200	SKNAS	input	Zimmerman	#C1C1C1	c.c.
<input type="checkbox"/>		GSE101295	gsm2700776-et200	SKNAS	H3K27ac	Zimmerman	#AA0000	c.c.
<input type="checkbox"/>		GSE90683	gsm2664371-et200	CLB-GA	HAND2	Boeva	#0000AA	c.c.
<input type="checkbox"/>		GSE90683	gsm2664370-et200	CLB-GA	GATA3	Boeva	#0000AA	c.c.
<input type="checkbox"/>		GSE90683	gsm2664369-et200	CLB-GA	PHOX2B	Boeva	#0000AA	c.c.

Additional interface elements include a navigation menu on the left, a search bar for genes, and various genomic tracks on the right such as Conservation, CpG Islands, RefSeq, and ENCODE. The browser address bar shows the URL: hgserver1.amc.nl/cgi-bin/r2/main.cgi.

# Multiple ChIP profiles

Get DNA sequence (UCSC)

Load / Store Profile

**properties**

**ChIP**

Chip data: chip\_raw

Sample label size: 12

Sample label placement: center

Grid: on

Range: a to a

Height: 80

Scale to library size: 20000000

Slider: no

**Available Datasets**

nr. selected: 1

Select / Adapt ChIP-Experiments

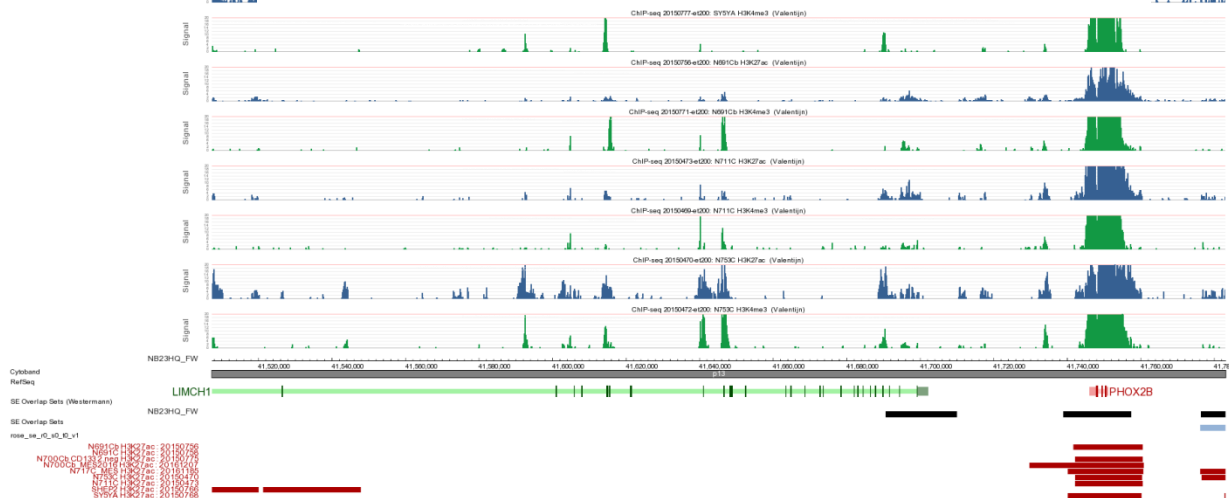
hg19: chr4

**Load/Delete a stored profile**

Profile: personal - Li\_coloredcms\_H3K27ac\_H3K4me1

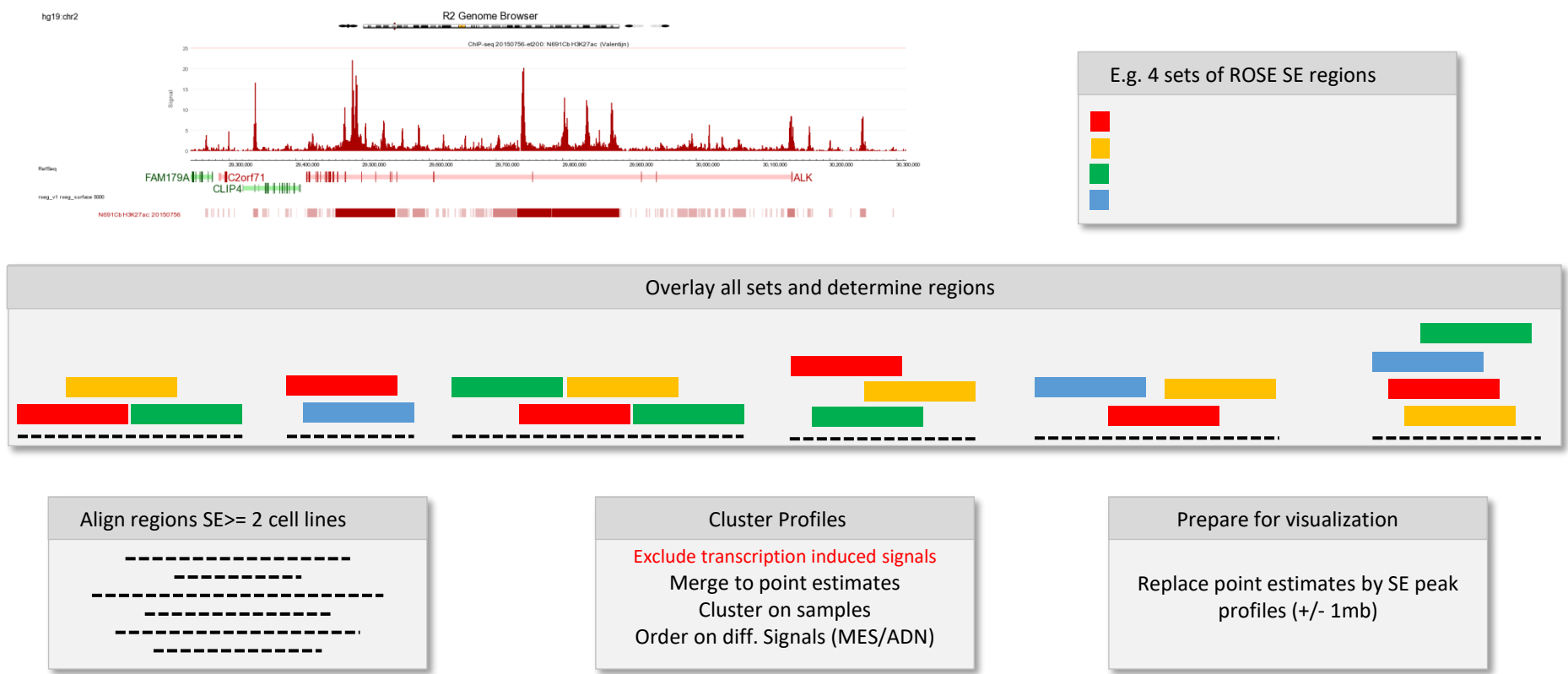
Operation: personal - Li\_coloredcms\_H3K27ac\_H3K4me1

- personal - Li\_normal\_tumor
- cemm - 220409\_CRL1\_Laura
- cemm - 220413\_CRL1\_rmdup\_Laura
- cemm - 220414\_CRL1\_mac3\_peaks
- cemm - 240215\_erna\_view\_for\_sinejan
- cemm - CRC\_cms\_meta\_profiles\_Li
- nb\_anc - 200708\_MVR\_SHEP2\_SY5Y\_001\_jk**
- nb\_anc - 200708\_MVR\_SHEP\_SY5Y\_001\_jk
- nb\_anc - 200721\_ARID1a\_fig4c1\_jk1
- nb\_anc - 201205\_6marks\_shep\_sy5y\_ses
- nb\_anc - 210525\_cut\_n\_run\_set\_001
- nb\_anc - 210622\_CLB-GA\_ALK\_SE\_HiChIP\_jk
- nb\_anc - 210729\_Ellen\_CLB\_SKNAS\_supp
- nb\_anc - chromhmm\_10\_profiles\_fcc\_v1
- nb\_anc - Ellen\_CLB\_SKNAS\_Suppl\_
- nb\_anc - Ellen\_TF\_adrn\_suppl\_
- nb\_anc - GSE128590\_HCT116\_crotonylation
- nb\_anc - GSE128590\_HCT116\_CR\_MVR\_example1
- nb\_anc - GSE147408\_NB\_RA\_jk1

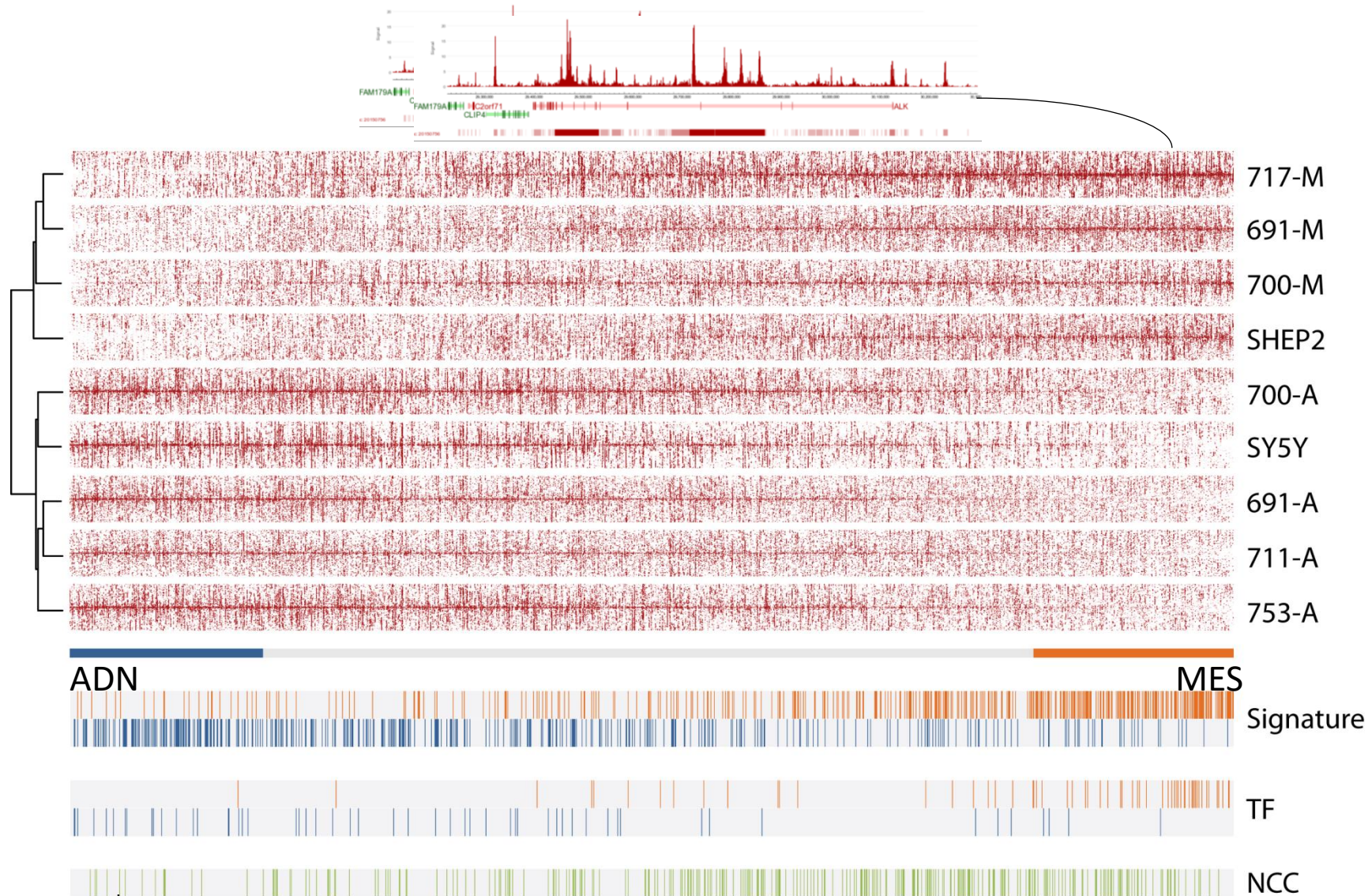


# Enhancer Landscape

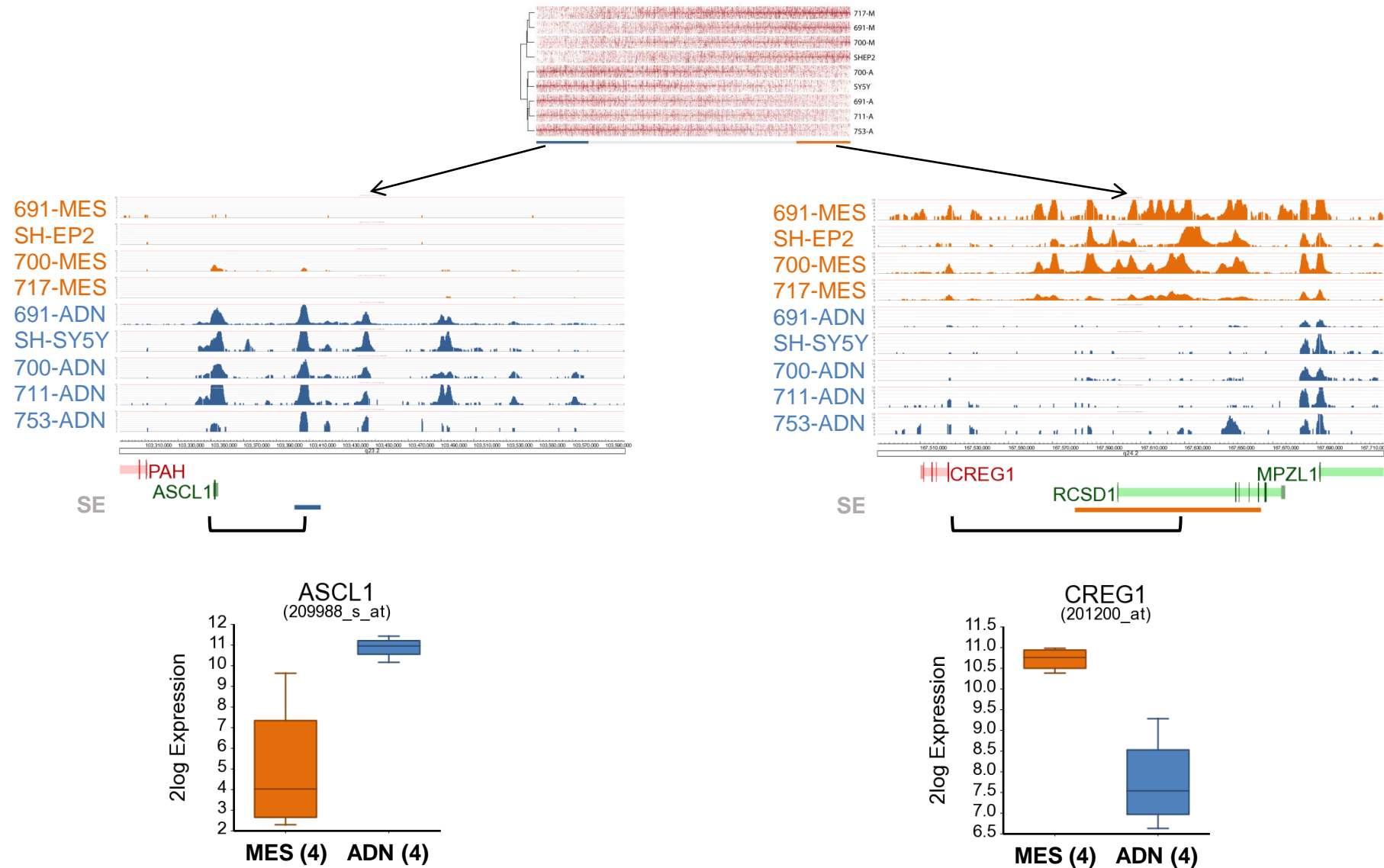
- ROSE SE data of multiple samples
  - Find overlapping SE's and assemble set of SE's seen more than once



# SuperEnhancer Landscape MES/ADN



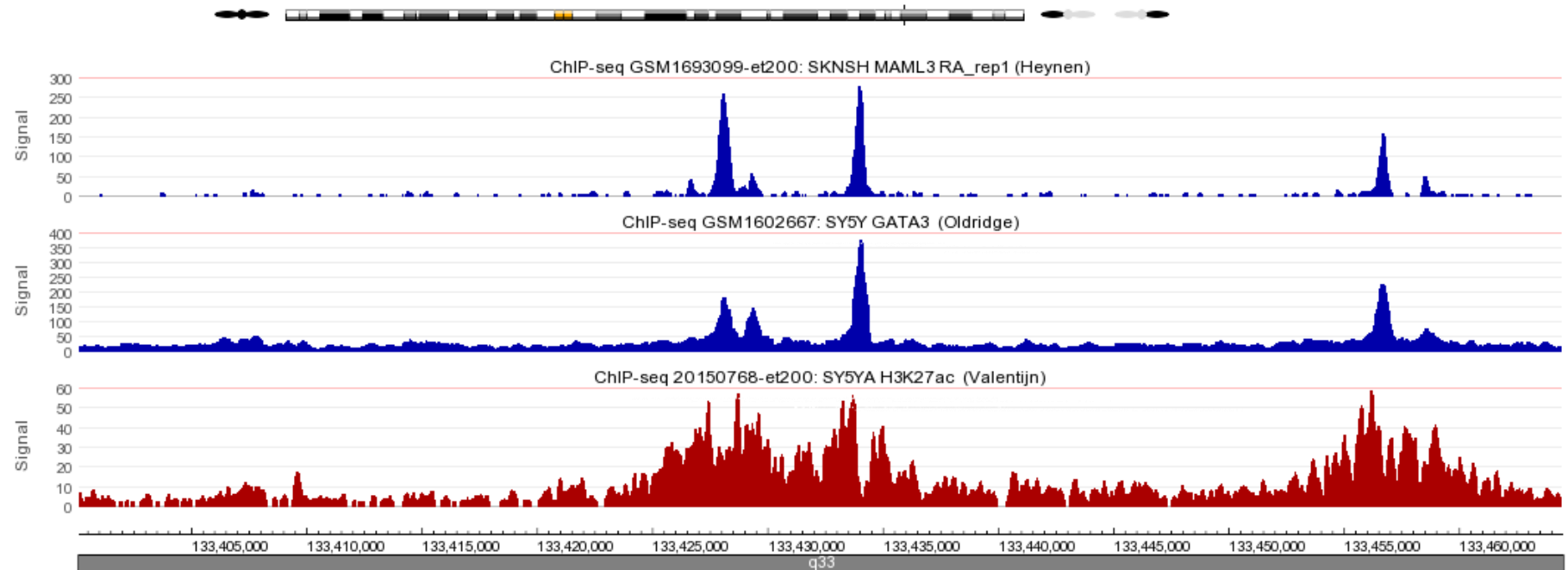
# SuperEnhancer Landscape MES/ADN



# ChIPseq analysis

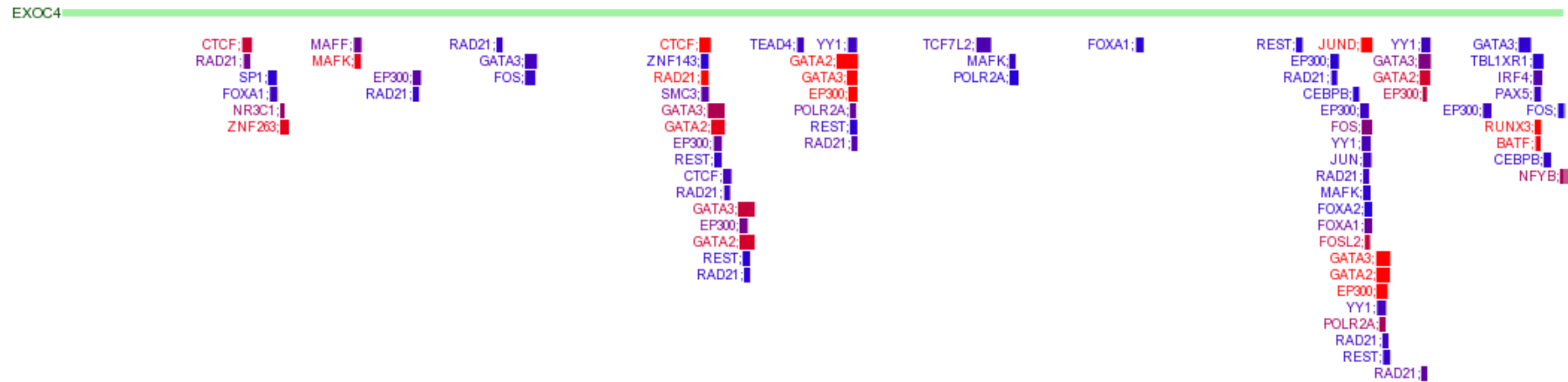
hg19:chr7

## R2 Genome Browser



Peaks for **161 transcription factors** in **91 cell types** are combined here into clusters to produce a summary display showing occupancy regions for each factor and motif sites within the regions when identified.

Scores were assigned to peaks by multiplying the input signal values by a normalization factor calculated as the ratio of the maximum score value (1000) to the signal value at one standard deviation from the mean. The cluster score is the highest score for any peak contributing to the cluster



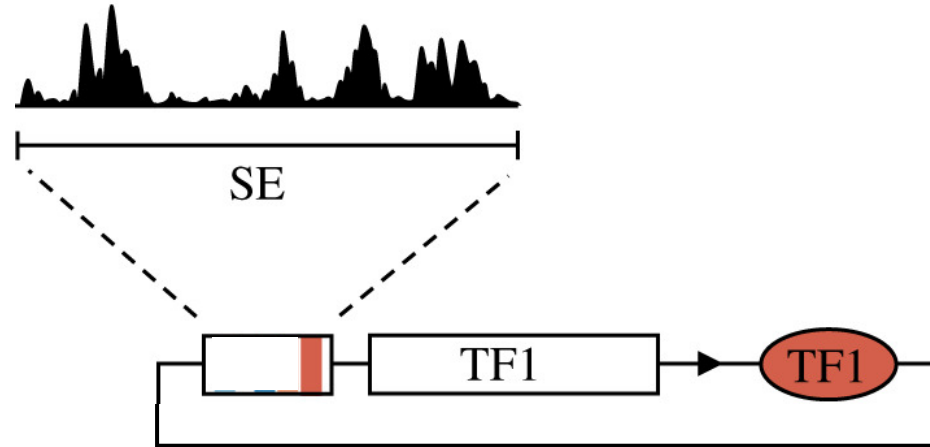
ENCODE re-analysis

<http://genome.ucsc.edu/cgi-bin/hgTrackUi?db=hg19&g=wgEncodeRegTfbsClusteredV3>

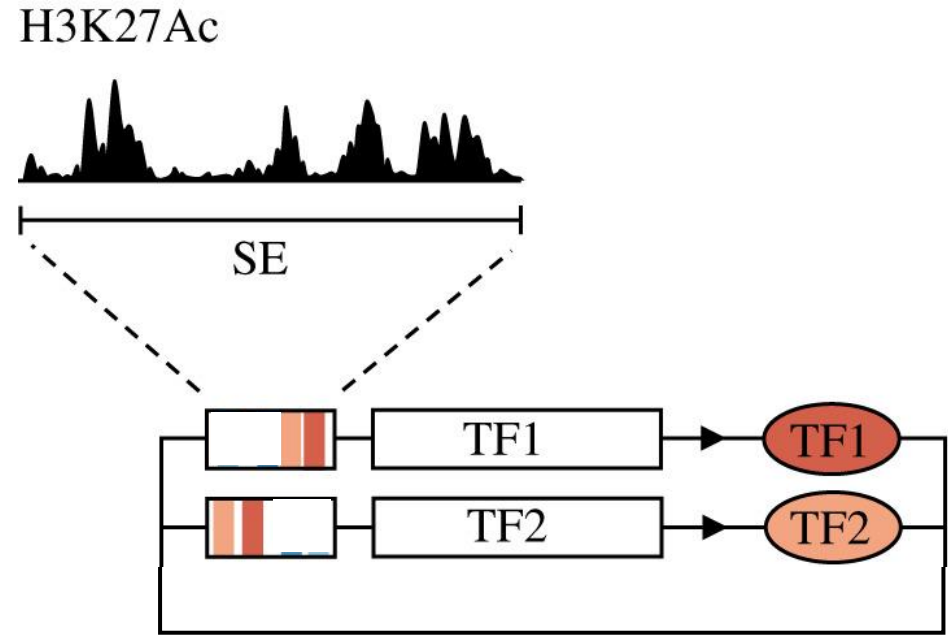


# Core Regulatory Circuitries (CRC)

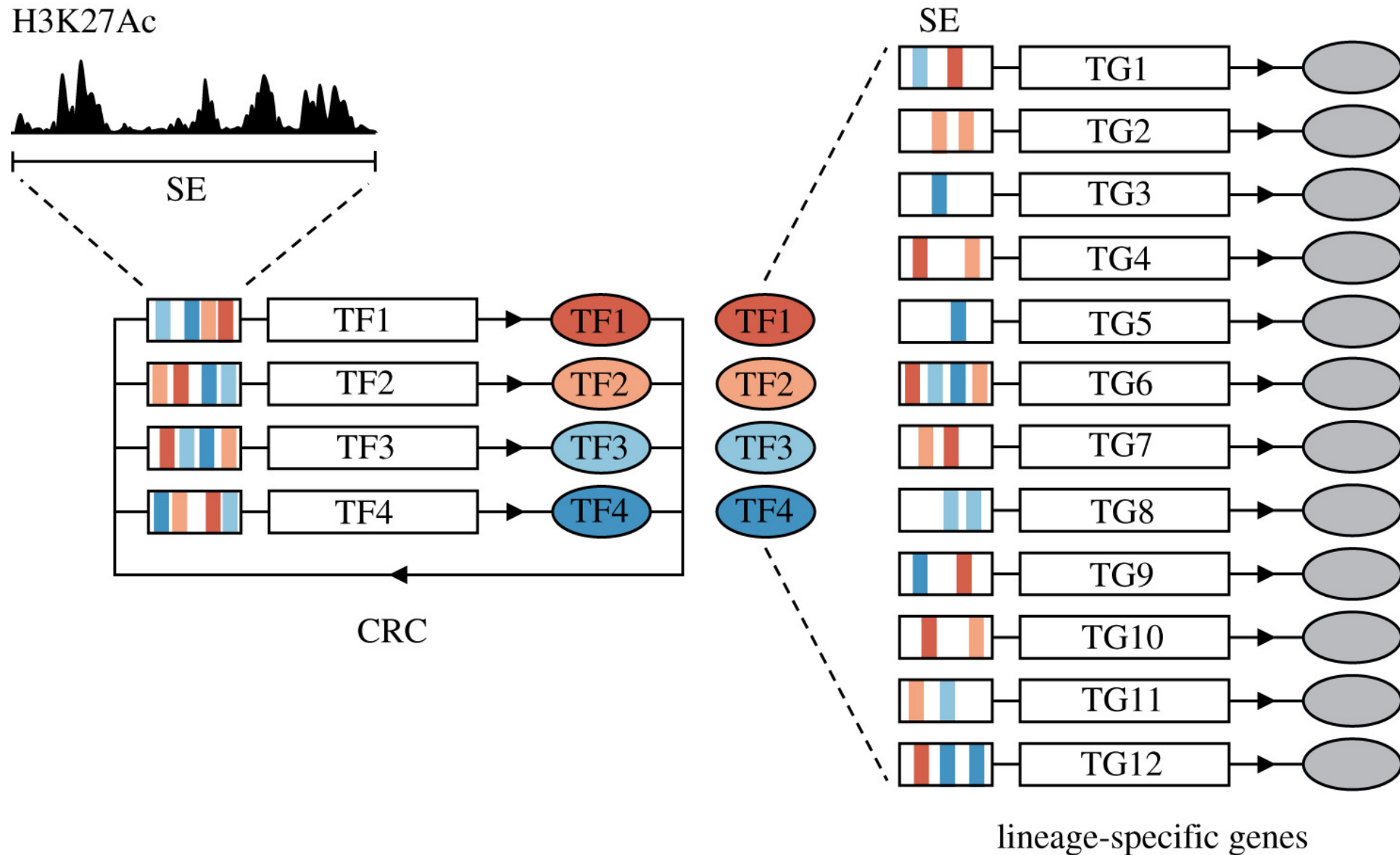
H3K27Ac



# Core Regulatory Circuitries (CRC)

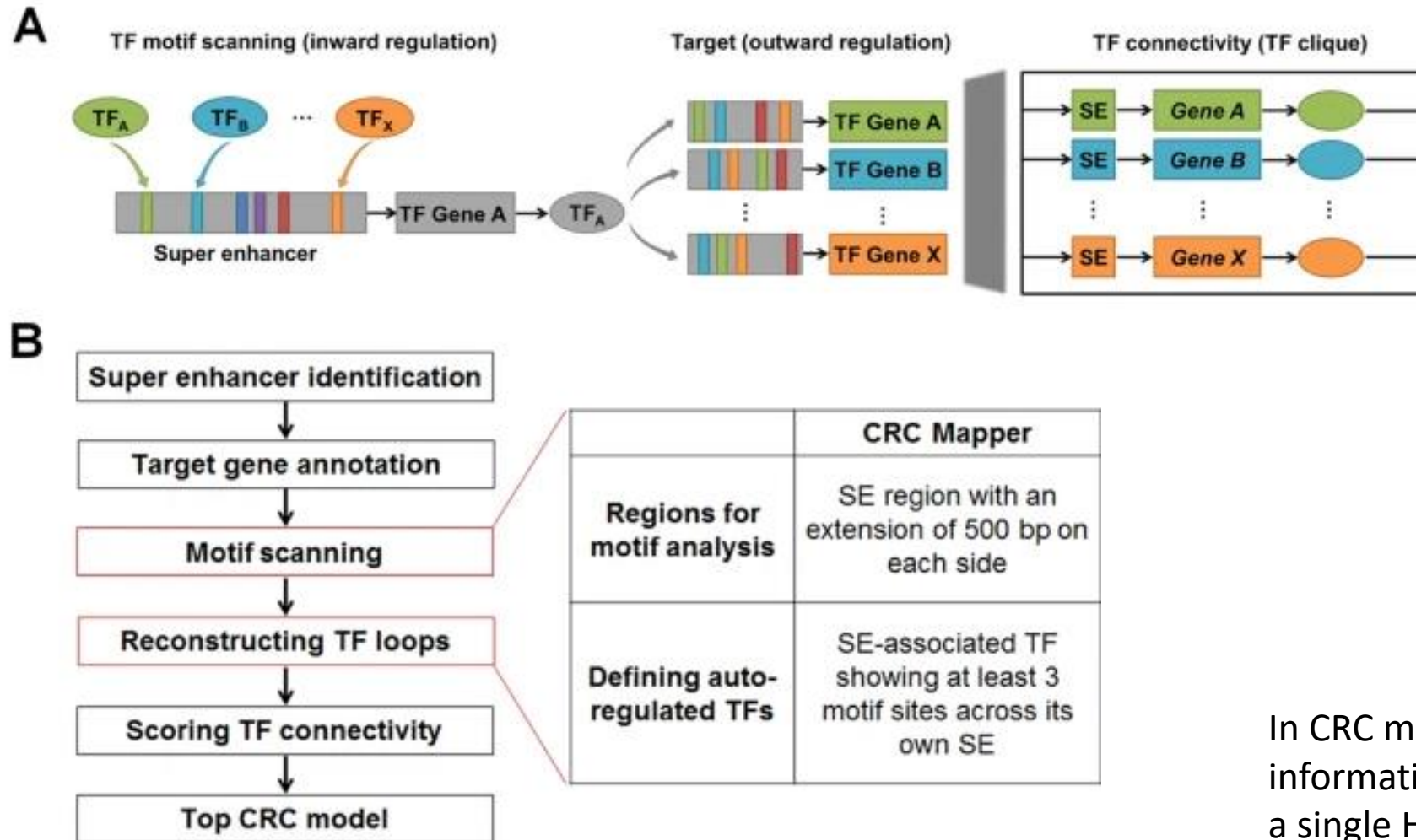


# Core Regulatory Circuitries (CRC)



# Identification of CRCs with CRC-Mapper

Fig. 2: Modeling of core transcriptional regulatory circuitry.



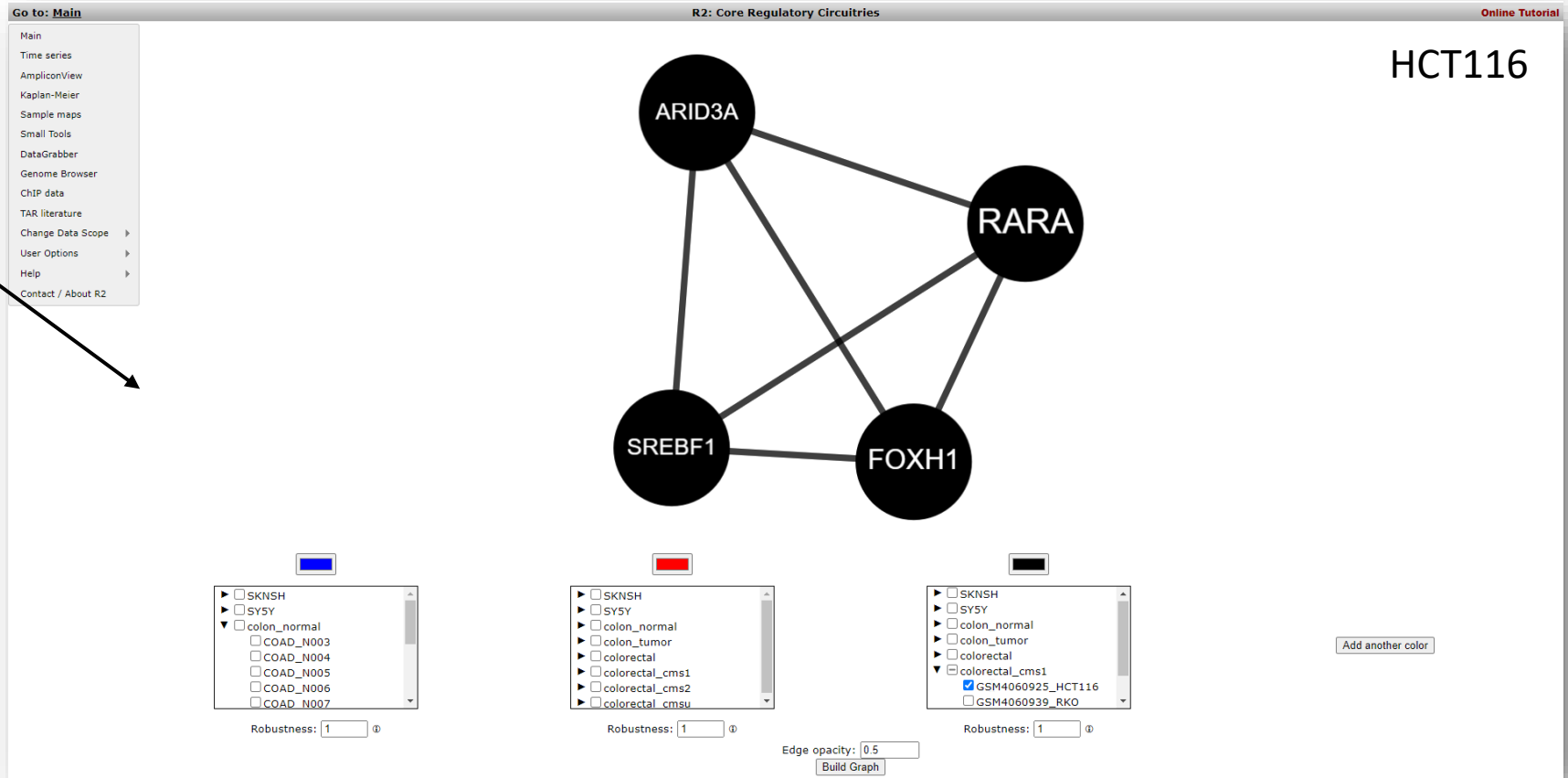
In CRC mapper all information is inferred from a single H3K27ac profile

# CRC graph in R2

- Main
- Time series
- AmpliconView
- Kaplan-Meier
- Sample maps
- Small Tools
- DataGrabber
- Genome Browser
- ChIP data**
- TAR literature
- Change Data Scope ▶
- User Options ▶
- Help ▶
- Contact / About R2

- ChIP (Transcriptional Start Site)
- TSS\_graphs within experiment
- TSS\_graphs
- TSS\_genesets scan
- TSS\_XY\_plot 2 experiments
- ChIPseq**
- ROSE Super Enhancer Plot
- MACS Plot
- RSEG Plot
- ChIPseq TSS Peak/Coverage Plotter
- ChIP Browser**
- ChIP Genome Browser
- ChIP Genome Browser + Gex
- Core Regulatory Circuitries**
- CRC\_graph

- Go to: Main
- Main
  - Time series
  - AmpliconView
  - Kaplan-Meier
  - Sample maps
  - Small Tools
  - DataGrabber
  - Genome Browser
  - ChIP data
  - TAR literature
  - Change Data Scope ▶
  - User Options ▶
  - Help ▶
  - Contact / About R2



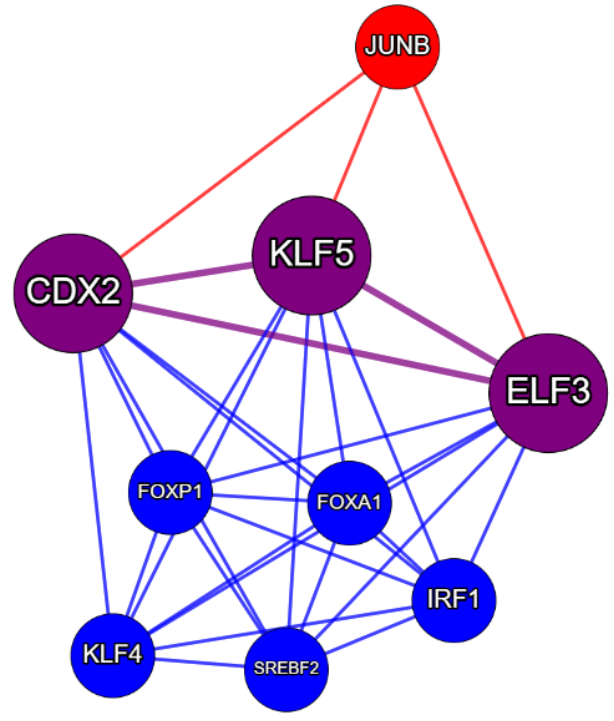
# CRC normal vs colon tumor

Go to: [Main](#) | [ITCC-P4 portal](#)

R2: Core Regulatory Circuitries

[Online Tutorial](#)

- Main
- ITCC-P4 (IMI2) ▶
- Cancer Portals
- Time series
- AmpliconView
- Kaplan-Meier
- Sample maps
- Small Tools
- DataGrabber
- Genome Browser
- ChIP data
- TAR literature
- Change Data Scope ▶
- User Options ▶
- Administrator ▶
- Help ▶
- Contact / About R2



ADRN  
 MES  
 SKNSH  
 SY5Y  
 colon\_normal  
 COAD\_N003  
 COAD\_N004  
 COAD\_N005

Robustness:  Ⓟ

ADRN  
 MES  
 SKNSH  
 SY5Y  
 colon\_normal  
 COAD\_T001  
 colon\_tumor  
 COAD\_T003

Robustness:  Ⓟ

ADRN  
 MES  
 SKNSH  
 SY5Y  
 colon\_normal  
 colon\_tumor  
 colorectal  
 colorectal cms1

Robustness:  Ⓟ

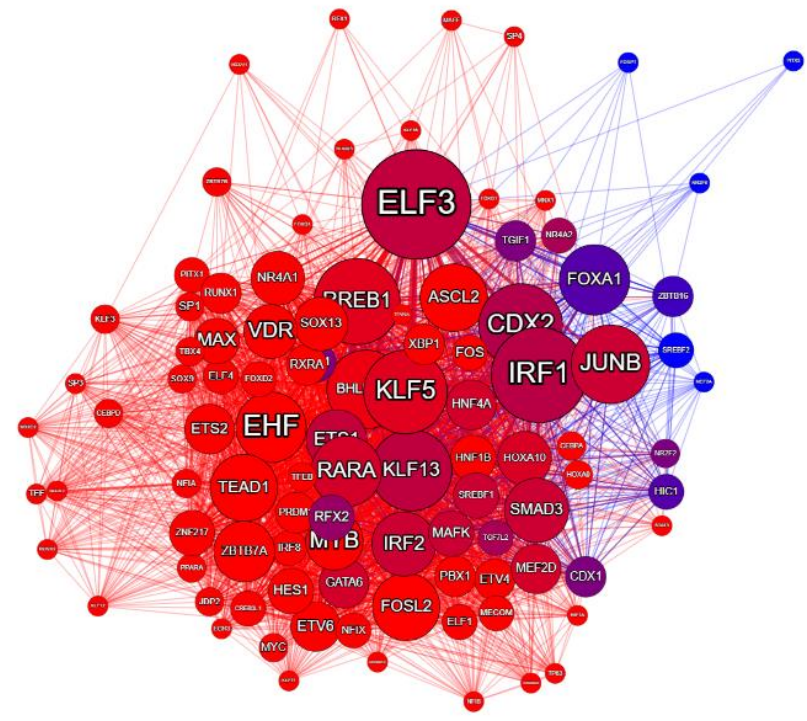
Edge opacity:

# CRC normal vs colon tumors

- Go to: **Main**
- Main
  - Time series
  - AmpliconView
  - Kaplan-Meier
  - Sample maps
  - Small Tools
  - DataGrabber
  - Genome Browser
  - ChIP data
  - TAR literature
  - Change Data Scope ▶
  - User Options ▶
  - Help ▶
  - Contact / About R2

R2: Core Regulatory Circuitries

Online Tutorial



SKNSH  
 SY5Y  
 colon\_normal  
 COAD\_N003  
 COAD\_N004  
 COAD\_N005  
 COAD\_N006  
 COAD\_N007

Robustness:  ⓘ

SKNSH  
 SY5Y  
 colon\_normal  
 colon\_tumor  
 colorectal  
 colorectal\_cms1  
 colorectal\_cms2  
 colorectal\_cmsu

Robustness:  ⓘ

SKNSH  
 SY5Y  
 colon\_normal  
 colon\_tumor  
 colorectal  
 colorectal\_cms1  
 GSM4060925\_HCT116  
 GSM4060939\_RKO

Robustness:  ⓘ

Add another color

Edge opacity:

# Fun things to do:

## Tutorial

- Go to a chapter of interest and follow the examples
- Toy around with a similar pipeline on a different dataset
- Perform a follow up analysis from the tutorial

## R2 Butterfly Training Courses

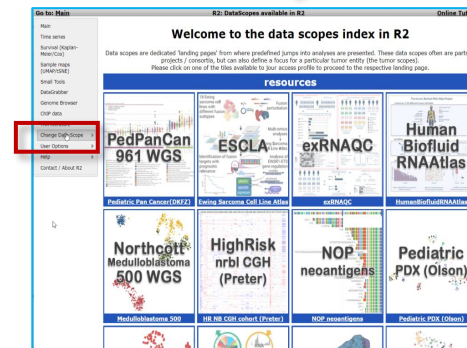
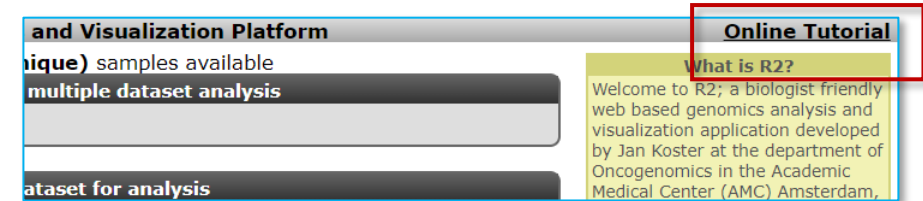
- Help => Training Courses => Butterfly Training Courses
- Read carefully and follow the research line step by step
- Feel free to toy around
  - Who knows

## Do your own research

- Find a dataset of your interest
- Think about a good biological research question
- Try to find the analyses that can help you gain insight

## Datascopes

- Find an interesting datascope to dive into





# Ither datascope

The screenshot displays the iTher datascope interface. At the top, it reads "R2: iTher (PMC)" and "R2: Genomics analysis and visualization platform". A navigation menu on the left includes options like "Main", "Time series", "Survival (Kaplan-Meier/Cox)", "Sample maps (UMAP/TSNE)", "Small Tools", "DataGrabber", "Genome Browser", "ChIP data", "TAR literature", "Change Data Scope", "User Options", "Help", and "Contact / About R2". A sub-menu for "resources" is open, listing various datasets and tools such as "Pediatric Pan Cancer (DKFZ)", "Ewing Sarcoma Cell Line Atlas", "exRNAQC", "HumanBiofluidRNAAtlas", "Medulloblastoma 500", "HR NB CGH cohort (Preter)", "NOP neantigens", "Pediatric PDX (Olson)", "PPTC PDX", "RNA Atlas", "Halbritter 2024 NC", "iTher (PMC)", and "PMC NRRL drug screens 2023".

The main content area is titled "Complete iTher cohort with provided events" and features a "Fixed Somatic Summary" plot. Below this, there are two sections of analysis tool tiles:

- iTher Tiles on PMC platform v2 (HG38 Gencode31):** This section includes tiles for "circos archive", "somatic variants", "genome browser", "Somatic Summary", "zscorer", "Mutation Burden", "find sample by omics", "cohort overview", "Mut vs Track", "CGH profile sorter", and "Gex UMAP".
- iTher Tiles on INFORM platform (HG19 Gencode19):** This section includes tiles for "circos archive", "somatic variants", "genome browser", "Somatic Summary", "zscorer", "Mutation Burden", and "find sample by omics".

Dr. Bianca Goemans and Dr. Karin Langenberg, pediatric oncologists and researchers, are involved in the iTher (individualized THERapy) study that has been ongoing for the past five years. Bianca Goemans says: 'This study involved children with a cancer diagnosis that is difficult to treat, so for which there is no standard treatment available. We wanted to map the DNA and RNA of cancer cells. If you know exactly what that looks like, you can find leads for a targeted medicine. We call this precision medicine.'

## Results

This study has shown that these DNA and RNA analyses are possible and that in the majority of patients (83%) changes were shown in their cancer cells that could potentially be used as a target for treatment. Bianca Goemans: 'Unfortunately, few patients were ultimately treated with such a targeted medicine, partly because the medicine was often not (yet) available for children or because children were already too sick to participate in a clinical trial. That's why we want to do these analyses earlier in the course of the disease so that children may still qualify for a targeted treatment.'

# Fun things to do:

## Tutorial

- Go to a chapter of interest and follow the examples
- Toy around with a similar pipeline on a different dataset
- Perform a follow up analysis from the tutorial

## R2 Butterfly Training Courses

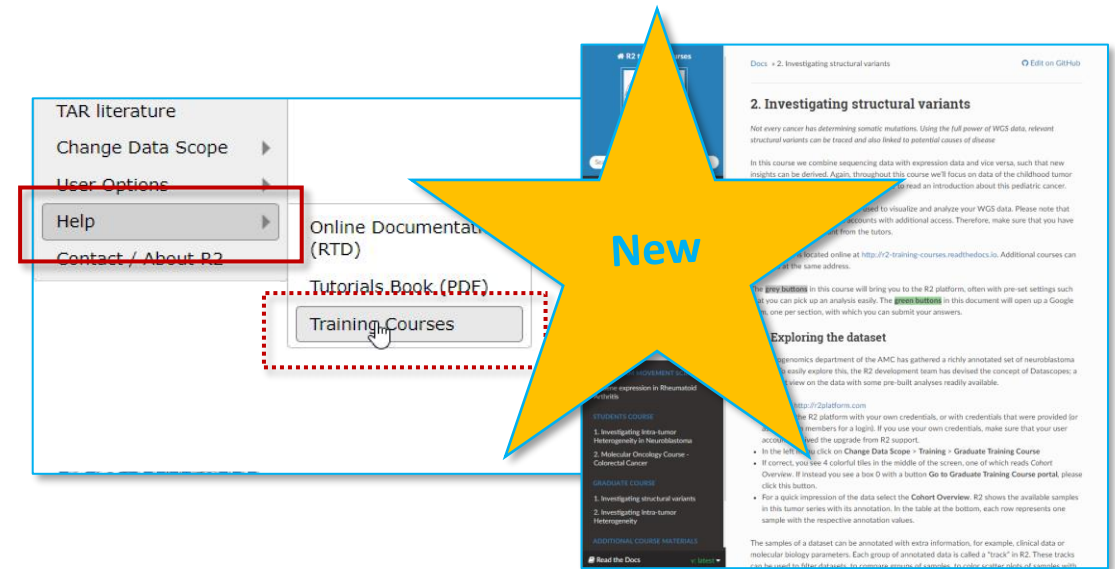
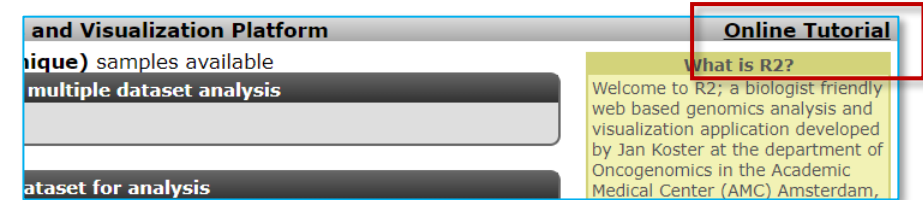
- Help => Training Courses => Butterfly Training Courses
- Read carefully and follow the research line step by step
- Feel free to toy around
  - Who knows

## Do your own research

- Find a dataset of your interest
- Think about a good biological research question
- Try to find the analyses that can help you gain insight

## Datascopes

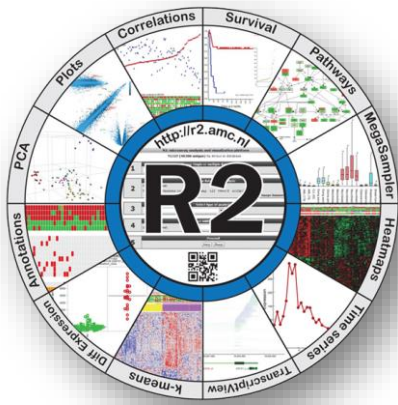
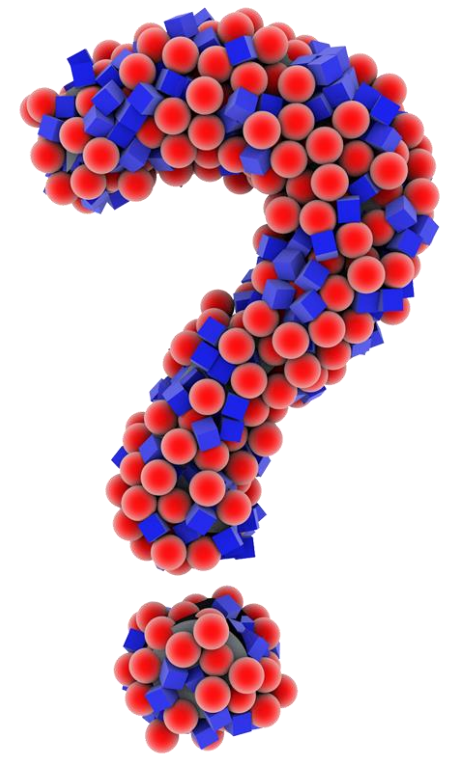
- Find an interesting datascope to dive into



**R2 Platform Team**

- Rianne van Drimmelen
- Romeo Willinge Prins
- Danny Zwijnenburg
- Richard Volckmann
- Christian Griffioen
- Lieke Hoyng
- Jan Koster

Koster J(151) Johnsen JI(18) Zhang F(6) Li M(14) Yang W(7) Li Z(32) Faber AC(7) Stallings RL(14) Capper D(9) Li Y(36)  
 Zhao L(10) Xu T(5) Vassal G(1) Li G(10) Garaventa A(11) Jager N(11) Gong Q(8) Zhang J(26) Westerhout EM(11) Buntch S(6)  
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